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(54) Title: COMPOSITIONS AND METHODS FOR THE DIAGNOSIS AND TREATMENT OF TUMOR

(57) Abstract: The present invention is directed to compositions of matter useful for the diagnosis and treatment of tumor in mammals and to methods of using those compositions of matter for the same.

COMPOSITIONS AND METHODS FOR THE DIAGNOSIS AND TREATMENT OF TUMOR

FIELD OF THE INVENTION

The present invention is directed to compositions of matter useful for the diagnosis and treatment of tumor in mammals and to methods of using those compositions of matter for the same.

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BACKGROUND OF THE INVENTION

Malignant tumors (cancers) are the second leading cause of death in the United States, after heart disease (Boring et al., CA Cancel J. Clin. 43:7 (1993)). Cancer is characterized by the increase in the number of abnormal, or neoplastic, cells derived from a normal tissue which proliferate to form a tumor mass, the invasion of adjacent tissues by these neoplastic tumor cells, and the generation of malignant cells which eventually spread via the blood or lymphatic system to regional lymph nodes and to distant sites via a process called metastasis. In a cancerous state, a cell proliferates under conditions in which normal cells would not grow. Cancer manifests itself in a wide variety of forms, characterized by different degrees of invasiveness and aggressiveness.

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In attempts to discover effective cellular targets for cancer diagnosis and therapy, researchers have sought to identify transmembrane or otherwise membrane-associated polypeptides that are specifically expressed on the surface of one or more particular type(s) of cancer cell as compared to on one or more normal non-cancerous cell(s). Often, such membrane-associated polypeptides are more abundantly expressed on the surface of the cancer cells as compared to on the surface of the non-cancerous cells. The identification of such tumor-associated cell surface antigen polypeptides has given rise to the ability to specifically target cancer cells for destruction via antibody-based therapies. In this regard, it is noted that antibody-based therapy has proved very effective in the treatment of certain cancers. For example, HERCEPTIN® and RITUXAN® (both from Genentech Inc., South San Francisco, California) are antibodies that have been used successfully to treat breast cancer and non-Hodgkin's lymphoma, respectively. More specifically, HERCEPTIN® is a recombinant DNA-derived humanized monoclonal antibody that selectively binds to the extracellular domain of the human epidermal growth factor receptor 2 (HER2) proto-oncogene. HER2 protein overexpression is observed in 25-30% of primary breast cancers. RITUXAN® is a genetically engineered chimeric murine/human monoclonal antibody directed against the CD20 antigen found on the surface of normal and malignant B lymphocytes. Both these antibodies are recombinantly produced in CHO cells.

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In other attempts to discover effective cellular targets for cancer diagnosis and therapy, researchers have sought to identify (1) non-membrane-associated polypeptides that are specifically produced by one or more particular type(s) of cancer cell(s) as compared to by one or more particular type(s) of non-cancerous normal cell(s), (2) polypeptides that are produced by cancer cells at an expression level that is significantly higher than that of one or more normal non-cancerous cell(s), or (3) polypeptides whose expression is specifically limited

to only a single (or very limited number of different) tissue type(s) in both the cancerous and non-cancerous state (e.g., normal prostate and prostate tumor tissue). Such polypeptides may remain intracellularly located or may be secreted by the cancer cell. Moreover, such polypeptides may be expressed not by the cancer cell itself, but rather by cells which produce and/or secrete polypeptides having a potentiating or growth-enhancing effect on cancer cells. Such secreted polypeptides are often proteins that provide cancer cells with a growth advantage over normal cells and include such things as, for example, angiogenic factors, cellular adhesion factors, growth factors, and the like. Identification of antagonists of such non-membrane associated polypeptides would be expected to serve as effective therapeutic agents for the treatment of such cancers. Furthermore, identification of the expression pattern of such polypeptides would be useful for the diagnosis of particular cancers in mammals.

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Despite the above identified advances in mammalian cancer therapy, there is a great need for additional diagnostic and therapeutic agents capable of detecting the presence of tumor in a mammal and for effectively inhibiting neoplastic cell growth, respectively. Accordingly, it is an objective of the present invention to identify: (1) cell membrane-associated polypeptides that are more abundantly expressed on one or more type(s) of cancer cell(s) as compared to on normal cells or on other different cancer cells, (2) non-membrane-associated polypeptides that are specifically produced by one or more particular type(s) of cancer cell(s) (or by other cells that produce polypeptides having a potentiating effect on the growth of cancer cells) as compared to by one or more particular type(s) of non-cancerous normal cell(s), (3) non-membrane-associated polypeptides that are produced by cancer cells at an expression level that is significantly higher than that of one or more normal noncancerous cell(s), or (4) polypeptides whose expression is specifically limited to only a single (or very limited number of different) tissue type(s) in both a cancerous and non-cancerous state (e.g., normal prostate and prostate tumor tissue), and to use those polypeptides, and their encoding nucleic acids, to produce compositions of matter useful in the therapeutic treatment and diagnostic detection of cancer in mammals. It is also an objective of the present invention to identify cell membrane-associated, secreted or intracellular polypeptides whose expression is limited to a single or very limited number of tissues, and to use those polypeptides, and their encoding nucleic acids, to produce compositions of matter useful in the therapeutic treatment and diagnostic detection of cancer in mammals.

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SUMMARY OF THE INVENTION

A. Embodiments

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In the present specification, Applicants describe for the first time the identification of various cellular polypeptides (and their encoding nucleic acids or fragments thereof) which are expressed to a greater degree on the surface of or by one or more types of cancer cell(s) as compared to on the surface of or by one or more types of normal non-cancer cells. Alternatively, such polypeptides are expressed by cells which produce and/or secrete polypeptides having a potentiating or growth-enhancing effect on cancer cells. Again alternatively, such polypeptides may not be overexpressed by tumor cells as compared to normal cells of the same tissue type, but rather may be specifically expressed by both tumor cells and normal cells of only a single or very limited

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number of tissue types (preferably tissues which are not essential for life, e.g., prostate, etc.). All of the above polypeptides are herein referred to as <u>Tumor-associated Antigenic Target polypeptides</u> ("TAT" polypeptides) and are expected to serve as effective targets for cancer therapy and diagnosis in mammals.

Accordingly, in one embodiment of the present invention, the invention provides an isolated nucleic acid molecule having a nucleotide sequence that encodes a tumor-associated antigenic target polypeptide or fragment thereof (a "TAT" polypeptide).

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In certain aspects, the isolated nucleic acid molecule comprises a nucleotide sequence having at least about 80% nucleic acid sequence identity, alternatively at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% nucleic acid sequence identity, to (a) a DNA molecule encoding a full-length TAT polypeptide having an amino acid sequence as disclosed herein, a TAT polypeptide amino acid sequence lacking the signal peptide as disclosed herein, an extracellular domain of a transmembrane TAT polypeptide, with or without the signal peptide, as disclosed herein or any other specifically defined fragment of a full-length TAT polypeptide amino acid sequence as disclosed herein, or (b) the complement of the DNA molecule of (a).

In other aspects, the isolated nucleic acid molecule comprises a nucleotide sequence having at least about 80% nucleic acid sequence identity, alternatively at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% nucleic acid sequence identity, to (a) a DNA molecule comprising the coding sequence of a full-length TAT polypeptide cDNA as disclosed herein, the coding sequence of a TAT polypeptide lacking the signal peptide as disclosed herein, the coding sequence of an extracellular domain of a transmembrane TAT polypeptide, with or without the signal peptide, as disclosed herein or the coding sequence of any other specifically defined fragment of the full-length TAT polypeptide amino acid sequence as disclosed herein, or (b) the complement of the DNA molecule of (a).

In further aspects, the invention concerns an isolated nucleic acid molecule comprising a nucleotide sequence having at least about 80% nucleic acid sequence identity, alternatively at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% nucleic acid sequence identity, to (a) a DNA molecule that encodes the same mature polypeptide encoded by the full-length coding region of any of the human protein cDNAs deposited with the ATCC as disclosed herein, or (b) the complement of the DNA molecule of (a).

Another aspect of the invention provides an isolated nucleic acid molecule comprising a nucleotide sequence encoding a TAT polypeptide which is either transmembrane domain-deleted or transmembrane domain-inactivated, or is complementary to such encoding nucleotide sequence, wherein the transmembrane domain(s) of such polypeptide(s) are disclosed herein. Therefore, soluble extracellular domains of the herein described TAT polypeptides are contemplated.

In other aspects, the present invention is directed to isolated nucleic acid molecules which hybridize to (a) a nucleotide sequence encoding a TAT polypeptide having a full-length amino acid sequence as disclosed herein, a TAT polypeptide amino acid sequence lacking the signal peptide as disclosed herein, an extracellular

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domain of a transmembrane TAT polypeptide, with or without the signal peptide, as disclosed herein or any other specifically defined fragment of a full-length TAT polypeptide amino acid sequence as disclosed herein, or (b) the complement of the nucleotide sequence of (a). In this regard, an embodiment of the present invention is directed to fragments of a full-length TAT polypeptide coding sequence, or the complement thereof, as disclosed herein, that may find use as, for example, hybridization probes useful as, for example, diagnostic probes, antisense oligonucleotide probes, or for encoding fragments of a full-length TAT polypeptide that may optionally encode a polypeptide comprising a binding site for an anti-TAT polypeptide antibody, a TAT binding oligopeptide or other small organic molecule that binds to a TAT polypeptide. Such nucleic acid fragments are usually at least about 5 nucleotides in length, alternatively at least about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600, 610, 620, 630, 640, 650, 660, 670, 680, 690, 700, 710, 720, 730, 740, 750, 760, 770, 780, 790, 800, 810, 820, 830, 840, 850, 860, 870, 880, 890, 900, 910, 920, 930, 940, 950, 960, 970, 980, 990, or 1000 nucleotides in length, wherein in this context the term "about" means the referenced nucleotide sequence length plus or minus 10% of that referenced length. It is noted that novel fragments of a TAT polypeptide-encoding nucleotide sequence may be determined in a routine manner by aligning the TAT polypeptide-encoding nucleotide sequence with other known nucleotide sequences using any of a number of well known sequence alignment programs and determining which TAT polypeptide-encoding nucleotide sequence fragment(s) are novel. All of such novel fragments of TAT polypeptide-encoding nucleotide sequences are contemplated herein. Also contemplated are the TAT polypeptide fragments encoded by these nucleotide molecule fragments, preferably those TAT polypeptide fragments that comprise a binding site for an anti-TAT antibody, a TAT binding oligopeptide or other small organic molecule that binds to a TAT polypeptide.

In another embodiment, the invention provides isolated TAT polypeptides encoded by any of the isolated nucleic acid sequences hereinabove identified.

In a certain aspect, the invention concerns an isolated TAT polypeptide, comprising an amino acid sequence having at least about 80% amino acid sequence identity, alternatively at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% amino acid sequence identity, to a TAT polypeptide having a full-length amino acid sequence as disclosed herein, a TAT polypeptide amino acid sequence lacking the signal peptide as disclosed herein, an extracellular domain of a transmembrane TAT polypeptide protein, with or without the signal peptide, as disclosed herein, an amino acid sequence encoded by any of the nucleic acid sequences disclosed herein or any other specifically defined fragment of a full-length TAT polypeptide amino acid sequence as disclosed herein.

In a further aspect, the invention concerns an isolated TAT polypeptide comprising an amino acid sequence having at least about 80% amino acid sequence identity, alternatively at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% amino acid

sequence identity, to an amino acid sequence encoded by any of the human protein cDNAs deposited with the ATCC as disclosed herein.

In a specific aspect, the invention provides an isolated TAT polypeptide without the N-terminal signal sequence and/or without the initiating methionine and is encoded by a nucleotide sequence that encodes such an amino acid sequence as hereinbefore described. Processes for producing the same are also herein described, wherein those processes comprise culturing a host cell comprising a vector which comprises the appropriate encoding nucleic acid molecule under conditions suitable for expression of the TAT polypeptide and recovering the TAT polypeptide from the cell culture.

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Another aspect of the invention provides an isolated TAT polypeptide which is either transmembrane domain-deleted or transmembrane domain-inactivated. Processes for producing the same are also herein described, wherein those processes comprise culturing a host cell comprising a vector which comprises the appropriate encoding nucleic acid molecule under conditions suitable for expression of the TAT polypeptide and recovering the TAT polypeptide from the cell culture.

In other embodiments of the present invention, the invention provides vectors comprising DNA encoding any of the herein described polypeptides. Host cells comprising any such vector are also provided. By way of example, the host cells may be CHO cells, *E. coli* cells, or yeast cells. A process for producing any of the herein described polypeptides is further provided and comprises culturing host cells under conditions suitable for expression of the desired polypeptide and recovering the desired polypeptide from the cell culture.

In other embodiments, the invention provides isolated chimeric polypeptides comprising any of the herein described TAT polypeptides fused to a heterologous (non-TAT) polypeptide. Example of such chimeric molecules comprise any of the herein described TAT polypeptides fused to a heterologous polypeptide such as, for example, an epitope tag sequence or a Fc region of an immunoglobulin.

In another embodiment, the invention provides an antibody which binds, preferably specifically, to any of the above or below described polypeptides. Optionally, the antibody is a monoclonal antibody, antibody fragment, chimeric antibody, humanized antibody, single-chain antibody or antibody that competitively inhibits the binding of an anti-TAT polypeptide antibody to its respective antigenic epitope. Antibodies of the present invention may optionally be conjugated to a growth inhibitory agent or cytotoxic agent such as a toxin, including, for example, a maytansinoid or calicheamicin, an antibiotic, a radioactive isotope, a nucleolytic enzyme, or the like. The antibodies of the present invention may optionally be produced in CHO cells or bacterial cells and preferably induce death of a cell to which they bind. For diagnostic purposes, the antibodies of the present invention may be detectably labeled, attached to a solid support, or the like.

In other embodiments of the present invention, the invention provides vectors comprising DNA encoding any of the herein described antibodies. Host cell comprising any such vector are also provided. By way of example, the host cells may be CHO cells, *E. coli* cells, or yeast cells. A process for producing any of the herein described antibodies is further provided and comprises culturing host cells under conditions suitable for expression of the desired antibody and recovering the desired antibody from the cell culture.

In another embodiment, the invention provides oligopeptides ("TAT binding oligopeptides") which

bind, preferably specifically, to any of the above or below described TAT polypeptides. Optionally, the TAT binding oligopeptides of the present invention may be conjugated to a growth inhibitory agent or cytotoxic agent such as a toxin, including, for example, a maytansinoid or calicheamicin, an antibiotic, a radioactive isotope, a nucleolytic enzyme, or the like. The TAT binding oligopeptides of the present invention may optionally be produced in CHO cells or bacterial cells and preferably induce death of a cell to which they bind. For diagnostic purposes, the TAT binding oligopeptides of the present invention may be detectably labeled, attached to a solid support, or the like.

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In other embodiments of the present invention, the invention provides vectors comprising DNA encoding any of the herein described TAT binding oligopeptides. Host cell comprising any such vector are also provided. By way of example, the host cells may be CHO cells, *E. coli* cells, or yeast cells. A process for producing any of the herein described TAT binding oligopeptides is further provided and comprises culturing host cells under conditions suitable for expression of the desired oligopeptide and recovering the desired oligopeptide from the cell culture.

In another embodiment, the invention provides small organic molecules ("TAT binding organic molecules") which bind, preferably specifically, to any of the above or below described TAT polypeptides. Optionally, the TAT binding organic molecules of the present invention may be conjugated to a growth inhibitory agent or cytotoxic agent such as a toxin, including, for example, a maytansinoid or calicheamicin, an antibiotic, a radioactive isotope, a nucleolytic enzyme, or the like. The TAT binding organic molecules of the present invention preferably induce death of a cell to which they bind. For diagnostic purposes, the TAT binding organic molecules of the present invention may be detectably labeled, attached to a solid support, or the like.

In a still further embodiment, the invention concerns a composition of matter comprising a TAT polypeptide as described herein, a chimeric TAT polypeptide as described herein, an anti-TAT antibody as described herein, a TAT binding oligopeptide as described herein, or a TAT binding organic molecule as described herein, in combination with a carrier. Optionally, the carrier is a pharmaceutically acceptable carrier.

In yet another embodiment, the invention concerns an article of manufacture comprising a container and a composition of matter contained within the container, wherein the composition of matter may comprise a TAT polypeptide as described herein, a chimeric TAT polypeptide as described herein, an anti-TAT antibody as described herein, a TAT binding oligopeptide as described herein, or a TAT binding organic molecule as described herein. The article may further optionally comprise a label affixed to the container, or a package insert included with the container, that refers to the use of the composition of matter for the therapeutic treatment or diagnostic detection of a tumor.

Another embodiment of the present invention is directed to the use of a TAT polypeptide as described herein, a chimeric TAT polypeptide as described herein, an anti-TAT polypeptide antibody as described herein, a TAT binding oligopeptide as described herein, or a TAT binding organic molecule as described herein, for the preparation of a medicament useful in the treatment of a condition which is responsive to the TAT polypeptide, chimeric TAT polypeptide, anti-TAT polypeptide antibody, TAT binding oligopeptide, or TAT

binding organic molecule.

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B. Additional Embodiments

Another embodiment of the present invention is directed to a method for inhibiting the growth of a cell that expresses a TAT polypeptide, wherein the method comprises contacting the cell with an antibody, an oligopeptide or a small organic molecule that binds to the TAT polypeptide, and wherein the binding of the antibody, oligopeptide or organic molecule to the TAT polypeptide causes inhibition of the growth of the cell expressing the TAT polypeptide. In preferred embodiments, the cell is a cancer cell and binding of the antibody, oligopeptide or organic molecule to the TAT polypeptide causes death of the cell expressing the TAT polypeptide. Optionally, the antibody is a monoclonal antibody, antibody fragment, chimeric antibody, humanized antibody, or single-chain antibody. Antibodies, TAT binding oligopeptides and TAT binding organic molecules employed in the methods of the present invention may optionally be conjugated to a growth inhibitory agent or cytotoxic agent such as a toxin, including, for example, a maytansinoid or calicheamicin, an antibiotic, a radioactive isotope, a nucleolytic enzyme, or the like. The antibodies and TAT binding oligopeptides employed in the methods of the present invention may optionally be produced in CHO cells or bacterial cells.

Yet another embodiment of the present invention is directed to a method of therapeutically treating a mammal having a cancerous tumor comprising cells that express a TAT polypeptide, wherein the method comprises administering to the mammal a therapeutically effective amount of an antibody, an oligopeptide or a small organic molecule that binds to the TAT polypeptide, thereby resulting in the effective therapeutic treatment of the tumor. Optionally, the antibody is a monoclonal antibody, antibody fragment, chimeric antibody, humanized antibody, or single-chain antibody. Antibodies, TAT binding oligopeptides and TAT binding organic molecules employed in the methods of the present invention may optionally be conjugated to a growth inhibitory agent or cytotoxic agent such as a toxin, including, for example, a maytansinoid or calicheamicin, an antibiotic, a radioactive isotope, a nucleolytic enzyme, or the like. The antibodies and oligopeptides employed in the methods of the present invention may optionally be produced in CHO cells or bacterial cells.

Yet another embodiment of the present invention is directed to a method of determining the presence of a TAT polypeptide in a sample suspected of containing the TAT polypeptide, wherein the method comprises exposing the sample to an antibody, oligopeptide or small organic molecule that binds to the TAT polypeptide and determining binding of the antibody, oligopeptide or organic molecule to the TAT polypeptide in the sample, wherein the presence of such binding is indicative of the presence of the TAT polypeptide in the sample. Optionally, the sample may contain cells (which may be cancer cells) suspected of expressing the TAT polypeptide. The antibody, TAT binding oligopeptide or TAT binding organic molecule employed in the method may optionally be detectably labeled, attached to a solid support, or the like.

A further embodiment of the present invention is directed to a method of diagnosing the presence of a tumor in a mammal, wherein the method comprises detecting the level of expression of a gene encoding a TAT polypeptide (a) in a test sample of tissue cells obtained from said mammal, and (b) in a control sample of known normal non-cancerous cells of the same tissue origin or type, wherein a higher level of expression of the

TAT polypeptide in the test sample, as compared to the control sample, is indicative of the presence of tumor in the mammal from which the test sample was obtained.

Another embodiment of the present invention is directed to a method of diagnosing the presence of a tumor in a mammal, wherein the method comprises (a) contacting a test sample comprising tissue cells obtained from the mammal with an antibody, oligopeptide or small organic molecule that binds to a TAT polypeptide and (b) detecting the formation of a complex between the antibody, oligopeptide or small organic molecule and the TAT polypeptide in the test sample, wherein the formation of a complex is indicative of the presence of a tumor in the mammal. Optionally, the antibody, TAT binding oligopeptide or TAT binding organic molecule employed is detectably labeled, attached to a solid support, or the like, and/or the test sample of tissue cells is obtained from an individual suspected of having a cancerous tumor.

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Yet another embodiment of the present invention is directed to a method for treating or preventing a cell proliferative disorder associated with altered, preferably increased, expression or activity of a TAT polypeptide, the method comprising administering to a subject in need of such treatment an effective amount of an antagonist of a TAT polypeptide. Preferably, the cell proliferative disorder is cancer and the antagonist of the TAT polypeptide is an anti-TAT polypeptide antibody, TAT binding oligopeptide, TAT binding organic molecule or antisense oligonucleotide. Effective treatment or prevention of the cell proliferative disorder may be a result of direct killing or growth inhibition of cells that express a TAT polypeptide or by antagonizing the cell growth potentiating activity of a TAT polypeptide.

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Yet another embodiment of the present invention is directed to a method of binding an antibody, oligopeptide or small organic molecule to a cell that expresses a TAT polypeptide, wherein the method comprises contacting a cell that expresses a TAT polypeptide with said antibody, oligopeptide or small organic molecule under conditions which are suitable for binding of the antibody, oligopeptide or small organic molecule to said TAT polypeptide and allowing binding therebetween.

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Other embodiments of the present invention are directed to the use of (a) a TAT polypeptide, (b) a nucleic acid encoding a TAT polypeptide or a vector or host cell comprising that nucleic acid, (c) an anti-TAT polypeptide antibody, (d) a TAT-binding oligopeptide, or (e) a TAT-binding small organic molecule in the preparation of a medicament useful for (i) the therapeutic treatment or diagnostic detection of a cancer or tumor, or (ii) the therapeutic treatment or prevention of a cell proliferative disorder.

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Another embodiment of the present invention is directed to a method for inhibiting the growth of a cancer cell, wherein the growth of said cancer cell is at least in part dependent upon the growth potentiating effect(s) of a TAT polypeptide (wherein the TAT polypeptide may be expressed either by the cancer cell itself or a cell that produces polypeptide(s) that have a growth potentiating effect on cancer cells), wherein the method comprises contacting the TAT polypeptide with an antibody, an oligopeptide or a small organic molecule that binds to the TAT polypeptide, thereby antagonizing the growth-potentiating activity of the TAT polypeptide and, in turn, inhibiting the growth of the cancer cell. Preferably the growth of the cancer cell is completely inhibited. Even more preferably, binding of the antibody, oligopeptide or small organic molecule to the TAT polypeptide induces the death of the cancer cell. Optionally, the antibody is a monoclonal antibody, antibody fragment,

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chimeric antibody, humanized antibody, or single-chain antibody. Antibodies, TAT binding oligopeptides and TAT binding organic molecules employed in the methods of the present invention may optionally be conjugated to a growth inhibitory agent or cytotoxic agent such as a toxin, including, for example, a maytansinoid or calicheamicin, an antibiotic, a radioactive isotope, a nucleolytic enzyme, or the like. The antibodies and TAT binding oligopeptides employed in the methods of the present invention may optionally be produced in CHO cells or bacterial cells.

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Yet another embodiment of the present invention is directed to a method of therapeutically treating a tumor in a mammal, wherein the growth of said tumor is at least in part dependent upon the growth potentiating effect(s) of a TAT polypeptide, wherein the method comprises administering to the mammal a therapeutically effective amount of an antibody, an oligopeptide or a small organic molecule that binds to the TAT polypeptide, thereby antagonizing the growth potentiating activity of said TAT polypeptide and resulting in the effective therapeutic treatment of the tumor. Optionally, the antibody is a monoclonal antibody, antibody fragment, chimeric antibody, humanized antibody, or single-chain antibody. Antibodies, TAT binding oligopeptides and TAT binding organic molecules employed in the methods of the present invention may optionally be conjugated to a growth inhibitory agent or cytotoxic agent such as a toxin, including, for example, a maytansinoid or calicheamicin, an antibiotic, a radioactive isotope, a nucleolytic enzyme, or the like. The antibodies and oligopeptides employed in the methods of the present invention may optionally be produced in CHO cells or bacterial cells.

Yet further embodiments of the present invention will be evident to the skilled artisan upon a reading of the present specification.

BRIEF DESCRIPTION OF THE DRAWINGS

In the list of figures for the present application, specific cDNA sequences which are upregulated in certain tumor tissues as compared to their normal tissue counterparts are individually identified with a designation beginning with the letters "DNA" followed by a specific numerical designation. A full or partial length protein sequence that is encoded by a cDNA sequence identified and shown herein is individually identified with a designation beginning with the letters "PRO" followed by a specific numerical designation. Figures showing encoded amino acid sequences immediately follow the figure showing the cDNA sequence encoding that specific amino acid sequence. If start and/or stop codons have been identified in a cDNA sequence shown in the attached figures, they are shown in bold and underlined font.

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Figure 20: DNA323729, XM_166599, gen.XM_166599	Figure 70: PRO80510
Figure 21: PRO80487	Figure 71: DNA323755, NM_003689, gen.NM_003689
Figure 22: DNA323730, NM_017900, gen.NM_017900	Figure 72: PRO80511
Figure 23: PRO80488	Figure 73: DNA323756, NM_016183, gen.NM_016183
Figure 24: DNA323731, XM_001589, gen.XM_001589	Figure 74: PRO80512
Figure 25: PRO80489	Figure 75: DNA323757, XM_015234, gen.XM_015234
Figure 26: DNA323732, NM_016176, gen.NM_016176	Figure 76A-B: DNA323758, XM_027916,
Figure 27: PRO80490	gen.XM_027916
Figure 28: DNA323733, XM_117692, gen.XM_117692	Figure 77: DNA323759, XM_033683, gen.XM_033683
Figure 29: DNA323734, XM_086360, gen.XM_086360	Figure 78: DNA323760, XM_001826, gen.XM_001826
Figure 30: PRO80492	Figure 79: DNA323761, XM_033654, gen.XM_033654
Figure 31: DNA287173, NM_001428, gen.NM_001428	Figure 80: PRO80517
Figure 32: PRO69463	Figure 81: DNA323762, NM _001791, gen.NM _001791
Figure 33: DNA323735, XM_001299, gen.XM_001299	Figure 82: PRO26194
Figure 34: DNA323736, NM_000983, gen.NM_000983	Figure 83: DNA323763, NM_005826, gen.NM_005826
Figure 35: PRO80493	Figure 84: PRO60815
Figure 36A-B: DNA227821, NM_014851,	Figure 85: DNA323764, XM_086357, gen.XM_086357
gen.NM_014851	Figure 86: PRO80518
Figure 37: PRO38284	Figure 87: DNA323765, NM_000975, gen.NM_000975
Figure 38A-B: DNA323737, XM_086204,	Figure 88: PRO80519
gen.XM_086204	Figure 89: DNA323766, NM _007260, gen.NM _007260
Figure 39: PRO80494	Figure 90: PRO61250
Figure 40: DNA323738, XM_030920, gen.XM_030920	Figure 91: DNA323767, NM_017761, gen.NM_017761
Figure 41: DNA323739, NM_018948, gen.NM_018948	Figure 92: PRO80520
Figure 42: DNA273712, NM_007262, gen.NM_007262	Figure 93: DNA323768, NM _006625, gen.NM _006625
Figure 43: PRO61679	Figure 94: PRO22196
Figure 44: DNA151148, NM_004781, gen.NM_004781	Figure 95: DNA323769, NM_054016, gen.NM_054016
Figure 45: PRO12618	Figure 96: PRO80521
Figure 46: DNA323740, XM_086151, gen.XM_086151	Figure 97: DNA323770, XM _086375, gen.XM _086375
Figure 47: PRO80497	Figure 98: DNA323771, XM _006290, gen.XM _006290
Figure 48: DNA171408, NM_004401, gen.NM_004401	Figure 99: DNA323772, NM_015484, gen.NM_015484
Figure 49: PRO20136	Figure 100: PRO80524
Figure 50: DNA323741, NM_003132, gen.NM_003132	Figure 101A-B: DNA323773, XM_001616,
Figure 51: PRO80498	gen.XM_001616
Figure 52: DNA323742, XM_086586, gen.XM_086586	Figure 102: DNA323774, XM_058240,

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Figure 207: PRO80575

Figure 208: DNA323828, XM_046557, gen.XM_059171 gen.XM_046557 Figure 172: PRO80563 Figure 173: DNA83085, NM_000760, gen.NM_000760 Figure 209: PRO80576 Figure 210: DNA323829, NM_001012, Figure 174: PRO2583 Figure 175: DNA323815, XM_165984, gen.NM_001012 Figure 211: PRO10760 gen.XM_165984 Figure 212: DNA323830, XM_046551, Figure 176: DNA323816, XM_029842, gen.XM_046551 gen.XM_029842 Figure 213A-B: DNA323831, XM_027983, Figure 177: PRO2851 gen.XM_027983 Figure 178: DNA323817, XM_086384, Figure 214: DNA323832, XM_086324, gen.XM_086384 gen.XM_086324 Figure 179: PRO80565 Figure 215: PRO80579 Figure 180A-C: DNA274487, NM_014747, Figure 216: DNA323833, XM_032391, gen.NM_014747 gen.XM_032391 Figure 181: PRO62389 Figure 217: PRO80580 Figure 182: DNA323818, XM_010712, Figure 218: DNA103214, NM_006066, gen.XM_010712 gen.NM_006066 Figure 183: DNA323819, NM_024664, Figure 219: PRO4544 gen.NM_024664 Figure 220: DNA304686, NM_002574, Figure 184: PRO80567 gen.NM_002574 Figure 185: DNA323820, XM_059214, Figure 221: PRO71112 gen.XM_059214 Figure 222: DNA323834, NM_032756, Figure 186: PRO80568 gen.NM_032756 Figure 187: DNA323821, XM_046349, Figure 223: PRO80581 gen.XM_046349 Figure 224: DNA323835, XM_059133, Figure 188: DNA103253, NM_006516, gen.XM_059133 gen.NM_006516 Figure 225: PRO80582 Figure 189: PRO4583 Figure 226: DNA323836, XM_027313, Figure 190: DNA323822, XM_086543, gen.XM_027313 gen.XM_086543 Figure 227: PRO80583 Figure 191: PRO80570 Figure 228: DNA323837, XM .054868, Figure 192: DNA274745, NM_006824, gen.XM_054868 gen.NM_006824 Figure 229: DNA323838, NM_001262, Figure 193: PRO62518 gen.NM_001262 Figure 194: DNA273060, NM_001255, Figure 230: PRO59546 gen.NM_001255 . Figure 231: DNA323839, XM_086391, Figure 195: PRO61125 gen.XM_086391 Figure 196: DNA323823, NM_030587, Figure 232: PRO80584 gen.NM_030587 Figure 233: DNA323840, XM_114798, Figure 197: PRO80571 gen.XM_114798 Figure 198: DNA323824, XM_097649, Figure 234: PRO80585 gen.XM_097649 Figure 235: DNA272748, NM_002979, Figure 199: DNA256503, NM _003780, gen.NM_002979 gen.NM_003780 Figure 236: PRO60860 Figure 200: PRO51539 Figure 237: DNA323841, XM_038911, Figure 201: DNA323825, XM_046450, gen.XM_038911 gen.XM_046450 Figure 238: PRO80586 Figure 202A-B: DNA272024, NM_014663, Figure 239: DNA323842, NM_018070, gen.NM_014663 gen.NM_018070 Figure 203: PRO60298 Figure 240: PRO80587 Figure 204: DNA323826, XM_046565, Figure 241: DNA323843, NM_024603, gen.XM_046565 gen.NM_024603 Figure 205: PRO80574 Figure 242: PRO80588 Figure 206: DNA323827, NM_024602, Figure 243: DNA323844, XM_086389, gen.NM_024602

gen.XM_086389

Figure 244: DNA323845, XM_038852,	Figure 278: PRO80607
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Figure 245: DNA323846, NM_032864,	gen.XM_086165
gen.NM_032864	Figure 280: DNA323866, XM_086167,
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Figure 247: DNA323847, NM_024586,	Figure 281: DNA323867, XM_086166,
gen.NM_024586	gen.XM_086166
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Figure 249A-B: DNA323848, XM_097565,	gen.XM_086138
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gen.XM_001472	gen.NM_000969
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Figure 256A-B: DNA323853, XM_059180,	gen.XM_165981
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Figure 257: DNA323854, XM_015717,	Figure 292: DNA275139, NM_013296,
gen.XM_015717	gen.NM_013296
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Figure 259: DNA323855, XM_114125,	Figure 294: DNA323872, XM_058702,
gen.XM_114125	gen.XM_058702
Figure 260: DNA323856, NM_015640,	Figure 295: DNA323873, XM 054978,
gen.NM_015640	gen.XM_054978
Figure 261: PRO80599	Figure 296: DNA323874, NM _032636,
Figure 262: DNA323857, NM_017768,	gen.NM_032636
gen.NM_017768	Figure 297: PRO80617
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Figure 264: DNA323858, XM_165977,	gen.NM_006513
gen.XM_165977	Figure 299: PRO80618
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gen.XM_086343	gen.NM_006621
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gen.NM_007034	gen.NM_007158
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gen.NM_001554	gen.XM_086132
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gen.NM_006769	gen.NM_004000 Figure 307: PRO80622
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gen.NM_004261	gen.NM_001688
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gen.XM_165983	gen.NM_019099
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	gen.NM_018116
gen.XM_114055	Figure 421: PRO80668
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gen.XM_113360	gen.NM_002004
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gen.XM_086564	gen.NM_005698
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gen.NM_005973	Figure 426: DNA323938, NM_052837,
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gen.XM_044077	Figure 428: DNA194600, NM_006589,
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gen.XM_044127	gen.NM_130898
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gen.XM_053245	gen.NM_006694
Figure 401: PRO80659	Figure 437: PRO37256
Figure 402: DNA257916, NM_032323,	Figure 438: DNA294794, NM _002870,
gen.NM_032323	gen.NM_002870
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gen.NM_005572	gen.NM_001030
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gen.XM_044166	gen.XM_036829
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gen.XM_044128	gen.NM_015449
Figure 409: DNA226125, NM_003145,	Figure 445: PRO80678 '
gen.NM_003145	Figure 446: DNA323946, NM .014847,
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gen.NM_014849	gen.NM_003617
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gen.XM_059094	gen.NM_025226
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	gen.XM_046464
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Figure 533: PRO80717	gen.XM_001914
Figure 534: DNA323995, XM_117181,	Figure 568: DNA324013, XM_001916,
gen.XM_117181	gen.XM_001916
Figure 535: DNA323996, NM_018122,	Figure 569: DNA324014, NM _018085,
gen.NM_018122	gen.NM_018085
Figure 536: PRO80719	Figure 570: PRO80734
Figure 537: DNA323997, XM_042967,	Figure 571: DNA324015, NM .006335,
gen.XM_042967	gen.NM_006335
Figure 538: DNA323998, XM _086494,	Figure 572: PRO80735
gen.XM_086494	Figure 573: DNA324016, XM_036500,
Figure 539: PRO80720	gen.XM_036500
Figure 540: DNA290234, NM _002923,	Figure 574: PRO80736
gen.NM_002923	Figure 575: DNA324017, XM _036507,
Figure 541: PRO70333	gen.XM_036507
Figure 542: DNA323999, XM_086328,	Figure 576: DNA196344, NM_004767,
gen.XM_086328	gen.NM_004767
Figure 543: DNA324000, XM_086282,	Figure 577: PRO24851
gen.XM_086282	Figure 578: DNA247474, NM_014176,
Figure 544: DNA324001, XM_053633,	gen.NM_014176
gen.XM_053633	Figure 579: PRO44999
Figure 545: DNA256905, NM_138391,	Figure 580A-B: DNA324018, XM_084055,
gen.NM_138391	gen.XM_084055
Figure 546: PRO51836	Figure 581: DNA324019, XM_010682,
Figure 547: DNA324002, XM_015434,	gen.XM_010682
gen.XM_015434	Figure 582: DNA324020, XM_117185,
Figure 548: DNA324003, NM_006763,	gen.XM_117185
gen.NM_006763	Figure 583: DNA324021, XM_055880,
Figure 549: PRO80725	gen.XM_055880
Figure 550: DNA227246, NM_005686,	Figure 584: PRO80740
gen.NM_005686	Figure 585: DNA193882, NM_014184,
Figure 551: PRO37709	gen.NM_014184
Figure 552: DNA324004, XM_058405,	Figure 586: PRO23300
gen.XM_058405	Figure 587: DNA324022, NM_018212,
Figure 553A-B: DNA226005, NM_000228,	gen.NM_018212
gen.NM_000228	Figure 588: PRO80741
Figure 554: PRO36468	Figure 589: DNA324023, XM_086431,
Figure 555: DNA324005, NM_015714,	gen.XM_086431
gen.NM_015714	Figure 590: PRO80742
Figure 556: PRO11582	Figure 591: DNA324024, XM_037329,
Figure 557: DNA324006, XM _086142,	gen.XM_037329
gen.XM_086142	Figure 592: DNA324025, XM_086432,
Figure 558: DNA83046, NM _000574, gen.NM _000574	· · · · · · · · · · · · · · · · · · ·
Figure 559: PRO2569	Figure 593A-B: DNA324026, XM _010732,
115010 337. 1102307	

gen.XM_010732 gen.XM_056970 Figure 629: PRO80762 Figure 594: DNA227504, NM_000447, gen.NM_000447 Figure 630: DNA324046, NM_032324, Figure 595: PRO37967 gen.NM_032324 Figure 631: PRO80763 Figure 596: DNA324027, NM_012486, Figure 632: DNA324047, XM_086257, gen.NM_012486 Figure 597: PRO80745 gen.XM_086257 Figure 633: PRO80764 Figure 598A-B: DNA324028, XM_113361, Figure 634: DNA324048, XM_114137, gen.XM_113361 Figure 599A-B: DNA324029, XM_001958, gen.XM_114137 gen.XM_001958 Figure 635: PRO80765 Figure 636: DNA324049, NM .000143, Figure 600: DNA324030, XM_016199, gen.NM_000143 gen.XM_016199 Figure 601: DNA324031, XM_086244, Figure 637: PRO62607 Figure 638: DNA324050, XM _090833, gen.XM_086244 gen.XM_090833 Figure 602: DNA324032, XM_086245, Figure 639: DNA324051, NM_130398, gen.XM_086245 gen.NM_130398 Figure 603: DNA254346, NM_024709, Figure 640: PRO80767 gen.NM_024709 Figure 641: DNA324052, XM_117196, Figure 604: PRO49457 Figure 605: DNA324033, XM_088107, gen.XM_117196 gen.XM_088107 Figure 642: DNA324053, XM_018041, Figure 606: DNA324034, NM_032890, gen.XM_018041 Figure 643: DNA324054, NM_001011, gen.NM_032890 Figure 607: PRO80752 gen.NM_001011 Figure 644: PRO10692 Figure 608: DNA324035, XM .052974, Figure 645: DNA324055, NM_024027, gen.XM_052974 gen.NM_024027 Figure 609: PRO80753 Figure 610: DNA324036, XM_047499, Figure 646: PRO1182 Figure 647: DNA324056, NM_016030, gen.XM_047499 gen.NM_016030 Figure 611: PRO80754 Figure 648: PRO80770 Figure 612: DNA324037, NM_000858, Figure 649: DNA103217, NM_003310, gen.NM_000858 gen.NM_003310 Figure 613: PRO80755 Figure 650: PRO4547 Figure 614: DNA324038, NM_024319, gen.NM_024319 Figure 651: DNA275195, NM_001034, gen.NM_001034 Figure 615: PRO80756 Figure 652: PRO62893 Figure 616: DNA324039, XM_047545, Figure 653: DNA324057, XM_059368, gen.XM_047545 Figure 617: PRO4914 gen.XM_059368 Figure 654: PRO80771 Figure 618A-B: DNA324040, XM_056884, Figure 655: DNA324058, NM_006826, gen.XM_056884 gen.NM_006826 Figure 619: DNA324041, XM_098599, gen.XM_098599 Figure 656: PRO70258 Figure 620: DNA324042, XM_165439, Figure 657: DNA324059, NM_005378, gen.XM_165439 gen.NM_005378 Figure 658: PRO80772 Figure 621: PRO80759 Figure 659: DNA324060, NM _002539, Figure 622: DNA324043, XM_089030, gen.NM_002539 gen.XM_089030 Figure 660: PRO80773 Figure 623: PRO80760 Figure 661: DNA324061, XM_096149, Figure 624: DNA82328, NM_000029, gen.NM_000029 gen.XM_096149 Figure 625: PRO1707 Figure 662: DNA275049, NM_004939, Figure 626: DNA324044, NM_014236, gen.NM_014236 gen.NM_004939 Figure 627: PRO80761 Figure 663: PRO62770 Figure 628: DNA324045, XM_056970, Figure 664A-B: DNA324062, XM_036450,

gen.NM_021095

Figure 699: PRO38008

Figure 700: DNA324079, XM_002435,

gen.XM_036450 gen.XM_002435 Figure 665: DNA324063, XM_103946, Figure 701: DNA324080, NM_000221, gen.XM_103946 gen.NM_000221 Figure 666: PRO80775 Figure 702: PRO80790 Figure 667: DNA324064, NM_014713, Figure 703: DNA271243, NM_006488, gen.NM_014713 gen.NM_006488 Figure 668: PRO80776 Figure 704: PRO59558 Figure 669: DNA324065, XM_087206, Figure 705: DNA324081, NM_007046, gen.XM_087206 gen.NM_007046 Figure 670: DNA324066, NM_106552, Figure 706: PRO9886 gen.NM_106552 Figure 707: DNA324082, NM_021831. Figure 671: PRO80778 gen.NM_021831 Figure 672: DNA324067, XM_092135, Figure 708: PRO80791 gen.XM_092135 Figure 709: DNA324083, NM_020134, Figure 673: PRO80779 gen.NM_020134 Figure 674: DNA324068, NM_017910, Figure 710: PRO80792 gen.NM_017910 Figure 711: DNA103593, NM_000183, Figure 675: PRO80780 gen.NM_000183 Figure 676: DNA324069, XM_092517, Figure 712: PRO4917 gen.XM_092517 Figure 713: DNA324084, NM_000182, Figure 677: PRO80781 gen.NM_000182 Figure 678A-B: DNA324070, NM_025203, Figure 714: PRO80793 gen.NM_025203 Figure 715: DNA324085, XM_097976, Figure 679: PRO80782 gen.XM_097976 Figure 680: DNA324071, XM_002480, Figure 716A-B: DNA324086, XM_039712, gen.XM_002480 gen.XM_039712 Figure 681: DNA324072, NM_002707, Figure 717: DNA324087, NM_022552, gen.NM_002707 gen.NM_022552 Figure 682: PRO12199 Figure 718: PRO80796 Figure 683: DNA324073, XM_087151, Figure 719: DNA324088, NM_024572, gen.XM_087151 gen.NM_024572 Figure 684: DNA227165, NM_014748, Figure 720: PRO80797 gen.NM_014748 Figure 721: DNA324089, NM_018607, Figure 685: PRO37628 gen.NM_018607 Figure 686: DNA324074, NM_015636, Figure 722: PRO80798 gen.NM_015636 Figure 723: DNA324090, XM_165448, Figure 687: PRO80785 gen.XM_165448 Figure 688: DNA273800, NM_001521, Figure 724: PRO80799 gen.NM_001521 Figure 725: DNA324091, XM_087195. Figure 689: PRO61761 gen.XM_087195 Figure 690: DNA324075, XM_047175, Figure 726: DNA324092, XM_087193. gen.XM_047175 gen.XM_087193 Figure 691: PRO80786 Figure 727: DNA324093, NM_138801, Figure 692A-B: DNA324076, NM_004341, gen.NM_138801 gen.NM_004341 Figure 728: PRO80802 Figure 693: PRO80787 Figure 729: DNA324094, XM_098004. Figure 694: DNA324077, NM_016085, gen.XM_098004 gen.NM_016085 Figure 730: PRO80803 Figure 695: PRO80788 Figure 731: DNA324095, XM_031519, Figure 696: DNA324078, NM_080592, gen.XM_031519 gen.NM_080592 Figure 732: PRO80804 Figure 697: PRO80789 Figure 733A-B: DNA324096, XM_031527, Figure 698: DNA227545, NM_021095, gen.XM_031527

gen.XM_038576

Figure 735: PRO80806

Figure 734: DNA324097, XM_038576,

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Figure 736: DNA324098, XM_117264, gen.XM_010881 gen.XM_117264 Figure 772: DNA324115, XM_087069, Figure 737: PRO80807 gen.XM_087069 Figure 738A-B: DNA324099, XM_031626, Figure 773: DNA324116, XM_016625, gen.XM_031626 gen.XM_016625 Figure 739: PRO80808 Figure 774: PRO80820 Figure 740: DNA324100, XM_057664, Figure 775: DNA324117, XM_087068, gen.XM_057664 gen.XM_087068 Figure 741: DNA226428, NM _000251, Figure 776: DNA324118, XM_002674, gen.NM_000251 gen.XM_002674 Figure 742: PRO36891 Figure 777: DNA324119, XM_065884. Figure 743: DNA324101, XM_087211, gen.XM_065884 gen.XM_087211 Figure 778: PRO80823 Figure 744A-B: DNA275066, NM_000179, Figure 779A-B: DNA324120, XM_002739, gen.NM_000179 gen.XM_002739 Figure 745: PRO62786 Figure 780: DNA324121, XM_031596, Figure 746A-C: DNA270154, NM_003128, gen.XM_031596 gen.NM_003128 Figure 781: PRO61325 Figure 747: PRO58543 Figure 782: DNA324122, XM_031585, Figure 748: DNA324102, XM_087051, gen.XM_031585 gen.XM_087051 Figure 783: DNA324123, XM_031586, Figure 749: DNA324103, NM _002954, gen.XM_031586 gen.NM_002954 Figure 784: DNA324124, XM_018039, Figure 750: PRO62239 gen.XM_018039 Figure 751: DNA271060, NM_002453, Figure 785: DNA324125, NM_032822, gen.NM_002453 gen.NM_032822 Figure 752: PRO59384 Figure 786: PRO80827 Figure 753: DNA324104, XM_048088, Figure 787A-B: DNA324126, XM_096172, gen.XM_048088 gen.XM_096172 Figure 788A-B: DNA324127, XM_002727, Figure 754: PRO80811 Figure 755: DNA324105, XM_010886, gen.XM_002727 gen.XM_010886 Figure 789: DNA324128, NM_003124, Figure 756: PRO80812 gen.NM_003124 Figure 757: DNA324106, XM_045283, Figure 790: PRO80830 gen.XM_045283 Figure 791: DNA324129, XM_086980, Figure 758: PRO80813 gen.XM_086980 Figure 759: DNA324107, NM_006430, Figure 792: DNA227795, NM_006429. gen.NM_006430 gen.NM_006429 Figure 760: PRO80814 Figure 793: PRO38258 Figure 761A-B: DNA324108, NM _003400, Figure 794: DNA287167, NM_006636, gen.NM_003400 gen.NM_006636 Figure 762: PRO59544 Figure 795: PRO59136 Figure 763: DNA324109, XM_018301, Figure 796: DNA324130, NM_033046, gen.XM_018301 gen.NM_033046 Figure 764: DNA324110, NM_005917, Figure 797: PRO80832 gen.NM_005917 Figure 798: DNA324131, NM_133637, Figure 765: PRO4918 gen.NM_133637 Figure 766: DNA324111, XM_016843, Figure 799: PRO80833 gen.XM_016843 Figure 800: DNA324132, XM_035220, Figure 767: PRO80816 gen.XM_035220 Figure 768: DNA324112, XM_088638, Figure 801: DNA324133, NM_013247, gen.XM_088638 gen.NM_013247 Figure 769: PRO80817 Figure 802: PRO80835 Figure 770: DNA324113, XM_002647, Figure 803: DNA227528, NM_021103, gen.XM_002647 gen.NM_021103 Figure 771: DNA324114, XM_010881, Figure 804: PRO37991

Figure 839: DNA324153, XM _087122,

Figure 805: DNA324134, XM .086920, gen, XM_087122 Figure 840: PRO80853 gen.XM_086920 Figure 806: DNA150725, NM_001747, Figure 841: DNA324154, XM_018540, gen.NM_001747 gen.XM_018540 Figure 807: PRO12792 Figure 842: DNA324155, XM_087040, Figure 808: DNA324135, NM .005911, gen.XM_087040 gen.NM_005911 Figure 843: DNA324156, NM_032212, Figure 809: PRO80837 gen.NM_032212 Figure 810: DNA324136, NM_032827, Figure 844: PRO80856 Figure 845: DNA324157, XM_002217, gen.NM_032827 Figure 811: PRO80838 gen.XM_002217 Figure 812: DNA324137, NM_017952, Figure 846: PRO80857 gen.NM_017952 Figure 847: DNA324158, NM_000576, gen.NM_000576 Figure 813: PRO80839 Figure 814: DNA227190, NM_006839, Figure 848: PRO65 gen.NM_006839 Figure 849: DNA324159, XM .086923, Figure 815: PRO37653 gen.XM_086923 Figure 850: DNA324160, XM_086925, Figure 816: DNA324138, XM_114215, gen.XM_086925 gen.XM_114215 Figure 851A-B: DNA324161, XM_114266, Figure 817: DNA324139, XM_052989, gen.XM_114266 gen.XM_052989 Figure 852: PRO80860 Figure 818: DNA324140, XM_049116, Figure 853: DNA324162, XM_002704, gen.XM_049116 gen.XM_002704 Figure 819: PRO80842 Figure 854: DNA194740, NM_005291, Figure 820A-B: DNA324141, XM_049108, gen.NM_005291 gen.XM_049108 Figure 855: PRO24028 Figure 821: PRO80843 Figure 856A-B: DNA324163, XM_114267, Figure 822: DNA324142, XM_049113, gen.XM_114267 gen.XM_049113 Figure 857: DNA324164, XM_034952, Figure 823: DNA324143, XM_002611, gen.XM_034952 gen.XM_002611 Figure 824A-B: DNA324144, XM_114247, Figure 858: DNA324165, XM_086950, gen.XM_086950 gen.XM_114247 Figure 859A-B: DNA255531, NM_017751, Figure 825: DNA324145, NM_017789, gen.NM_017751 gen.NM_017789 Figure 860: PRO50596 Figure 826: PRO80846 Figure 827: DNA324146, NM_001862, Figure 861: DNA324166, XM_017698, gen.NM_001862 gen.XM_017698. Figure 828: PRO80847 Figure 862: DNA324167, XM_030529, gen.XM_030529 Figure 829: DNA324147, NM .005783, Figure 863: PRO80866 gen.NM_005783 Figure 864: DNA275240, NM_005915, Figure 830: PRO80848 gen.NM_005915 Figure 831A-B: DNA324148, XM_037108, Figure 865: PRO62927 gen.XM_037108 Figure 866: DNA324168, XM_043173, Figure 832: DNA324149, NM_000993, gen.XM_043173 gen.NM_000993 Figure 833: PRO11197 Figure 867: DNA324169, XM_092489, Figure 834: DNA324150, NM_017546, gen.XM_092489 Figure 868: PRO80868 gen.NM_017546 Figure 869: DNA324170, XM_115672, Figure 835: PRO80850 gen.XM_115672 Figure 836: DNA324151, NM_001450, Figure 870: PRO80869 gen.NM_001450 Figure 871: DNA324171, NM_020548, Figure 837: PRO80851 gen.NM_020548 Figure 838: DNA324152, XM_114229, Figure 872: PRO60753 gen.XM_114229

Figure 873: DNA324172, XM_037101,

gen.XM_015920

gen.XM_037101 Figure 910: DNA324190, XM_166007, Figure 874: PRO80870 gen.XM_166007 Figure 875: DNA324173, NM_032390, Figure 911: DNA324191, XM_015922, gen.NM_032390 gen.XM_015922 Figure 876: PRO80871 Figure 912: DNA324192, XM_087061, Figure 877: DNA324174, XM_002447, gen.XM_087061 gen.XM_002447 Figure 913: PRO80888 Figure 878: DNA324175, NM_033416, Figure 914: DNA324193, XM_087062, gen.NM_033416 gen.XM_087062 Figure 879: PRO80873 Figure 915: PRO80889 Figure 880: DNA324176, XM_016288, Figure 916: DNA324194, NM_001463, gen.XM_016288 gen.NM_001463 Figure 881: DNA272127, NM_003937, Figure 917: PRO80890 gen.NM_003937 Figure 918: DNA324195, XM_092158, Figure 882: PRO60397 gen.XM_092158 Figure 883: DNA324177, XM_030582, Figure 919: PRO80891 gen.XM_030582 Figure 920: DNA324196, XM_059351. Figure 884: PRO80875 gen.XM_059351 Figure 885: DNA324178, NM_015702, Figure 921A-B: DNA324197, NM_000090, gen.NM_015702 gen.NM_000090 Figure 886: PRO80876 Figure 922: PRO2665 Figure 887: DNA324179, NM_016838. Figure 923: DNA324198, NM_014585, gen.NM_016838 gen.NM_014585 Figure 888: PRO80877 Figure 924: PRO37675 Figure 889: DNA324180, NM_016839, Figure 925: DNA324199, XM_010778, gen.NM_016839 gen.XM_010778 Figure 890: PRO80878 Figure 926: DNA324200, XM_086961, Figure 891: DNA324181, XM_087118, gen.XM_086961 gen.XM_087118 Figure 927: DNA324201, XM_165994, Figure 892: PRO80879 gen.XM_165994 Figure 893: DNA324182, XM_165998, Figure 928: DNA324202, XM_045170, gen.XM_165998 gen.XM_045170 Figure 894: DNA324183, NM_001935. Figure 929: DNA324203, XM_113390. gen.NM_001935 gen.XM_113390 Figure 895: PRO80881 Figure 930: DNA299899, NM_002157, Figure 896: DNA324184, NM_020675, gen.NM_002157 gen.NM_020675 Figure 931: PRO62760 Figure 897: PRO80882 Figure 932: DNA324204, XM_087045, Figure 898: DNA88051, NM_000079, gen.NM_000079 gen.XM_087045 Figure 899: PRO2146 Figure 933: DNA324205, XM_086944, Figure 900: DNA324185, XM_166008, gen.XM_086944 gen.XM_166008 Figure 934: DNA271608, NM_014670. Figure 901: DNA324186, XM_087240, gen.NM_014670 gen.XM_087240 Figure 935: PRO59895 Figure 902: PRO11403 Figure 936: DNA324206, XM_027963, Figure 903: DNA324187, NM_013341, gen.XM_027963 gen.NM_013341 Figure 937: PRO80900 Figure 904: PRO80883 Figure 938: DNA324207, XM_010852, Figure 905: DNA304805, NM_031942, gen.XM_010852 gen.NM_031942 Figure 939: PRO80901 Figure 906: PRO69531 Figure 940: DNA324208, XM_028034, Figure 907: DNA324188, XM_059465, gen.XM_028034 gen.XM_059465 Figure 941: DNA324209, NM_015934, Figure 908: PRO80884 gen.NM_015934 Figure 909: DNA324189, XM_015920. Figure 942: DNA324210, XM_087028,

gen.XM_087028

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Figure 943: PRO80903 Figure 979: DNA324230, XM_050638, Figure 944: DNA324211, XM_092346, gen.XM_050638 gen.XM_092346 Figure 980A-B: DNA324231, NM _002846, Figure 945: PRO80904 gen.NM_002846 Figure 946: DNA324212, XM_002669, Figure 981: PRO2610 gen.XM_002669 Figure 982: DNA324232, NM_006000, Figure 947: PRO80905 gen.NM_006000 Figure 948: DNA324213, NM_021121, Figure 983: PRO26228 gen.NM_021121 Figure 984: DNA324233, XM_050891, Figure 949: PRO23124 gen.XM_050891 Figure 950: DNA324214, NM _001959, Figure 985: DNA324234, XM_087162, gen.NM_001959 gen.XM_087162 Figure 951: PRO23124 Figure 986: DNA324235, XM_058098, Figure 952: DNA324215, XM_030834, gen.XM_058098 gen.XM_030834 Figure 987: PRO80920 Figure 953: PRO80906 Figure 988: DNA324236, NM_022453, Figure 954A-C: DNA324216, XM_055254, gen.NM_022453 gen.XM_055254 Figure 989: PRO80921 Figure 955: DNA324217, NM_004044, Figure 990: DNA324237, NM_032726, gen.NM_004044 gen.NM_032726 Figure 956: PRO80908 Figure 991: PRO70675 Figure 957: DNA324218, XM_114298, Figure 992: DNA324238, XM_010866, gen.XM_114298 gen.XM_010866 Figure 958: DNA324219, NM_021141, Figure 993: DNA324239, XM_087166, gen.NM_021141 gen.XM_087166 Figure 959: PRO59313 Figure 994: DNA254204, NM_001087, Figure 960A-B: DNA324220, XM_098048, gen.NM_001087 gen.XM_098048 Figure 995: PRO49316 Figure 961: PRO80910 Figure 996: DNA324240, NM_005731, Figure 962: DNA324221, XM_098047, gen.NM_005731 gen.XM_098047 Figure 997: PRO80924 Figure 963: PRO80911 Figure 998: DNA189697, NM_004846, Figure 964: DNA324222, XM_002636, gen.NM_004846 gen.XM_002636 Figure 999: PRO23123 Figure 965: DNA324223, XM_087181, Figure 1000: DNA324241, NM_025202, gen.XM_087181 gen.NM_025202 Figure 966: DNA324224, NM_000998, Figure 1001: PRO80925 gen.NM_000998 Figure 1002: DNA324242, XM_115825, Figure 967: PRO10498 gen.XM_115825 Figure 968: DNA324225, XM_059422, Figure 1003: PRO80926 Figure 1004: DNA324243, XM_010858, gen.XM_059422 Figure 969: PRO9984 gen.XM_010858 Figure 970: DNA324226, XM_092545, Figure 1005: PRO80927 gen.XM_092545 Figure 1006: DNA324244, XM .002540, Figure 971: DNA324227, XM_059461, gen.XM_002540 gen.XM_059461 Figure 1007: DNA324245, XM_048690, Figure 972: PRO80915 gen.XM_048690 Figure 973: DNA324228, NM_018674, Figure 1008: PRO80929 gen.NM_018674 Figure 1009: DNA324246, NM_030926, Figure 974: PRO80916 gen.NM_030926 Figure 1010: PRO80930 Figure 975: DNA324229, XM_050962, gen.XM_050962 Figure 1011: DNA324247, XM_087218, Figure 976: PRO80917 gen.XM_087218 Figure 977: DNA194827, NM_012100, Figure 1012: DNA324248, NM_004509, gen.NM_012100 gen.NM_004509 Figure 978: PRO24091 Figure 1013: PRO80932

Figure 1014: DNA324249, NM_004510,	Figure 1049: DNA324269, NM_006354,
gen.NM_004510	gen.NM_006354
Figure 1015: PRO80933	Figure 1050: PRO80952
Figure 1016: DNA324250, NM_080424,	Figure 1051: DNA324270, NM_133480,
gen.NM_080424	gen.NM_133480
Figure 1017: PRO80934	Figure 1052: PRO80953
Figure 1018: DNA324251, NM_018410,	Figure 1053: DNA324271, NM_133481,
gen.NM_018410	gen.NM_133481
Figure 1019: PRO80935	Figure 1054: PRO80954
Figure 1020: DNA324252, NM_017974,	Figure 1055: DNA324272, NM_005718,
gen.NM_017974	gen.NM_005718
Figure 1021: PRO80936	Figure 1056: PRO80955
Figure 1022A-B: DNA324253, XM_096169,	Figure 1057: DNA324273, NM_015644,
gen.XM_096169	gen.NM_015644
Figure 1023: PRO80937	Figure 1058: PRO80956
Figure 1024: DNA150884, NM_005855,	Figure 1059: DNA324274, XM_059561,
gen.NM_005855	gen.XM_059561
Figure 1025: PRO12520	Figure 1060: DNA324275, XM_052310,
Figure 1026A-B: DNA324254, NM_004735,	gen.XM_052310
gen.NM_004735	Figure 1061: PRO80958
Figure 1027: PRO80938	Figure 1062: DNA269910, NM _006395,
Figure 1028A-C: DNA324255, XM_030203,	gen.NM_006395
gen.XM_030203	Figure 1063: PRO58308
Figure 1029: DNA324256, XM_059372,	Figure 1064: DNA324276, NM_000994,
gen.XM_059372	gen.NM_000994
Figure 1030: DNA324257, NM_002712,	Figure 1065: PRO80959
gen.NM_002712	Figure 1066: DNA151017, NM_004844,
Figure 1031: PRO80941	gen.NM_004844
Figure 1032A-B: DNA324258, XM_042326,	Figure 1067: PRO12841
gen.XM_042326	Figure 1068: DNA324277, XM_059557,
Figure 1033: PRO80942	gen.XM_059557
Figure 1034: DNA324259, NM_004404,	Figure 1069: PRO80960
gen.NM_004404	Figure 1070A-B: DNA324278, XM_042860,
Figure 1035: PRO80943	gen.XM_042860
Figure 1036: DNA324260, XM_002742,	Figure 1071: PRO80961
gen.XM_002742	Figure 1072: DNA324279, XM_042841,
Figure 1037: DNA324261, NM_138483,	gen.XM_042841
gen.NM_138483	Figure 1073: PRO80962
Figure 1038: PRO80945	Figure 1074: DNA324280, XM_053712,
Figure 1039: DNA324262, XM_115706,	gen.XM_053712
gen.XM_115706	Figure 1075: DNA324281, XM_087284,
Figure 1040: DNA324263, XM_115722,	gen.XM_087284
gen.XM_115722	Figure 1076: DNA324282, NM_002948,
Figure 1041: DNA324264, XM_084141,	gen.NM_002948
gen.XM_084141	Figure 1077: PRO6360
Figure 1042: DNA324265, XM_005086,	Figure 1078: DNA324283, XM_053323,
gen.XM_005086	gen.XM_053323
Figure 1043: DNA324266, NM_015453,	Figure 1079A-B: DNA324284, NM_001068,
gen.NM_015453	gen.NM_001068
Figure 1044: PRO80949	Figure 1080: PRO80966
Figure 1045: DNA324267, NM_022485,	Figure 1081: DNA252367, NM_017801,
gen.NM_022485	gen.NM_017801
Figure 1046: PRO80950	Figure 1082: PRO48357
Figure 1047A-B: DNA324268, XM_054520,	Figure 1083: DNA324285, XM_093624,
gen.XM_054520	gen.XM_093624
Figure 1048: PRO80951	Figure 1084: PRO80967
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gen.XM_087588 Figure 1085: DNA324286, XM_046401, gen.XM_046401 Figure 1121: DNA324302, XM_166011, gen.XM_166011 Figure 1086: DNA324287, NM_022461, Figure 1122A-B: DNA324303, XM_114364, gen.NM_022461 Figure 1087: PRO80969 gen.XM_114364 Figure 1123A-B: DNA324304, XM_033294, Figure 1088: DNA324288, XM_113410, gen.XM_113410 gen.XM_033294 Figure 1089: DNA88100, NM_000404, Figure 1124: PRO80983 gen.NM_000404 Figure 1125: DNA324305, NM_138614, Figure 1090: PRO2172 gen.NM_138614 Figure 1091: DNA324289, XM .091076, Figure 1126: PRO80984 Figure 1127: DNA324306, XM_002899, gen.XM_091076 Figure 1092: PRO80970 gen.XM_002899 Figure 1128: DNA225910, NM_004345, Figure 1093A-B: DNA271187, NM_005109, gen.NM_004345 gen.NM_005109 Figure 1129: PRO36373 Figure 1094: PRO59504 Figure 1095: DNA324290, NM_002468, Figure 1130: DNA324307, XM_010953, gen.NM_002468 gen.XM_010953 Figure 1096: PRO36735 Figure 1131: DNA324308, XM_051518, gen.XM_051518 Figure 1097: DNA269930, NM_001607, Figure 1132A-D: DNA324309, NM_001407, gen.NM_001607 gen.NM_001407 Figure 1098: PRO58328 Figure 1133: PRO50095 Figure 1099: DNA270401, NM _003149, Figure 1134: DNA324310, NM_003365, gen.NM_003149 gen.NM_003365 Figure 1100: PRO58784 Figure 1135: PRO80988 Figure 1101: DNA324291, XM_087370, Figure 1136: DNA324311, XM_003245, gen.XM_087370 gen.XM_003245 Figure 1102: PRO80971 Figure 1137: DNA324312, XM_047561, Figure 1103: DNA324292, XM_098158, gen.XM_098158 gen.XM_047561 Figure 1104: PRO80972 Figure 1138: PRO80990 Figure 1139: DNA324313, XM_116853, Figure 1105: DNA324293, XM_017364, gen.XM_116853 gen.XM_017364 Figure 1140A-B: DNA324314, XM_113405, Figure 1106: DNA324294, XM_087349, gen.XM_113405 gen.XM_087349 Figure 1141: DNA324315, XM_114323, Figure 1107: PRO80974 gen.XM_114323 Figure 1108: DNA226547, NM _002295, Figure 1142: PRO80993 gen.NM_002295 Figure 1143: DNA324316, XM_002828, Figure 1109: PRO37010 gen.XM_002828 Figure 1110: DNA324295, NM _003973, gen.NM_003973 Figure 1144: PRO80994 Figure 1111: PRO80975 Figure 1145: DNA150976, NM _022171, Figure 1112: DNA324296, XM_030417, gen.NM_022171 gen.XM_030417 Figure 1146: PRO12565 Figure 1147: DNA324317, XM_041507, Figure 1113: DNA324297, NM_020347, gen.XM_041507 gen.NM_020347 Figure 1148: PRO71103 Figure 1114: PRO80977 Figure 1149: DNA103505, NM_004636, Figure 1115: DNA324298, XM _087346, gen.NM_004636 gen.XM_087346 Figure 1150: PRO4832 Figure 1116: PRO80978 Figure 1117: DNA324299, XM_096198, Figure 1151: DNA324318, NM_006764, gen.XM_096198 gen.NM_006764 Figure 1152: PRO80995 Figure 1118: PRO80979 Figure 1153: DNA150562, NM_007275, Figure 1119: DNA324300, XM_003222, gen.NM_007275 gen.XM_003222 Figure 1154: PRO12779 Figure 1120: DNA324301, XM_087588,

Figure 1191: PRO81010 Figure 1155: DNA254582, NM_004635, Figure 1192: DNA324336, XM_166015, gen.NM_004635 gen.XM_166015 Figure 1156: PRO49685 Figure 1193: DNA324337, XM_113395, Figure 1157: DNA324319, NM_052859, gen.XM_113395 gen.NM_052859 Figure 1194: PRO81012 Figure 1158: PRO80996 Figure 1195: DNA269730, NM_014814, Figure 1159: DNA324320, NM_001064, gen.NM_014814 gen.NM_001064 Figure 1196: PRO58140 Figure 1160: PRO80997 Figure 1197: DNA324338, XM_036938, Figure 1161: DNA324321, XM_041211, gen.XM_036938 gen.XM_041211 Figure 1198: DNA324339, XM_029369, Figure 1162: DNA324322, XM_003213, gen.XM_029369 gen.XM_003213 Figure 1199: DNA324340, XM_076414, Figure 1163A-C: DNA324323, XM_037423, gen.XM_076414 gen.XM_037423 Figure 1200: PRO81015 Figure 1164: PRO80999 Figure 1201: DNA324341, XM_093546, Figure 1165A-B: DNA227307, NM_007184, gen.XM_093546 gen.NM_007184 Figure 1202: DNA324342, XM_113409, Figure 1166: PRO37770 gen.XM_113409 Figure 1167: DNA324324, NM_000688, Figure 1203: DNA324343, XM_087268, gen.NM_000688 gen.XM_087268 Figure 1168: PRO81000 Figure 1204: DNA324344, XM_116071, Figure 1169: DNA324325, XM_067715, gen.XM_116071 gen.XM_067715 Figure 1205: DNA324345, XM_116072, Figure 1170: DNA324326, NM_000992, gen.XM_116072 gen.NM_000992 Figure 1206: DNA324346, NM_000986, Figure 1171: PRO62153 gen.NM_000986 Figure 1172: DNA324327, NM_000666, Figure 1207: PRO10602 gen.NM_000666 Figure 1208: DNA324347, XM_015462, Figure 1173: PRO81002 Figure 1174: DNA324328, NM .032750, gen.XM_015462 Figure 1209: DNA324348, XM_167366, gen.NM_032750 gen.XM_167366 Figure 1175: PRO81003 Figure 1210: PRO81022 Figure 1176: DNA324329, NM_033008, Figure 1211: DNA324349, XM .. 087331, gen.NM_033008 gen.XM_087331 Figure 1177: PRO81004 Figure 1212: PRO81023 Figure 1178: DNA324330, NM_033010, Figure 1213: DNA324350, XM_039952, gen.NM_033010 gen.XM_039952 Figure 1179: PRO81005 Figure 1214: DNA324351, XM_045290, Figure 1180: DNA324331, NM_020418, gen.XM_045290 gen.NM_020418 Figure 1215: PRO81025 Figure 1181: PRO81006 Figure 1216A-B: DNA324352, NM .007085, Figure 1182: DNA273919, NM_004704, gen.NM_007085 gen.NM_004704 Figure 1217: PRO2077 Figure 1183: PRO61870 Figure 1184A-B: DNA324332, XM_087448, Figure 1218: DNA324353, NM _004547, gen.NM_004547 gen.XM_087448 Figure 1219: PRO81026 Figure 1185: PRO81007 Figure 1220: DNA324354, XM_027161, Figure 1186: DNA324333, XM_002855, gen.XM_002855 gen.XM_027161 Figure 1221A-B: DNA324355, XM .032269, Figure 1187: DNA324334, XM_002854, gen.XM_032269 gen.XM_002854 Figure 1222: PRO81028 Figure 1188: DNA0, NM_002854, gen.NM_002854 Figure 1223: DNA88547, NM_006810, Figure 1189: PRO gen.NM_006810 Figure 1190: DNA324335, XM_096195, Figure 1224: PRO2837 gen.XM_096195

Figure 1225: DNA324356, XM_114301, Figure 1259: PRO81046 Figure 1260: DNA324378, NM_000532, gen.XM_114301 Figure 1226: PRO81029 gen.NM_000532 Figure 1227: DNA324357, XM_098173, Figure 1261: PRO81047 Figure 1262: DNA324379, XM_036118, gen.XM_098173 Figure 1228: PRO81030 gen.XM_036118 Figure 1229: DNA324358, XM_042618, Figure 1263: DNA324380, XM_084123, gen.XM_084123 gen.XM_042618 Figure 1230: PRO81031 Figure 1264: DNA324381, XM_018149, gen.XM_018149 Figure 1231: DNA324359, XM_084129, Figure 1265: DNA324382, XM_087342, gen.XM_084129 gen.XM_087342 Figure 1232: DNA324360, XM_098154, Figure 1266: DNA324383, XM_059516, gen.XM_098154 Figure 1233: PRO81033 gen.XM_059516 Figure 1234: DNA324361, XM_050552, Figure 1267: DNA324384, XM_087341, gen.XM_087341 gen.XM_050552 Figure 1235: DNA324362, NM_032343, Figure 1268: DNA324385, XM_165451, gen.XM_165451 gen.NM_032343 Figure 1269: PRO81053 Figure 1236: PRO81034 Figure 1237: DNA324363, XM_051264, Figure 1270: DNA269858, NM _004766, gen.NM_004766 gen.XM_051264 Figure 1271: PRO58259 Figure 1238A-B: DNA324364, NM_013336, Figure 1272: DNA324386, NM_030921, gen.NM_013336 Figure 1239: PRO1314 gen.NM_030921 Figure 1240: DNA324365, XM_067264, Figure 1273: PRO51109 Figure 1274: DNA324387, XM_002859, gen.XM_067264 Figure 1241: PRO81036 gen.XM_002859 Figure 1242: DNA324366, XM_114309, Figure 1275: DNA324388, XM_166014, gen.XM_114309 gen.XM_166014 Figure 1243: DNA324367, XM_084111, Figure 1276: DNA324389, NM_013363, gen.XM_084111 gen.NM_013363 Figure 1244: DNA324368, XM_113397, Figure 1277: PRO287 gen.XM_113397 Figure 1278: DNA324390, XM_058267, gen.XM_058267 Figure 1245: DNA324369, XM_098111, Figure 1279: PRO81056 gen.XM_098111 Figure 1280A-B: DNA324391, NM_032383, Figure 1246: DNA324370, NM_004637, gen.NM_004637 gen.NM_032383 Figure 1281: PRO81057 Figure 1247: PRO81040 Figure 1282: DNA324392, NM_015472, Figure 1248: DNA324371, NM_020701, gen.NM_020701 gen.NM_015472 Figure 1249: PRO81041 Figure 1283: PRO81058 Figure 1250: DNA324372, NM_003418, Figure 1284: DNA324393, NM_014445, gen.NM_003418 gen.NM_014445 Figure 1251: PRO81042 Figure 1285: PRO11048 Figure 1252: DNA324373, XM_059583, Figure 1286: DNA324394, XM_042168, gen.XM_042168 gen.XM_059583 Figure 1287: PRO81059 Figure 1253: PRO81043 Figure 1254: DNA324374, XM_113417, Figure 1288A-B: DNA324395, XM_114356, gen.XM_113417 gen.XM_114356 Figure 1289: DNA324396, XM_105236, Figure 1255: DNA324375, XM_093487, gen.XM_093487 gen.XM_105236 Figure 1290: DNA324397, XM_010978, Figure 1256A-B: DNA324376, XM_030812, gen.XM_030812 gen.XM_010978 Figure 1291: DNA324398, XM_017356, Figure 1257: PRO58177 gen.XM_017356 Figure 1258A-B: DNA324377, XM_039805, Figure 1292A-B: DNA324399, XM_039796, gen.XM_039805

gen.XM_039796 Figure 1327: DNA89239, NM_000893, Figure 1293: PRO81064 gen.NM_000893 Figure 1294: DNA324400, XM_016334, Figure 1328: PRO2906 gen.XM_016334 Figure 1329: DNA324420, XM_113422, Figure 1295: DNA324401, XM_116058, gen.XM_113422 gen.XM_116058 Figure 1330: DNA225592, NM_001622, Figure 1296: DNA324402, XM_113408, gen.NM_001622 gen.XM_113408 Figure 1331: PRO36055 Figure 1297: DNA324403, NM _002492, Figure 1332: DNA324421, XM_005180, gen.NM_002492 gen.XM_005180 Figure 1298: PRO81068 Figure 1333: DNA324422, XM_087392, Figure 1299: DNA324404, XM_037381, gen.XM_087392 gen.XM_037381 Figure 1334: PRO81086 Figure 1300: DNA324405, XM_037377, Figure 1335A-B: DNA272605, NM_003722, gen.XM_037377 gen.NM_003722 Figure 1301: PRO69681 Figure 1336: PRO60741 Figure 1302A-B: DNA324406, XM_087254, Figure 1337: DNA324423, XM_117311, gen.XM_087254 gen.XM_117311 Figure 1303: PRO81070 Figure 1338: DNA324424, XM_116034, Figure 1304: DNA324407, XM_037600, gen.XM_116034 gen.XM_037600 Figure 1339: PRO81088 Figure 1305: PRO81071 Figure 1340A-B: DNA324425, XM_084110, Figure 1306: DNA324408, NM_018023, gen.XM_084110 gen.NM_018023 Figure 1341: DNA324426, XM_038243, Figure 1307: PRO81072 gen.XM_038243 Figure 1308: DNA324409, XM_093423, Figure 1342: PRO81090 gen.XM_093423 Figure 1343: DNA324427, XM_087359, Figure 1309: PRO81073 gen.XM_087359 Figure 1310: DNA324410, XM_029136, Figure 1344: DNA324428, XM_114328, gen.XM_029136 gen.XM_114328 Figure 1311: PRO81074 Figure 1345: DNA324429, XM_098109, Figure 1312: DNA324411, XM_087322, gen.XM_098109 gen.XM_087322 Figure 1346: PRO81093 Figure 1313A-B: DNA324412, XM_029132, Figure 1347: DNA324430, XM_087410, gen.XM_087410 gen.XM_029132 Figure 1314A-B: DNA324413, XM_029104, Figure 1348: DNA324431, NM_033316, gen.XM_029104 gen.NM_033316 Figure 1315: DNA324414, XM_084120, Figure 1349: PRO81095 gen.XM_084120 Figure 1350: DNA324432, XM_166017, Figure 1316: DNA254620, NM_005787, gen.XM_166017 Figure 1351: PRO81096 gen.NM_005787 Figure 1317: PRO49722 Figure 1352: DNA79129, NM _001647, Figure 1318: DNA324415, NM_032331, gen.NM_001647 gen.NM_032331 Figure 1353: PRO2551 Figure 1319: PRO81079 Figure 1354: DNA324433, NM_032288, Figure 1320: DNA324416, XM_011074, gen.NM_032288 gen.XM_011074 Figure 1355: PRO81097 Figure 1321: PRO81080 Figure 1356: DNA324434, XM_086228, Figure 1322: DNA324417, XM_087295, gen.XM_086228 Figure 1357: PRO81098 gen.XM_087295 Figure 1323: DNA324418, XM_087289, Figure 1358: DNA324435, XM_087278, gen.XM_087289 gen.XM_087278 Figure 1324: PRO81082 Figure 1359: DNA324436, XM_018523, Figure 1325: DNA324419, XM_105658, gen.XM_018523 gen.XM_105658 Figure 1360: DNA324437, XM_087297, Figure 1326: PRO81083 gen.XM_087297

Figure 1397: PRO60542 Figure 1361: DNA324438, XM_002255, Figure 1398A-B: DNA324455, XM_052626, gen.XM_002255 gen.XM_052626 Figure 1362: PRO81102 Figure 1363: DNA324439, XM_053122, Figure 1399: PRO81118 Figure 1400: DNA324456, NM_016930, gen.XM_053122 gen.NM_016930 Figure 1364: DNA324440, XM_042695, Figure 1401: PRO81119 gen.XM_042695 Figure 1402: DNA324457, XM _035824, Figure 1365: DNA324441, XM_011160, gen.XM_035824 gen.XM_011160 Figure 1403: PRO81120 Figure 1366: DNA324442, NM_007100, Figure 1404: DNA324458, NM_033296, gen.NM_007100 gen.NM_033296 Figure 1367: PRO81106 Figure 1405: PRO81121 Figure 1368: DNA139747, NM_002477, gen.NM_002477 Figure 1406: DNA324459, NM_138699, gen.NM_138699 Figure 1369: PRO9785 Figure 1407: PRO81122 Figure 1370: DNA253804, NM_032219, Figure 1408: DNA324460, XM_116285, gen.NM_032219 gen.XM_116285 Figure 1371: PRO49209 Figure 1409: PRO81123 Figure 1372: DNA324443, NM_138385, gen.NM_138385 Figure 1410: DNA324461, XM_041221, gen.XM_041221 Figure 1373: PRO81107 Figure 1374: DNA324444, NM_006342, Figure 1411: PRO81124 Figure 1412: DNA324462, XM_117351, gen.NM_006342 gen.XM_117351 Figure 1375: PRO81108 Figure 1413: DNA324463, XM_039165, Figure 1376A-C: DNA324445, NM_133330, gen.XM_039165 gen.NM_133330 Figure 1414: DNA324464, NM_025205, Figure 1377: PRO81109 Figure 1378A-C: DNA324446, NM_014919, gen.NM_025205 Figure 1415: PRO81127 gen.NM_014919 Figure 1379: PRO81110 Figure 1416: DNA324465, XM_039173, gen.XM_039173 Figure 1380A-C: DNA324447, NM_133332, Figure 1417: DNA324466, XM_039176, gen.NM_133332 gen.XM_039176 Figure 1381: PRO81111 Figure 1418: DNA324467, XM_087583, Figure 1382: DNA324448, NM_005663, gen.XM_087583 gen.NM_005663 Figure 1419: DNA324468, NM_017491, Figure 1383: PRO81112 Figure 1384A-B: DNA324449, XM _098248, gen.NM_017491 gen.XM_098248 Figure 1420: PRO12077 Figure 1421: DNA324469, NM_005112, Figure 1385: PRO81113 gen.NM_005112 Figure 1386: DNA270615, NM_002938, Figure 1422: PRO81131 gen.NM_002938 Figure 1423: DNA324470, XM_011129, Figure 1387: PRO58986 Figure 1388A-B: DNA324450, NM_014190, gen.XM_011129 Figure 1424A-B: DNA324471, XM _052530, gen.NM_014190 gen.XM_052530 Figure 1389: PRO81114 Figure 1390A-B: DNA324451, NM_014189, Figure 1425: DNA324472, NM_000661, gen.NM_000661 gen.NM_014189 Figure 1426: PRO81134 Figure 1391: PRO81115 Figure 1427A-B: DNA324473, NM_002913, Figure 1392: DNA324452, XM_035572, gen.NM_002913 gen.XM_035572 Figure 1428: PRO81135 Figure 1393: PRO81116 Figure 1429A-B: DNA324474, XM_047477, Figure 1394A-B: DNA324453, NM_014556, gen.NM_014556 gen.XM_047477 Figure 1430: DNA324475, NM_004181, Figure 1395: PRO81117 Figure 1396: DNA324454, NM_001313, gen.NM_004181 Figure 1431: PRO81137 gen.NM_001313

Figure 1432: DNA324476, XM_003435,	gen.XM_096203
gen.XM_003435	Figure 1465: DNA324498, XM_084158,
Figure 1433: DNA324478, XM_010941,	gen.XM_084158
gen.XM_010941	Figure 1466: DNA324499, XM_034710,
Figure 1434: DNA324479, XM_059593,	gen.XM_034710
gen.XM_059593	Figure 1467: PRO81156
Figure 1435: DNA324480, NM_001553,	Figure 1468: DNA324500, XM_034713,
gen.NM_001553	gen.XM_034713
Figure 1436: PRO81141	Figure 1469: DNA324501, XM_059633,
Figure 1437: DNA257511, NM_032313,	gen.XM_059633
gen.NM_032313	Figure 1470: DNA324502, XM_114426,
Figure 1438: PRO52083	gen.XM_114426
Figure 1439: DNA324481, XM_071623,	Figure 1471: DNA324503, XM_056957,
gen.XM_071623	gen.XM_056957
Figure 1440A-B: DNA324482, XM_036002,	Figure 1472: DNA324504, XM_088472,
gen.XM_036002	gen.XM_088472
Figure 1441: DNA324483, XM_058927,	Figure 1473: DNA324505, XM_114424,
gen.XM_058927	gen.XM_114424
Figure 1442: DNA324484, XM_059628,	Figure 1474A-B: DNA324506, XM_042301,
gen.XM_059628	gen.XM_042301
Figure 1443: DNA324485, XM_046057,	Figure 1475: PRO81163
gen.XM_046057	Figure 1476: DNA324507, XM_017925,
Figure 1444: PRO81146	gen.XM_017925
Figure 1445: DNA324486, XM 031320,	Figure 1477: DNA324508, XM_052336,
gen.XM_031320	gen.XM_052336
Figure 1446: DNA225919, NM _001134,	Figure 1478: DNA324509, NM_002106,
gen.NM.001134	gen.NM_002106
Figure 1447: PRO36382	Figure 1479: PRO10297
Figure 1448A-B: DNA324487, XM_003511,	Figure 1480: DNA324510, XM_085068,
gen.XM_003511	gen.XM_085068
Figure 1449: DNA324488, NM .006835,	Figure 1481: PRO81166
gen.NM_006835	Figure 1482: DNA324511, XM_165473,
Figure 1450: PRO4605	gen.XM_165473
Figure 1451: DNA324489, XM _003305,	Figure 1483: DNA324512, XM_087514,
gen.XM_003305	gen.XM_087514
Figure 1452: DNA324490, XM_113425,	Figure 1484: DNA324513, XM_116247,
gen.XM_113425	gen.XM_116247
Figure 1453: DNA324491, XM_001389,	Figure 1485: DNA324514, NM_002358,
gen.XM_001389	gen.NM_002358
Figure 1454: PRO81148	Figure 1486: PRO81169
Figure 1455: DNA324492, XM_087527,	Figure 1487: DNA324515, XM_050200,
gen.XM_087527	gen.XM_050200
Figure 1456: DNA324493, XM_035986,	Figure 1488: PRO81170
gen.XM_035986	Figure 1489: DNA225584, NM_001154,
Figure 1457A-B: DNA324494, NM_014933,	gen.NM_001154
gen.NM_014933	Figure 1490: PRO36047
Figure 1458: PRO81150	Figure 1491: DNA324516, NM_024900,
Figure 1459: DNA290585, NM_000582,	gen.NM_024900
gen.NM_000582	Figure 1492: PRO81171
Figure 1460: PRO70536	Figure 1493: DNA324517, XM_040752,
Figure 1461: DNA324495, XM_055551,	gen.XM_040752
gen.XM_055551	Figure 1494: DNA324518, NM_002413,
Figure 1462: PRO81151	gen.NM_002413
Figure 1463: DNA324496, XM_087498,	Figure 1495: PRO60956
gen.XM_087498	Figure 1496: DNA324519, XM_114401,
Figure 1464: DNA324497, XM_096203,	gen.XM_114401
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Figure 1497: DNA324520, XM_068164,	Figure 1532: DNA324538, XM_116204,
gen.XM_068164	gen.XM_116204
Figure 1498: PRO81174	Figure 1533: DNA324539, XM_116205,
Figure 1499: DNA324521, XM_060067,	gen.XM_116205
gen.XM_060067	Figure 1534: DNA324540, XM_098405,
Figure 1500: DNA324522, XM_003555,	gen.XM_098405
gen.XM_003555	Figure 1535: DNA324541, XM _052313,
Figure 1501: PRO81176	gen.XM_052313
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Figure 1502: DNA324523, XM_034321,	Figure 1536: PRO81195
gen.XM_034321	Figure 1537: DNA324542, XM_087659,
Figure 1503: PRO81177	gen.XM_087659
Figure 1504: DNA324524, NM_006439,	Figure 1538: PRO81196
gen.NM_006439	Figure 1539: DNA324543, XM_029096,
Figure 1505: PRO81178	gen.XM_029096
Figure 1506: DNA324525, NM_001006,	Figure 1540: DNA324544, XM_003825,
gen.NM_001006	gen.XM_003825
Figure 1507: PRO81179	Figure 1541: DNA324545, XM_057994,
Figure 1508: DNA227575, NM_005141,	gen.XM_057994
gen.NM_005141	Figure 1542: PRO81199
Figure 1509: PRO38038	Figure 1543: DNA324546, XM_087686,
Figure 1510: DNA324526, XM_114368,	gen.XM_087686
gen.XM_114368	Figure 1544: DNA324547, XM_017641,
Figure 1511A-B: DNA225920, NM_000508,	gen.XM_017641
gen.NM_000508	Figure 1545: DNA324548, NM_030782,
Figure 1512: PRO36383	gen.NM_030782
Figure 1513: DNA324527, NM_021871,	Figure 1546: PRO81202
gen.NM_021871	Figure 1547: DNA324549, XM_084168,
Figure 1514: PRO81181	gen.XM_084168
Figure 1515: DNA225921, NM_000509,	Figure 1548: DNA324550, XM_057492,
gen.NM_000509	gen.XM_057492
Figure 1516: PRO36384	Figure 1549: DNA324551, XM_087597,
Figure 1517: DNA324528, NM_021870,	gen.XM_087597
gen.NM_021870	Figure 1550: DNA324552, XM_087601,
Figure 1518: PRO81182	gen.XM_087601
Figure 1519: DNA324529, XM_059623,	Figure 1551: DNA324554, XM_087599,
gen.XM_059623	gen.XM_087599
Figure 1520: DNA324530, XM_106246,	Figure 1552: DNA324555, XM_114435,
gen.XM_106246	gen.XM_114435
Figure 1521: PRO81184	Figure 1553: DNA324556, XM_087600,
Figure 1522: DNA324531, NM_002129,	gen.XM_087600
gen.NM_002129	Figure 1554: DNA324557, XM_016170,
Figure 1523: PRO81185	gen.XM_016170
Figure 1524: DNA324532, XM_040321,	Figure 1555: DNA324558, XM_114434,
gen.XM_040321	gen.XM_114434
Figure 1525: DNA324533, XM_015563,	Figure 1556: DNA324559, XM_113452,
gen.XM_015563	gen.XM_113452
Figure 1526: DNA324534, NM_024748,	Figure 1557: DNA324560, XM_071580,
gen.NM_024748	gen.XM_071580
Figure 1527: PRO81188	Figure 1558: PRO81213
Figure 1528: DNA324535, XM_165470,	Figure 1559: DNA324561, XM_087713,
gen.XM_165470	gen.XM_087713
Figure 1529: PRO81189	Figure 1560: PRO81214
Figure 1530A-E: DNA324536, XM_003477,	Figure 1561: DNA324562, XM_094440,
gen.XM_003477	gen.XM_094440
Figure 1531: DNA324537, XM_165465,	Figure 1562: DNA324563, XM_106739,
gen.XM_165465	gen.XM_106739
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Figure 1597: DNA324584, XM_087610, Figure 1563: PRO81216 gen.XM_087610 Figure 1564: DNA324564, XM_087614, Figure 1598: DNA288259, NM_031966, gen.XM_087614 Figure 1565: DNA324565, XM_004009, gen.NM_031966 gen.XM_004009 Figure 1599: PRO4676 Figure 1600: DNA324585, XM_042025, Figure 1566: PRO81219 Figure 1567: DNA324566, XM_114437, gen.XM_042025 gen.XM_114437 Figure 1601: PRO81238 Figure 1568: DNA324567, XM_043771, Figure 1602: DNA324586, NM_005713, gen.XM_043771 gen.NM_005713 Figure 1603: PRO81239 Figure 1569: PRO81221 Figure 1604: DNA324587, XM .059709, Figure 1570: DNA324568, NM _000997, gen.NM_000997 gen.XM_059709 Figure 1605: PRO81240 Figure 1571: PRO11077 Figure 1606: DNA324588, XM_116447, Figure 1572: DNA324569, XM_003869, gen.XM_116447 gen.XM_003869 Figure 1573: DNA227173, NM _001465, Figure 1607: PRO81241 gen.NM_001465 Figure 1608: DNA324589, XM_037260, gen.XM_037260 Figure 1574: PRO37636 Figure 1609: DNA324590, XM_098351, Figure 1575: DNA324570, NM_018034, gen.XM_098351 gen.NM_018034 Figure 1610: DNA324591, XM_098354, Figure 1576: PRO81223 gen.XM_098354 Figure 1577: DNA324571, NM_032637, Figure 1611: DNA324592, XM .098352, gen.NM_032637 Figure 1578: PRO81224 gen.XM_098352 Figure 1579: DNA324572, NM_005983, Figure 1612: DNA324593, XM_166037, gen.NM_005983 gen.XM_166037 Figure 1613: PRO81246 Figure 1580: PRO81225 Figure 1614: DNA324594, XM_041694, Figure 1581A-B: DNA324573, XM .003896, gen.XM_041694 gen.XM_003896 Figure 1615: DNA324595, XM_165488, Figure 1582: DNA287282, NM .. 002130, gen.XM_165488 gen.NM_002130 Figure 1583: PRO69554 Figure 1616: PRO81248 Figure 1617: DNA324596, XM_059669, Figure 1584: DNA324574, XM_114442, gen.XM_059669 gen.XM_114442 Figure 1618: PRO81249 Figure 1585: PRO81227 Figure 1619: DNA324597, XM _027964, Figure 1586: DNA324575, XM_114439, gen.XM_027964 gen.XM_114439 Figure 1620: PRO81250 Figure 1587: DNA324576, XM_114440, Figure 1621: DNA324598, XM_088020, gen.XM_114440 Figure 1588A-B: DNA324577, XM_032902, gen.XM_088020 Figure 1622: DNA324599, XM_117387, gen.XM_032902 gen.XM_117387 Figure 1589: PRO81230 Figure 1623: DNA324600, XM_114469, Figure 1590: DNA324578, XM_032895, gen.XM_114469 gen.XM_032895 Figure 1624: DNA324601, NM..001207, Figure 1591: DNA324579, XM_084179, gen.NM_001207 gen.XM_084179 Figure 1625: PRO22771 Figure 1592: DNA324580, XM_041712, gen.XM_041712 Figure 1626A-B: DNA324602, XM_032553, gen.XM_032553 Figure 1593: DNA324581, XM_116439, Figure 1627: DNA254147, NM_000521, gen.XM_116439 gen.NM_000521 Figure 1594: PRO81234 Figure 1628: PRO49262 Figure 1595: DNA324582, XM_087611, Figure 1629: DNA324603, NM_031482, gen.XM_087611 gen.NM_031482 Figure 1596: DNA324583, XM_059653, gen.XM_059653 Figure 1630: PRO81254

Figure 1631: DNA324604, XM_087790, Figure 1666: DNA324622, XM_003830, gen.XM_087790 gen.XM_003830 Figure 1632: DNA324605, NM_001025, Figure 1667: PRO81269 gen.NM_001025 Figure 1668: DNA324623, XM_037002, Figure 1633: PRO10685 gen.XM_037002 Figure 1634: DNA324606, XM_098362, Figure 1669: DNA324624, XM_166026, gen.XM_098362 gen.XM_166026 Figure 1635: PRO81256 Figure 1670: DNA324625, XM_041059, Figure 1636: DNA324607, NM_003401, gen.XM_041059 gen.NM_003401 Figure 1671: DNA83020, NM_000358, Figure 1637: PRO70327 gen.NM_000358 Figure 1638: DNA290231, NM_022550, Figure 1672: PRO2561 gen.NM_022550 Figure 1673: DNA324626, NM_003687, Figure 1639: PRO70327 gen.NM_003687 Figure 1640: DNA324608, XM_017857, Figure 1674: PRO81272 gen.XM_017857 Figure 1675: DNA324627, XM_034862, Figure 1641: DNA324609, XM_117398, gen.XM_034862 gen.XM_117398 Figure 1676: PRO34544 Figure 1642A-B: DNA257253, NM_032280, Figure 1677: DNA103380, NM_003374, gen.NM_032280 gen.NM_003374 Figure 1643: PRO51851 Figure 1678: PRO4710 Figure 1644: DNA324610, XM_003771, Figure 1679: DNA324628, XM_017474, gen.XM_003771 gen.XM_017474 Figure 1645: PRO81259 Figure 1680: PRO63082 Figure 1646A-B: DNA269816, NM_002397, Figure 1681A-B: DNA324629, NM_014829, gen.NM_002397 gen.NM_014829 Figure 1647: PRO58219 Figure 1682: PRO81273 Figure 1683A-B: DNA324630, XM_114482, Figure 1648: DNA324611, XM_116427, gen.XM_116427 gen.XM_114482 Figure 1649: PRO81260 Figure 1684: PRO81274 Figure 1650: DNA324612, NM_004772, Figure 1685: DNA324631, NM_004893, gen.NM_004772 gen.NM-004893 Figure 1651: PRO81261 Figure 1686: PRO81275 Figure 1652: DNA324613, XM_016674, Figure 1687: DNA269809, NM_006805, gen.XM_016674 gen.NM_006805 Figure 1653: PRO81262 Figure 1688: PRO58213 Figure 1654: DNA324614, XM_113463, Figure 1689: DNA226872, NM_001964, gen.XM_113463 gen.NM_001964 Figure 1655: DNA324615, XM_034744, Figure 1690: PRO37335 gen.XM_034744 Figure 1691: DNA324632, XM_116307, Figure 1656: DNA324616, XM_087745, gen.XM_116307 gen.XM_087745 Figure 1692: PRO81276 Figure 1657: PRO81264 Figure 1693: DNA324633, NM .004134, Figure 1658: DNA324617, XM_018473, gen.NM_004134 gen.XM_018473 Figure 1694: PRO81277 Figure 1659: PRO81265 Figure 1695: DNA324634, XM_038221, Figure 1660: DNA324618, XM_087635, gen.XM_038221 gen.XM_087635 Figure 1696: PRO81278 Figure 1661: PRO81266 Figure 1697: DNA271931, NM_005754, Figure 1662: DNA324619, XM_087637, gen.NM_005754 gen.XM_087637 Figure 1698: PRO60207 Figure 1663: DNA324620, XM_166027, Figure 1699: DNA324635, XM_003841, gen.XM_166027 gen.XM_003841 Figure 1664: DNA324621, NM_014035, Figure 1700: DNA324636, XM_032759, gen.NM_014035 gen.XM_032759 Figure 1665: PRO1285 Figure 1701: DNA324637, XM_017591,

gen.XM_017591 gen.NM_018913 Figure 1702: DNA324638, NM_006058, Figure 1737: PRO81293 gen.NM_006058 Figure 1738A-B: DNA324656, NM_018914, Figure 1703: PRO81280 gen.NM_018914 Figure 1704: DNA324639, NM_002084, Figure 1739: PRO81294 Figure 1740A-B: DNA324657, NM_018915, gen.NM_002084 Figure 1705: PRO81281 gen.NM_018915 Figure 1706: DNA324640, NM_018047, Figure 1741: PRO36020 gen.NM_018047 Figure 1742A-B: DNA324658, NM_018916, Figure 1707: PRO81282 gen.NM_018916 Figure 1743: PRO81295 Figure 1708: DNA324641, NM_005617, gen.NM_005617 Figure 1744A-B: DNA324659, NM_018917, Figure 1709: PRO10849 gen.NM_018917 Figure 1745: PRO81296 Figure 1710: DNA324642, XM_003937, Figure 1746A-B: DNA324660, NM_018918, gen.XM_003937 Figure 1711: DNA324643, XM_087621, gen.NM_018918 gen.XM_087621 Figure 1747: PRO81297 Figure 1748A-B: DNA324661, NM_018919, Figure 1712A-B: DNA324644, XM_003789, gen.NM_018919 gen.XM_003789 Figure 1749: PRO81298 Figure 1713: DNA324645, XM_087652, Figure 1750A-B: DNA324662, NM_018920, gen.XM_087652 gen.NM_018920 Figure 1714: DNA324646, XM_068853, Figure 1751: PRO81299 gen.XM_068853 Figure 1715: PRO81286 Figure 1752A-B: DNA324663, NM_018921, Figure 1716: DNA324647, XM_116465, gen.NM_018921 Figure 1753: PRO81300 gen.XM_116465 Figure 1754A-B: DNA324664, NM_018922, Figure 1717: PRO81287 Figure 1718: DNA302020, NM _005573, gen.NM_018922 gen.NM_005573 Figure 1755: PRO81301 Figure 1756A-B: DNA324665, NM_018923, Figure 1719: PRO70993 Figure 1720: DNA324648, XM_113467, gen.NM_018923 gen.XM_113467 Figure 1757: PRO81302 Figure 1758A-B: DNA324666, NM_018924, Figure 1721: DNA271626, NM_014773, gen.NM_018924 gen.NM_014773 Figure 1759: PRO81303 Figure 1722: PRO59913 Figure 1760A-B: DNA324667, NM_018925, Figure 1723A-B: DNA324649, XM_056315, gen.NM_018925 gen.XM_056315 Figure 1761: PRO81304 Figure 1724: DNA324650, NM_024668, gen.NM_024668 Figure 1762A-B: DNA324668, NM_018926, Figure 1725: PRO81289 gen.NM_018926 Figure 1726: DNA324651, NM_080670, Figure 1763: PRO81305 gen.NM_080670 Figure 1764A-B: DNA324669, NM_018927, gen.NM_018927 Figure 1727: PRO81290 Figure 1765: PRO37091 Figure 1728A-B: DNA324652, NM_002588, Figure 1766A-B: DNA324670, NM_018928, gen.NM_002588 gen.NM_018928 Figure 1729: PRO81291 Figure 1767: PRO81306 Figure 1730A-B: DNA324653, NM_003735, Figure 1768A-B: DNA324671, NM_018929, gen.NM_003735 Figure 1731: PRO81292 gen.NM_018929 Figure 1769: PRO81307 Figure 1732A-B: DNA150679, NM_003736, Figure 1770A-B: DNA324672, NM_032088, gen.NM_003736 gen.NM_032088 Figure 1733: PRO12416 Figure 1771: PRO81308 Figure 1734A-B: DNA324654, NM_018912, Figure 1772A-B: DNA324673, NM_032092, gen.NM_018912 gen.NM_032092 Figure 1735: PRO36058

Figure 1736A-B: DNA324655, NM_018913,

Figure 1773: PRO81309

Figure 1774: DNA324674, NM_032403, Figure 1809: PRO81327 gen.NM_032403 Figure 1810: DNA324694, XM_116856, Figure 1775: PRO81310 gen.XM_116856 Figure 1776: DNA324675, NM_032402, Figure 1811: DNA324695, XM_003716, gen.NM_032402 gen.XM_003716 Figure 1777: PRO81311 Figure 1812: DNA227320, NM _003714, Figure 1778: DNA324676, XM_098387, gen.NM_003714 gen.XM_098387 Figure 1813: PRO37783 Figure 1779: DNA324677, NM_002109, Figure 1814: DNA324696, NM_032361, gen.NM_002109 gen.NM_032361 Figure 1780: PRO4908 Figure 1815: PRO81330 Figure 1781: DNA324678, XM_084180, Figure 1816: DNA324697, XM_087773, gen.XM_084180 gen.XM_087773 Figure 1817: DNA324698, XM_114457, Figure 1782: PRO81313 Figure 1783: DNA324679, XM_039975, gen.XM_114457 Figure 1818: DNA324699, XM_165483, gen.XM_039975 Figure 1784: PRO81314 gen.XM_165483 Figure 1785: DNA324680, NM_033551, Figure 1819: DNA324700, XM_114453, gen.XM_114453 gen.NM_033551 Figure 1820: DNA324701, XM_165484, Figure 1786: PRO81315 Figure 1787: DNA324681, NM_004821, gen.XM_165484 Figure 1821: DNA324702, XM_030771, gen.NM_004821 Figure 1788: PRO81316 gen.XM_030771 Figure 1789: DNA324682, XM_068395, Figure 1822: PRO19615 gen.XM_068395 Figure 1823: DNA324703, XM_030777, Figure 1790: PRO81317 gen.XM_030777 Figure 1791: DNA226418, NM_004060, Figure 1824: DNA324704, XM_030782, gen.NM_004060 gen.XM_030782 Figure 1825: PRO81336 Figure 1792: PRO36881 Figure 1793A-B: DNA324683, XM_056963, Figure 1826: DNA324705, NM_030567, gen.XM_056963 gen.NM_030567 Figure 1794: PRO81318 Figure 1827: PRO81337 Figure 1795: DNA324684, NM_004219, Figure 1828: DNA225909, NM_000505, gen.NM_004219 gen.NM_000505 Figure 1829: PRO36372 Figure 1796: PRO81319 Figure 1797: DNA324685, XM_094243, Figure 1830: DNA274206, NM .006816, gen.XM_094243 gen.NM_006816 Figure 1831: PRO62135 Figure 1798A-B: DNA324686, XM_047964, gen.XM_047964 Figure 1832: DNA324706, NM_031300, Figure 1799: DNA324687, XM_016345, gen.NM_031300 gen.XM_016345 Figure 1833: PRO81338 Figure 1800: DNA324688, NM_002887, Figure 1834: DNA324707, NM_013237, gen.NM_002887 gen.NM_013237 Figure 1835: PRO81339 Figure 1801: PRO81323 Figure 1802: DNA324689, XM_166029, Figure 1836: DNA324708, NM_002011, gen.NM_002011 gen.XM_166029 Figure 1803: DNA324690, NM_002520, Figure 1837: PRO81340 Figure 1838: DNA324709, NM_022963, gen.NM_002520 Figure 1804: PRO58993 gen.NM_022963 Figure 1805: DNA324691, XM_043340, Figure 1839: PRO81341 gen.XM_043340 Figure 1840: DNA324710, XM_038946, Figure 1806: PRO81325 gen.XM_038946 Figure 1807: DNA324692, XM_116340, Figure 1841: DNA324711, XM_113454, gen.XM_113454 gen.XM_116340 Figure 1808A-B: DNA324693, XM_043388, Figure 1842: DNA324712, XM_166028, gen.XM_043388 gen.XM_166028

Figure 1843: DNA324713, NM_015043, Figure 1877: DNA324731, XM_168123, gen.NM_015043 gen.XM_168123 Figure 1844: PRO81345 Figure 1878: DNA324732, XM_166457, Figure 1845: DNA324714, XM_113468, gen.XM_166457 gen.XM_113468 Figure 1879: DNA324733, XM_166469, Figure 1846: DNA324715, NM_014275, gen.XM_166469 gen.NM_014275 Figure 1880: DNA324734, NM_018135, Figure 1847: PRO1927 gen.NM_018135 Figure 1848: DNA324716, NM_054013, Figure 1881: PRO81359 gen.NM_054013 Figure 1882A-B: DNA324735, XM_166340, Figure 1849: PRO81347 gen.XM_166340 Figure 1850: DNA270675, NM_005520, Figure 1883: DNA324736, XM_087960, gen.NM_005520 gen.XM_087960 Figure 1851: PRO59040 Figure 1884: DNA324737, XM_166362, Figure 1852: DNA324717, NM .006098, gen.XM_166362 gen.NM_006098 Figure 1885: PRO81362 Figure 1853: PRO25849 Figure 1886: DNA227204, NM_015388. Figure 1854: DNA269593, NM_005110, gen.NM_015388 gen.NM_005110 Figure 1887: PRO37667 Figure 1855: PRO58006 Figure 1888: DNA324738, XM_166425, Figure 1856: DNA324718, XM_116365, gen.XM_166425 gen.XM_116365 Figure 1889: PRO81363 Figure 1857: DNA324719, XM_116511, Figure 1890: DNA324739, NM_057161, gen.XM_116511 gen.NM_057161 Figure 1858: DNA324720, XM_087823, Figure 1891: PRO81364 gen.XM_087823 Figure 1892: DNA270613, NM_006245, Figure 1859A-C: DNA324721, XM_053955, gen.NM_006245 Figure 1893: PRO58984 gen.XM_053955 Figure 1860: DNA324722, XM_113476, Figure 1894: DNA324740, NM_006586, gen.XM_113476 gen.NM_006586 Figure 1861: DNA324723, XM_116514, Figure 1895: PRO81365 gen.XM_116514 Figure 1896: DNA324741, XM_166402, Figure 1862: DNA324724, XM_094741, gen.XM_166402 gen.XM_094741 Figure 1897: PRO81366 Figure 1863: DNA324725, NM_025168, Figure 1898: DNA324742, NM_001760, gen.NM_025168 gen.NM_001760 Figure 1864: PRO81354 Figure 1899: PRO81367 Figure 1865A-B: DNA324726, XM_165740, Figure 1900: DNA287246, NM_004053, gen.NM_004053 gen.XM_165740 Figure 1866: DNA272171, NM _002388, Figure 1901: PRO69521 Figure 1902: DNA324743, NM_017601, gen.NM_002388 Figure 1867: PRO60438 gen.NM_017601 Figure 1868: DNA324727, XM_167169, Figure 1903: PRO81368 gen.XM_167169 Figure 1904: DNA275630, NM_006708, Figure 1869: PRO81355 gen.NM_006708 Figure 1870: DNA324728, NM_014452, Figure 1905: PRO63253 gen.NM_014452 Figure 1906: DNA324744, NM_014341, Figure 1871: PRO868 gen.NM_014341 Figure 1872: DNA324729, XM_166349, Figure 1907: PRO81369 gen.XM_166349 Figure 1908: DNA304460, NM_016059, Figure 1873: PRO81356 gen.NM_016059 Figure 1874: DNA304680, NM_007355, Figure 1909: PRO4984 gen.NM_007355 Figure 1910: DNA324745, XM_166412, Figure 1875: PRO71106 gen.XM_166412 Figure 1876: DNA324730, XM_165772, Figure 1911: PRO81370 gen.XM_165772 Figure 1912: DNA304716, NM_078467,

gen.NM_022551 gen.NM_078467 Figure 1947: PRO71088 Figure 1913: PRO71142 Figure 1948: DNA324767, XM_165747, Figure 1914: DNA324746, XM_166417, gen.XM_165747 gen.XM_166417 Figure 1949: DNA324768, XM_165698, Figure 1915: PRO81371 Figure 1916A-B: DNA324747, NM_003137, gen.XM_165698 gen.NM_003137 Figure 1950: PRO4884 Figure 1951A-B: DNA324769, XM_165770, Figure 1917: PRO81372 Figure 1918A-B: DNA324748, NM_004117, gen.XM_165770 Figure 1952: DNA287227, NM_004159, gen.NM_004117 gen.NM_004159 Figure 1919: PRO36841 Figure 1920: DNA324749, XM_166419, Figure 1953: PRO69506 Figure 1954: DNA324770, XM_165717, gen.XM_166419 gen.XM_165717 Figure 1921: DNA324750, XM_165794, Figure 1955: DNA324771, XM_166480, gen.XM_165794 gen.XM_166480 Figure 1922: DNA324751, NM_007104, Figure 1956: DNA324772, XM_165801, gen.NM_007104 gen.XM_165801 Figure 1923: PRO10360 Figure 1957A-B: DNA324773, NM_000592, Figure 1924: DNA324752, NM_024294, gen.NM_000592 gen.NM_024294 Figure 1958: PRO36316 Figure 1925: PRO81375 Figure 1959: DNA324774, NM .001710, Figure 1926: DNA324753, NM_022758, gen.NM_022758 gen.NM_001710 Figure 1960: PRO36305 Figure 1927: PRO50582 Figure 1928: DNA324754, XM_168070, Figure 1961: DNA227607, NM_005346, gen.XM_168070 gen.NM_005346 Figure 1962: PRO38070 Figure 1929: DNA324755, NM_012391, Figure 1963: DNA304668, NM_005345, gen.NM_012391 gen.NM_005345 Figure 1930: PRO81377 Figure 1964: PRO71095 Figure 1931: DNA324756, XM_166459, Figure 1965: DNA324775, NM_021177, gen.XM_166459 Figure 1932: DNA324757, XM_166333, gen.NM_021177 Figure 1966: PRO81394 gen.XM_166333 Figure 1967A-B: DNA272263, NM_006295, Figure 1933: PRO81379 gen.NM_006295 Figure 1934: DNA324758, XM_058039, Figure 1968: PRO70138 gen.XM_058039 Figure 1969: DNA287319, NM_001288, Figure 1935: PRO81380 Figure 1936: DNA324759, XM_087990, gen.NM_001288 gen.XM_087990 Figure 1970: PRO69584 Figure 1971: DNA324776, NM .001320, Figure 1937: DNA324760, XM_165743, gen.NM_001320 gen.XM_165743 Figure 1972: PRO63052 Figure 1938: DNA324761, XM_166360, Figure 1973A-B: DNA324777, NM_004639, gen.XM_166360 gen.NM_004639 Figure 1939: DNA324763, XM_059801, Figure 1974: PRO81395 gen.XM_059801 Figure 1975A-B: DNA324778, NM .080703, Figure 1940: DNA324764, XM_166363, gen.NM_080703 gen.XM_166363 Figure 1976: PRO81396 Figure 1941: DNA324765, XM_016857, gen.XM_016857 Figure 1977A-B: DNA324779, NM_080702, Figure 1942: DNA227442, NM_001350, gen.NM_080702 gen.NM_001350 Figure 1978: PRO81397 Figure 1979A-B: DNA324780, NM_004638, Figure 1943: PRO37905 gen.NM_004638 Figure 1944: DNA324766, NM_005452, Figure 1980: PRO81398 gen.NM_005452 Figure 1981A-B: DNA324781, NM_080686, Figure 1945: PRO81387 Figure 1946: DNA304661, NM_022551, gen.NM_080686

gen.NM_018950 Figure 1982: PRO81399 Figure 1983: DNA324782, XM_165771, Figure 2018: PRO81414 Figure 2019: DNA324800, XM_166392, gen.XM_165771 gen.XM_166392 Figure 1984: DNA324783, NM_080598, gen.NM_080598 Figure 2020: PRO81415 Figure 2021: DNA324801, XM_166336, Figure 1985: PRO71125 gen.XM_166336 Figure 1986: DNA304699, NM_004640, Figure 2022: PRO81416 gen.NM_004640 Figure 2023: DNA324802, XM_167128, Figure 1987: PRO71125 gen.XM_167128 Figure 1988: DNA324784, XM_165765, Figure 2024: PRO23797 gen.XM_165765 Figure 1989: PRO81400 Figure 2025: DNA324803, XM_167161, Figure 1990: DNA324785, XM_087945, gen.XM_167161 Figure 2026: PRO81417 gen.XM_087945 Figure 2027: DNA324804, NM_013375, Figure 1991: PRO81401 gen.NM_013375 Figure 1992: DNA324786, XM_166381, Figure 2028: PRO81418 gen.XM_166381 Figure 2029: DNA324805, NM_007047, Figure 1993: PRO81402 gen.NM_007047 Figure 1994: DNA324787, XM_168104, Figure 2030: PRO81419 gen.XM_168104 Figure 2031: DNA324806, XM_167179, Figure 1995: DNA324788, XM_166401, gen.XM_167179 gen.XM_166401 Figure 2032: DNA290785, NM_003107, Figure 1996: PRO81404 Figure 1997: DNA271040, NM_001517, gen.NM_003107 gen.NM_001517 Figure 2033: PRO70544 Figure 1998: PRO59365 Figure 2034: DNA150772, NM _003472, gen.NM_003472 Figure 1999A-B: DNA324789, XM_165738, Figure 2035: PRO12797 gen.XM_165738 Figure 2036A-B: DNA324807, XM_165728, Figure 2000: DNA324790, XM_087939, gen.XM_165728 gen.XM_087939 Figure 2037: DNA324808, XM_165749, Figure 2001: PRO81406 Figure 2002: DNA324791, XM_166353, gen.XM_165749 Figure 2038: PRO81421 gen.XM_166353 Figure 2039A-B: DNA324809, NM_004973, Figure 2003: PRO1112 gen.NM_004973 Figure 2004A-B: DNA324792, XM_166376, gen.XM_166376 Figure 2040: PRO81422 Figure 2005: PRO81407 Figure 2041: DNA324810, XM_167196, Figure 2006A-B: DNA324793, XM_165799, gen.XM_167196 Figure 2042: DNA324811, XM_166446, gen.XM_165799 gen.XM_166446 Figure 2007: DNA290264, NM_025263, Figure 2043: PRO81424 gen.NM_025263 Figure 2044A-C: DNA324812, XM_165777, Figure 2008: PRO70393 Figure 2009: DNA324794, XM_166361, gen.XM_165777 Figure 2045: DNA324813, XM_037875, gen.XM_166361 Figure 2010: PRO81409 gen, XM_037875 Figure 2011: DNA324795, XM_165764, Figure 2046: PRO81426 gen.XM_165764 Figure 2047: DNA324814, XM_167225, gen.XM_167225 Figure 2012: PRO81410 Figure 2013: DNA324796, XM_165758, Figure 2048: PRO81427 Figure 2049: DNA324815, XM_166357, gen.XM_165758 Figure 2014: PRO81411 gen.XM_166357 Figure 2050: DNA324816, NM_001069, Figure 2015: DNA324797, XM_166406, gen.NM_001069 gen.XM_166406 Figure 2016: DNA324798, XM_165809, Figure 2051: PRO81429 gen.XM_165809 Figure 2052: DNA324817, NM_001500, Figure 2017: DNA324799, NM_018950, gen.NM_001500

Figure 2053: PRO81430 Figure 2087: DNA324839, XM_167016, Figure 2054A-B: DNA324818, XM_166042, gen.XM_167016 Figure 2088: PRO81449 gen.XM_166042 Figure 2055: PRO51389 Figure 2089: DNA324840, XM_087855, Figure 2056: DNA324819, XM_052721, gen.XM_087855 gen.XM_052721 Figure 2090: DNA324841, XM_087853, Figure 2057: DNA324820, XM_165499, gen.XM_087853 Figure 2091: DNA324842, XM_165669, gen.XM_165499 Figure 2058: DNA324821, XM_114497, gen.XM_165669 Figure 2092: DNA324843, XM_166303, gen.XM_114497 gen.XM_166303 Figure 2059: DNA324822, XM_011117, Figure 2093: PRO81453 gen.XM_011117 Figure 2094: DNA324844, XM_167027, Figure 2060: DNA324823, XM_094855, gen.XM_094855 gen.XM_167027 Figure 2061: PRO81435 Figure 2095: PRO81454 Figure 2062: DNA324824, XM_059776, Figure 2096: DNA324845, XM_167037, gen.XM_059776 gen.XM_167037 Figure 2097: PRO81455 Figure 2063: PRO81436 Figure 2064: DNA324825, XM_055641, Figure 2098: DNA324846, XM_018182, gen.XM_018182 gen.XM_055641 Figure 2099: DNA227924, NM _000165, Figure 2065: DNA324826, XM_004151, gen.XM_004151 gen.NM_000165 Figure 2066: DNA324827, NM_133645, Figure 2100: PRO38387 Figure 2101: DNA324847, XM_166310, gen.NM_133645 Figure 2067: PRO81439 gen.XM_166310 Figure 2102: PRO81457 Figure 2068: DNA324828, XM_097453, gen.XM_097453 Figure 2103: DNA324848, XM_168054, Figure 2069: DNA324829, XM_029228, gen.XM_168054 gen.XM_029228 Figure 2104: DNA271418, NM_003287, gen.NM'_003287 Figure 2070: DNA103471, NM_006670, gen.NM_006670 Figure 2105: PRO59717 Figure 2071: PRO4798 Figure 2106: DNA324849, XM_114492, Figure 2072: DNA324830, XM_068963, gen.XM_114492 Figure 2107: DNA324850, XM_037056, gen.XM_068963 gen.XM_037056 Figure 2073: PRO81441 Figure 2108: DNA324851, XM_098468, Figure 2074: DNA324831, XM _040623, gen.XM_098468 gen.XM_040623 Figure 2109: PRO19933 Figure 2075: DNA324832, NM_020320, Figure 2110: DNA324852, XM_004526, gen.NM_020320 Figure 2076: PRO81443 gen.XM_004526 Figure 2077: DNA324833, NM _014107, Figure 2111: DNA324853, NM_001016, gen.NM_014107 gen.NM_001016 Figure 2078: PRO81444 Figure 2112: PRO81462 Figure 2113: DNA324854, XM_004297, Figure 2079A-B: DNA324834, XM_084204, gen.XM_004297 gen.XM_084204 Figure 2114: DNA324855, XM_004256, Figure 2080: DNA324835, XM_017517, gen.XM_004256 gen.XM_017517 Figure 2115: PRO81464 Figure 2081: DNA324836, NM_032929, gen.NM_032929 Figure 2116: DNA324856, NM_014320, Figure 2082: PRO81446 gen.NM_014320 Figure 2083: DNA324837, XM _003611, Figure 2117: PRO81465 gen.XM_003611 Figure 2118: DNA324857, XM_059741, Figure 2084: PRO81447 gen.XM_059741 Figure 2085: DNA324838, XM_068919, Figure 2119: DNA324858, XM_017831, gen.XM_068919 gen.XM_017831 Figure 2086: PRO81448 Figure 2120: PRO81467

Figure 2121: DNA324859, XM_049899, Figure 2154: DNA324883, XM_087991, gen.XM_049899 gen.XM_087991 Figure 2155: DNA324884, NM_005514, Figure 2122: DNA324860, XM_004379, gen.XM_004379 gen.NM_005514 Figure 2123A-C: DNA324861, XM_087834, Figure 2156: PRO81490 Figure 2157: DNA324885, XM_166327, gen.XM_087834 Figure 2124A-B: DNA324862, XM_087836, gen.XM_166327 Figure 2158: PRO81491 gen.XM_087836 Figure 2159: DNA324886, XM_165692, Figure 2125: PRO81471 Figure 2126: DNA324863, NM_005389, gen.XM_165692 Figure 2160: DNA324887, XM_117449, gen.NM_005389 Figure 2127: PRO66279 gen.XM_117449 Figure 2128A-C: DNA324864, XM_029746, Figure 2161: DNA324888, XM_086428, gen.XM_086428 gen.XM_029746 Figure 2162: PRO81494 Figure 2129: PRO66282 Figure 2163: DNA324889, NM_032350, Figure 2130: DNA324865, XM_004383, gen.NM_032350 gen.XM_004383 Figure 2164: PRO81495 Figure 2131: DNA324866, XM_059745, Figure 2165: DNA324890, NM_013393, gen.XM_059745 gen.NM_013393 Figure 2132: DNA324867, XM.033912, gen.XM_033912 Figure 2166: PRO81496 Figure 2167: DNA324891, XM_165860, Figure 2133: PRO81474 gen.XM_165860 Figure 2134: DNA324868, XM_033910, Figure 2168: DNA324892, XM_166541, gen.XM_033910 Figure 2135: DNA324870, NM_003181, gen.XM_166541 Figure 2169: PRO81498 gen.NM_003181 Figure 2170A-B: DNA324893, XM_166523, Figure 2136: PRO81476 gen.XM_166523 Figure 2137: DNA324871, NM_002793, Figure 2171: PRO81499 gen.NM_002793 Figure 2138: PRO81477 Figure 2172: DNA324894, NM_016003, Figure 2139: DNA324872, XM_044866, gen.NM_016003 Figure 2173: PRO81500 gen.XM_044866 Figure 2174: DNA225631, NM_001101, Figure 2140: DNA324873, XM_116524, gen.XM_116524 gen.NM_001101 Figure 2175: PRO36094 Figure 2141: DNA324874, XM_059773, gen.XM_059773 Figure 2176: DNA274326, NM_003088, Figure 2142: DNA324875, XM_084998, gen.NM_003088 Figure 2177: PRO62244 gen.XM_084998 Figure 2178: DNA324895, NM_006303, Figure 2143: PRO81481 gen.NM_006303 Figure 2144: DNA324876, XM_058266, Figure 2179: PRO81501 gen.XM_058266 Figure 2180: DNA324896, NM_014413, Figure 2145: DNA324877, XM_042422, gen.NM_014413 gen.XM_042422 Figure 2181: PRO60579 Figure 2146A-B: DNA324878, XM_054706, Figure 2182: DNA247595, NM_006908, gen.XM_054706 Figure 2147: DNA324879, XM_166049, gen.NM_006908 Figure 2183: PRO45014 gen.XM_166049 Figure 2184: DNA324897, NM_006854, Figure 2148: DNA324880, XM_042473, gen.NM_006854 gen.XM_042473 Figure 2185: PRO12468 Figure 2149: PRO81486 Figure 2186: DNA324898, NM_024067, Figure 2150: DNA324881, XM_167046, gen.NM_024067 gen.XM_167046 Figure 2187: PRO81502 Figure 2151: PRO23797 Figure 2188: DNA324899, NM_002947, Figure 2152: DNA324882, XM_071937, gen.XM_071937 gen.NM_002947 Figure 2153: PRO81487 Figure 2189: PRO81503

Figure 2190: DNA324900, XM-166531, . gen.XM_166494 gen.XM_166531 Figure 2225: DNA324920, XM_107825, Figure 2191: DNA324901, XM_166540, gen.XM_107825 Figure 2226A-B: DNA324921, NM_022748, gen.XM_166540 gen.NM_022748 Figure 2192: PRO81505 Figure 2227: PRO81523 Figure 2193: DNA193955, NM_002489, Figure 2228: DNA324922, NM_000598, gen.NM_002489 Figure 2194: PRO23362 gen.NM_000598 Figure 2195: DNA324902, XM_088264, Figure 2229: PRO119 Figure 2230A-B: DNA324923, XM_166594, gen.XM_088264 gen.XM_166594 Figure 2196: PRO81506 Figure 2231: PRO81524 Figure 2197: DNA324903, XM_165841, Figure 2232A-B: DNA275334, NM_030900, gen.XM_165841 gen.NM_030900 Figure 2198: DNA324904, XM_166521, Figure 2233: PRO63009 gen.XM_166521 Figure 2234: DNA324924, NM_031443, Figure 2199: PRO81508 Figure 2200: DNA324905, XM_166506, gen.NM_031443 Figure 2235: PRO81525 gen.XM_166506 Figure 2201: PRO81509 Figure 2236: DNA324925, NM_012412, gen.NM_012412 Figure 2202: DNA324906, XM_166505, Figure 2237: PRO61812 gen.XM_166505 Figure 2238: DNA324926, NM_021130, Figure 2203: DNA324907, XM_166514, gen.NM_021130 gen.XM_166514 Figure 2239: PRO7427 Figure 2204: DNA324908, XM_166515, Figure 2240A-B: DNA324927, XM_165877, gen.XM_166515 gen.XM_165877 Figure 2205: DNA324909, XM_166512, Figure 2241: PRO81526 gen.XM_166512 Figure 2242: DNA227268, NM_019082, Figure 2206: DNA227929, NM_019059, gen.NM_019082 gen.NM_019059 Figure 2243: PRO37731 Figure 2207: PRO38392 Figure 2244: DNA324928, XM_015258, Figure 2208A-B: DNA324910, NM_018947, gen.XM_015258 gen.NM_018947 Figure 2209: PRO81514 Figure 2245: DNA324929, XM_165870, gen.XM_165870 Figure 2210: DNA324911, NM _002137, Figure 2246: DNA273865, NM_006230, gen.NM_002137 gen.NM_006230 Figure 2211: PRO81515 Figure 2247: PRO61824 Figure 2212: DNA324912, NM _031243, Figure 2248A-B: DNA324930, XM_165882, gen.NM_031243 gen.XM_165882 Figure 2213: PRO6373 Figure 2249: DNA324931, XM_165867, Figure 2214: DNA324913, NM_007276, gen.NM_007276 gen.XM_165867 Figure 2215: PRO81516 Figure 2250: PRO61688 Figure 2216: DNA324914, NM_016587, Figure 2251: DNA324932, NM_014063, gen.NM_016587 gen.NM_014063 Figure 2252: PRO81529 Figure 2217: PRO81517 Figure 2253: DNA324933, XM_165872, Figure 2218: DNA324915, XM_040853, gen.XM_165872 gen.XM_040853 Figure 2254: DNA304707, NM_002787, Figure 2219: DNA324916, XM_166509, gen.NM_002787 gen.XM_166509 Figure 2255: PRO71133 Figure 2220: DNA324917, XM_166513, Figure 2256: DNA324934, XM_016733, gen.XM_166513 Figure 2221: PRO81520 gen.XM_016733 Figure 2257: PRO81531 Figure 2222: DNA324918, XM_166504, Figure 2258: DNA324935, XM_165876, gen.XM_166504 gen.XM_165876 Figure 2223: PRO81521 Figure 2259A-B: DNA324936, NM_014800, Figure 2224: DNA324919, XM_166494,

gen.NM_014800 Figure 2295A-B: DNA324954, NM_032999, Figure 2260: DNA324937, NM_130442, gen.NM_032999 gen.NM_130442 Figure 2296: PRO81551 Figure 2297: DNA324955, XM_088239, Figure 2261: PRO81534 Figure 2262: DNA226416, NM_000385, gen.XM_088239 Figure 2298: PRO81552 gen.NM_000385 Figure 2263: PRO36879 Figure 2299A-B: DNA324956, XM_167500, Figure 2264A-B: DNA324938, XM_167339, gen.XM_167500 Figure 2300A-B: DNA324957, XM_167504, gen.XM_167339 gen.XM_167504 Figure 2265: DNA287189, NM_002047, gen.NM_002047 Figure 2301: DNA324958, XM_167498, Figure 2266: PRO69475 gen.XM_167498 Figure 2302: DNA324959, XM_168454, Figure 2267: DNA324939, XM_170195, gen.XM_168454 gen.XM_170195 Figure 2303: PRO81556 Figure 2268: PRO81536 Figure 2269: DNA324940, XM_168378, Figure 2304: DNA324960, NM_031925, gen.XM_168378 gen.NM_031925 Figure 2270: PRO81537 Figure 2305: PRO81557 Figure 2306: DNA324961, NM_005918, Figure 2271: DNA324941, XM_168354, gen.XM_168354 gen.NM_005918 Figure 2272: PRO81538 Figure 2307: PRO81558 Figure 2273: DNA324942, XM_167494, Figure 2308: DNA304710, NM_001540, gen.XM_167494 gen.NM_001540 Figure 2274: DNA103588, NM_001762, Figure 2309: PRO71136 Figure 2310: DNA324962, XM_168470, gen.NM_001762 Figure 2275: PRO4912 gen.XM_168470 Figure 2276: DNA324943, XM_037741, Figure 2311: DNA324963, XM_168461, gen.XM_168461 gen.XM_037741 Figure 2277: PRO81540 Figure 2312A-B: DNA324964, XM_167502, Figure 2278: DNA324944, XM_050265, gen.XM_167502 gen.XM_050265 Figure 2313: DNA324965, XM_017442, Figure 2279: PRO81541 gen.XM_017442 Figure 2314: PRO81561 Figure 2280: DNA324945, XM_017483, gen.XM_017483 Figure 2315: DNA324966, XM_168450, Figure 2281A-B: DNA324946, XM_018359, gen.XM_168450 Figure 2316: DNA324967, XM_168435, gen.XM_018359 Figure 2282: DNA324947, XM_059876, gen.XM_168435 Figure 2317: DNA324968, XM_168464, gen.XM_059876 Figure 2283: PRO81544 gen.XM.168464 Figure 2318: DNA324969, XM_170427, Figure 2284: DNA324948, NM_032951, gen.NM_032951 gen.XM_170427 Figure 2285: PRO81545 Figure 2319A-B: DNA324971, NM_015068, Figure 2286: DNA324949, NM_032953, gen.NM_015068 Figure 2320: PRO81566 gen.NM_032953 Figure 2287: PRO81546 Figure 2321A-B: DNA324972, XM_167476, Figure 2288: DNA324950, NM_022170, gen.XM_167476 Figure 2322: DNA324973, XM_168181, gen.NM_022170 Figure 2289: PRO81547 gen.XM_168181 Figure 2290: DNA324951, NM_031992, Figure 2323: DNA324974, XM_168251, gen.NM_031992 gen.XM_168251 Figure 2291: PRO81548 Figure 2324: PRO81569 Figure 2292: DNA324952, XM_004901, Figure 2325: DNA324975, XM_167477, gen.XM_004901 gen.XM_167477 Figure 2293: DNA324953, NM_016328, Figure 2326: DNA324976, NM_005837, gen.NM_005837 gen.NM_016328 Figure 2294: PRO81550 Figure 2327: PRO81571

gen.NM_057089 Figure 2328: DNA324977, XM_167483, Figure 2364: PRO81588 gen.XM_167483 Figure 2365: DNA324995, NM _001283, Figure 2329: DNA324978, XM_167484, gen.NM_001283 gen.XM_167484 Figure 2366: PRO41882 Figure 2330: PRO81572 Figure 2367: DNA324996, NM_003378, Figure 2331: DNA324979, NM_030935, gen.NM_030935 gen.NM_003378 Figure 2368: PRO81589 Figure 2332: PRO81573 Figure 2369: DNA324997, NM_001084, Figure 2333: DNA324980, NM _019606, gen.NM_001084 gen.NM_019606 Figure 2370: PRO58437 Figure 2334: PRO81574 Figure 2371: DNA270711, NM_006349, Figure 2335: DNA324981, NM _024070, gen.NM_006349 gen.NM_024070 Figure 2372: PRO59074 Figure 2336: PRO81575 Figure 2373: DNA324998, NM_024653, Figure 2337: DNA324982, XM_084241, gen.XM_084241 gen.NM_024653 Figure 2374: PRO81590 Figure 2338: DNA324983, NM_006833, Figure 2375: DNA324999, XM_168548, gen.NM_006833 gen.XM_168548 Figure 2339: PRO22897 Figure 2376: DNA325000, NM_032958, Figure 2340: DNA324984, NM_032164, gen.NM_032958 gen.NM_032164 Figure 2377: PRO81591 Figure 2341: PRO81578 Figure 2378: DNA325001, NM_002803, Figure 2342: DNA304801, NM_004889, gen.NM_002803 gen.NM_004889 Figure 2379: PRO81592 Figure 2343: PRO71211 Figure 2380: DNA325002, XM_168572, Figure 2344: DNA324985, NM _006693, gen.XM_168572 gen.NM_006693 Figure 2381: DNA325003, XM_071605, Figure 2345: PRO81579 gen.XM_071605 Figure 2346: DNA324986, XM_165839, Figure 2382: PRO81594 gen.XM_165839 Figure 2347: PRO81580 Figure 2383: DNA325004, XM .033876, Figure 2348: DNA272090, NM_005720, gen.XM_033876 Figure 2384: PRO81595 gen.NM_005720 Figure 2385A-B: DNA325005, XM_027214, Figure 2349: PRO60360 gen.XM_027214 Figure 2350: DNA324987, XM_165836, Figure 2386: DNA325006, XM_088073, gen.XM_165836 gen.XM_088073 Figure 2351A-B: DNA324988, XM_166482, Figure 2387: DNA325007, XM_072430, gen.XM_166482 gen.XM_072430 Figure 2352: DNA324989, XM .088180, Figure 2388: PRO81598 gen.XM_088180 Figure 2389: DNA325008, XM_050430, Figure 2353A-B: DNA324990, XM_166485, gen.XM_050430 gen.XM_166485 Figure 2390: PRO81599 Figure 2354: PRO81584 Figure 2391: DNA325009, NM _001753, Figure 2355: DNA324991, NM_001673, gen.NM_001753 gen.NM_001673 Figure 2392: PRO81600 Figure 2356: PRO81585 Figure 2393: DNA226560, NM .006136, Figure 2357: DNA324992, NM_133436, gen.NM_006136 gen.NM_133436 Figure 2394: PRO37023 Figure 2358: PRO81586 Figure 2395: DNA325010, XM_012284, Figure 2359: DNA324993, XM_168586, gen.XM_168586 gen.XM_012284 Figure 2396: DNA325011, NM .005000, . Figure 2360: PRO81587 gen.NM_005000 Figure 2361: DNA83141, NM_000602, Figure 2397: PRO59380 gen.NM_000602 Figure 2398: DNA325012, NM _001662, Figure 2362: PRO2604 gen.NM_001662 Figure 2363: DNA324994, NM_057089,

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gen.NM_017760

Figure 2433: DNA325034, XM_016700,

Figure 2502: PRO81652

Figure 2503: DNA325073, NM_025232, Figure 2468: PRO81637 gen.NM_025232 Figure 2469: DNA325054, XM_036413, Figure 2504: PRO81653 gen.XM_036413 Figure 2470A-B: DNA325055, XM_032944, Figure 2505: DNA325074, XM_027440, gen.XM_027440 gen.XM_032944 Figure 2506: DNA225671, NM_001831, Figure 2471: DNA325056, XM_117444, gen.NM_001831 gen.XM_117444 Figure 2507: PRO36134 Figure 2472: DNA325057, XM_117452, Figure 2508: DNA325075, NM .024567, gen.XM_117452 gen.NM_024567 Figure 2473: DNA325058, XM _070203, Figure 2509: PRO81654 gen.XM_070203 Figure 2510: DNA325076, NM_018250, Figure 2474: PRO81641 gen.NM_018250 Figure 2475: DNA325059, XM_095371, Figure 2511: PRO81655 gen.XM_095371 Figure 2512: DNA227267, NM_018660, Figure 2476: DNA325060, NM_004084, gen.NM_018660 gen.NM_004084 Figure 2513: PRO37730 Figure 2477: PRO2570 Figure 2514A-B: DNA325077, XM_095545, Figure 2478: DNA325061, NM_005217, gen.XM_095545 gen.NM_005217 Figure 2515: DNA325078, XM_088338, Figure 2479: PRO9980 Figure 2480: DNA325062, XM_070188, gen.XM_088338 Figure 2516: PRO81657 gen.XM_070188 Figure 2517: DNA325079, XM_114617, Figure 2481: PRO81643 gen.XM_114617 Figure 2482: DNA325063, XM .035680, Figure 2518: PRO81658 gen.XM_035680 Figure 2519: DNA325080, XM_088336, Figure 2483: DNA325064, XM_035662, gen.XM_088336 gen.XM_035662 Figure 2520: PRO81659 Figure 2484: PRO3344 Figure 2521: DNA325081, XM_047083, Figure 2485: DNA325065, XM_005305, gen.XM_047083 gen.XM_005305 Figure 2522: PRO81660 Figure 2486: PRO81645 Figure 2523: DNA325082, XM_114618, Figure 2487: DNA325066, XM_050293, gen.XM_114618 gen.XM_050293 Figure 2524: PRO81661 Figure 2488A-B: DNA325067, XM_027679, Figure 2525: DNA325083, XM_050215, gen.XM_027679 gen.XM_050215 Figure 2489: PRO81647 Figure 2526: DNA325084, XM_113531, Figure 2490A-B: DNA325068, XM_027651, gen.XM_113531 gen_XM_027651 Figure 2527: DNA325085, NM_018310, Figure 2491: DNA274178, NM_005775, gen.NM_018310 gen.NM_005775 Figure 2528: PRO81664 Figure 2492: PRO62108 Figure 2529: DNA325086, XM_088294, Figure 2493: DNA325069, XM_113557, gen.XM_088294 gen.XM_113557 Figure 2530: DNA325087, XM_013112, Figure 2494: PRO81649 gen.XM_013112 Figure 2495: DNA83022, NM_001199, Figure 2531: DNA325088, XM_059933, gen.NM_001199 gen.XM_059933 Figure 2496: PRO2042 Figure 2532: PRO1108 Figure 2497: DNA325070, NM_006128, Figure 2533: DNA325089, XM_011629, gen.NM_006128 gen.XM_011629 Figure 2498: PRO81650 Figure 2534: DNA325090, NM_000930, Figure 2499: DNA325071, NM_006131, gen.NM_000930 gen.NM_006131 Figure 2535: PRO4 Figure 2500: PRO81651 Figure 2536: DNA325091, NM .000931, Figure 2501: DNA325072, NM_006132, gen.NM_000931 gen.NM_006132 Figure 2537: PRO81668

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gen.XM_084514	gen.XM_015705
Figure 3181: PRO81965	Figure 3215: DNA272728, NM_003146,
Figure 3182: DNA325442, XM_084516,	gen.NM_003146
gen.XM_084516	Figure 3216: PRO60847
Figure 3183: DNA325443, XM_084515,	Figure 3217: DNA325461, XM_165611,
gen.XM_084515	gen.XM_165611
Figure 3184: DNA325444, XM_084517,	Figure 3218: DNA287417, NM_024098,
gen.XM_084517	gen.NM_024098
Figure 3185: DNA325445, XM_034431,	Figure 3219: PRO69674
gen.XM_034431	Figure 3220: DNA227088, NM_014502,
Figure 3186: PRO11691	gen.NM_014502
Figure 3187: DNA325446, XM _030326,	Figure 3221: PRO37551
gen.XM_030326	Figure 3222: DNA325462, XM_165610,
Figure 3188: DNA325447, NM_057174,	gen.XM_165610
gen.NM_057174	Figure 3223A-B: DNA325463, XM_165612,
Figure 3189: PRO81970	gen.XM_165612
Figure 3190: DNA325448, NM_004813,	Figure 3224: DNA325464, XM_166234,
gen.NM_004813	gen.XM_166234
Figure 3191: PRO81971	Figure 3225: DNA325465, NM_015533,
Figure 3192: DNA325449, XM_167437,	gen.NM_015533
gen.XM_167437	Figure 3226: PRO81988
Figure 3193: DNA325450, XM_054856,	Figure 3227: DNA325466, XM_166232,
gen.XM_054856	gen.XM_166232
Figure 3194: DNA325451, XM_004330,	Figure 3228A-B: DNA325467, XM_167748,
gen.XM_004330	gen.XM_167748
Figure 3195: DNA325452, XM_084681,	Figure 3229: PRO81990
gen.XM_084681	Figure 3230: DNA325468, NM_004739,
Figure 3196: DNA325453, XM_006297,	gen.NM_004739
gen.XM_006297	Figure 3231: PRO81991
Figure 3197: DNA325454, NM_003646,	Figure 3232: DNA325469, NM_014610,

gen.NM_014610 Figure 3268: DNA325488, XM_113223, Figure 3233: PRO81992 gen.XM_113223 Figure 3269: DNA325489, XM_045642, Figure 3234: DNA325470, XM_167747, gen.XM_045642 gen.XM_167747 Figure 3270: DNA325490, XM_006533, Figure 3235: PRO81993 gen.XM_006533 Figure 3236: DNA287254, NM_024099, gen.NM_024099 Figure 3271: DNA325491, XM_045613, gen.XM_045613 Figure 3237: PRO69528 Figure 3272: PRO59721 Figure 3238: DNA325471, NM_015853, Figure 3273A-B: DNA325492, XM_045612, gen.NM_015853 gen.XM_045612 Figure 3239: PRO81994 Figure 3240: DNA325472, NM_032667, Figure 3274: PRO82009 Figure 3275: DNA325493, XM_113224, gen.NM_032667 gen.XM_113224 Figure 3241: PRO81995 Figure 3276: DNA325494, XM_045499, Figure 3242: DNA325473, NM _006362, gen.XM_045499 gen.NM_006362 Figure 3277: PRO82011 Figure 3243: PRO81996 Figure 3278: DNA325495, XM_045525, Figure 3244: DNA325474, XM_167716, gen.XM_045525 gen.XM_167716 Figure 3245: DNA75863, NM _002411, Figure 3279: DNA325496, NM_013265, gen.NM_013265 gen.NM_002411 Figure 3280: PRO82013 Figure 3246: PRO2018 Figure 3281: DNA325497, XM _006529, Figure 3247: DNA325475, XM_087710, gen.XM_006529 gen.XM_087710 Figure 3282: PRO60008 Figure 3248: DNA325476, XM_167726, Figure 3283: DNA325498, XM_053787, gen.XM_167726 gen.XM_053787 Figure 3249: DNA325477, NM_004265, gen.NM_004265 Figure 3284: DNA269803, NM _001667, gen.NM_001667 Figure 3250: PRO12878 Figure 3251A-B: DNA325478, NM_013402, Figure 3285: PRO58207 Figure 3286: DNA325499, XM_115031, gen.NM_013402 Figure 3252: PRO81999 gen.XM_115031 Figure 3287: DNA325500, XM_084702, Figure 3253: DNA325479, NM_004111, gen.XM_084702 gen.NM_004111 Figure 3288: DNA325501, XM_053796, Figure 3254: PRO69568 gen.XM_053796 Figure 3255: DNA325480, XM_048286, Figure 3289: DNA325502, NM -002689, gen.XM_048286 gen.NM_002689 Figure 3256: DNA325481, NM_004322, Figure 3290: PRO82018 gen.NM_004322 Figure 3291A-D: DNA325503, XM_167804, Figure 3257: PRO20117 gen.XM_167804 Figure 3258: DNA325482, NM _032989, Figure 3292: PRO82019 gen.NM_032989 Figure 3293: DNA325504, XM_166235, Figure 3259: PRO20117 Figure 3260: DNA325483, XM_011988, gen.XM_166235 Figure 3294: DNA325505, XM_166236, gen.XM_011988 gen.XM_166236 Figure 3261: DNA325484, NM_031472, Figure 3295: DNA270721, NM_006842, gen.NM_031472 gen.NM_006842 Figure 3262: PRO82002 Figure 3296: PRO59084 Figure 3263: DNA325485, XM_037808, Figure 3297: DNA189687, NM _000852, gen.XM_037808 Figure 3264: DNA325486, NM_004074, gen.NM_000852 gen.NM_004074 Figure 3298: PRO25845 Figure 3265: PRO82004 Figure 3299: DNA325506, NM_007103, Figure 3266: DNA325487, NM_017670, gen.NM_007103 Figure 3300: PRO58606 gen.NM_017670 Figure 3301: DNA325507, NM_005851, Figure 3267: PRO82005

gen.XM_166253 gen.NM_005851 Figure 3337: DNA325526, NM _001293, Figure 3302: PRO69461 Figure 3303A-B: DNA325508, XM_165598, gen.NM_001293 Figure 3338: PRO82034 gen.XM_165598 Figure 3304: DNA325509, NM_006019, Figure 3339: DNA325527, XM _042852, gen.XM_042852 gen.NM_006019 Figure 3340: PRO82035 Figure 3305: PRO82023 Figure 3341: DNA325528, XM_165628, Figure 3306: DNA325510, NM_006053, gen.XM_165628 gen.NM_006053 Figure 3342A-B: DNA325529, NM_080491, Figure 3307: PRO24831 Figure 3308: DNA325511, XM_166196, gen.NM_080491 Figure 3343: PRO82037 gen.XM_166196 Figure 3344A-B: DNA325530, NM_012296, Figure 3309: PRO82024 gen.NM_012296 Figure 3310: DNA325512, XM_165600, Figure 3345: PRO60311 gen.XM_165600 Figure 3346: DNA325531, NM _032379, Figure 3311A-B: DNA325513, NM 053056, gen.NM_032379 gen.NM_053056 Figure 3347: PRO82038 Figure 3312: PRO4870 Figure 3348: DNA325532, NM_007173, Figure 3313: DNA103474, NM_003824, gen.NM_007173 gen.NM_003824 Figure 3349: DNA325533, XM_166239, Figure 3314: PRO4801 gen.XM_166239 Figure 3315: DNA325514, XM_096486, Figure 3350: DNA325534, XM _084610, gen.XM_096486 gen.XM_084610 Figure 3316A-B: DNA325515, NM_003626, gen.NM_003626 Figure 3351: PRO82040 Figure 3352: DNA325535, XM .058450, Figure 3317: PRO82027 Figure 3318A-B: DNA325516, XM_167853, gen.XM_058450 Figure 3353: DNA325536, XM_084601, gen.XM_167853 gen.XM_084601 Figure 3319: PRO82028 Figure 3354: PRO82042 Figure 3320: DNA325517, NM _014042, Figure 3355A-B: DNA325537, XM_006464, gen.NM_014042 gen.XM_006464 Figure 3321: PRO82029 Figure 3356: PRO82043 Figure 3322A-B: DNA325518, NM _001567, Figure 3357: DNA325538, XM_084570, gen.NM_001567 gen.XM_084570 Figure 3323: PRO61238 Figure 3358: DNA325539, XM .051435, Figure 3324: DNA325519, XM_167433, gen.XM_051435 gen.XM_167433 Figure 3359: DNA325540, NM .001467, Figure 3325: DNA325520, XM_165616, gen.NM_001467. gen.XM_165616 Figure 3360: PRO82045 Figure 3326: DNA325521, NM_032871, Figure 3361: DNA325541, NM_001028, gen.NM_032871 gen.NM_001028 Figure 3327: PRO57307 Figure 3362: PRO82046 Figure 3328: DNA325522, XM_165631, Figure 3363: DNA325542, XM_113230, gen.XM_165631 Figure 3329: DNA254186, NM_014752, gen.XM_113230 Figure 3364: DNA325543, XM_115062, gen.NM_014752 gen.XM_115062 Figure 3330: PRO49298 Figure 3331: DNA325523, NM_001005, Figure 3365: DNA325544, XM_115063, gen.XM_115063 gen.NM_001005 Figure 3366: DNA325545, XM_113229, Figure 3332: PRO82032 gen.XM_113229 Figure 3333: DNA88176, NM_001235, Figure 3367A-B: DNA325546, XM_051489, gen.NM_001235 gen.XM_051489 Figure 3334: PRO2685 Figure 3368: PRO82050 Figure 3335A-B: DNA325524, XM_165627, Figure 3369: DNA325547, NM_022003, gen.XM_165627 gen.NM_022003 Figure 3336: DNA325525, XM_166253,

Figure 3370: PRO82051 Figure 3405: PRO82066 Figure 3371: DNA325548, XM_006432, Figure 3406: DNA325565, XM_166177, gen.XM_006432 gen.XM_166177 Figure 3407: DNA325566, XM_165571, Figure 3372: PRO82052 Figure 3373: DNA325549, XM_051716, gen.XM_165571 Figure 3408: PRO82068 gen.XM_051716 Figure 3409: DNA325567, XM_166174, Figure 3374: DNA325550, NM_025164, gen.XM_166174 gen.NM_025164 Figure 3375: PRO82054 Figure 3410: PRO82069 Figure 3411: DNA325568, NM _001274, Figure 3376: DNA225752, NM_000039, gen.NM_001274 gen.NM_000039 Figure 3412: PRO12187 Figure 3377: PRO36215 Figure 3413: DNA325569, XM_165586, Figure 3378: DNA325551, XM_052113, gen.XM_165586 gen.XM_052113 Figure 3414: DNA325570, XM_165584, Figure 3379: PRO82055 gen.XM_165584 Figure 3380: DNA271324, NM_006169, Figure 3415: DNA257965, NM_032873, gen.NM_006169 gen.NM_032873 Figure 3381: PRO59629 Figure 3416: PRO52492 Figure 3382: DNA325552, XM _084658, Figure 3417: DNA325571, XM_167780, gen.XM_084658 gen.XM_167780 Figure 3383: PRO82056 Figure 3418: DNA325572, XM_166743, Figure 3384: DNA325553, NM _000795, gen.XM_166743 gen.NM_000795 Figure 3419: PRO82072 Figure 3385: PRO12448 Figure 3420: DNA325573, NM_012101, Figure 3386: DNA325554, NM_017868, gen.NM_012101 gen.NM_017868 Figure 3421: PRO82073 Figure 3387: PRO82057 Figure 3422: DNA325574, NM_058193, Figure 3388: DNA325555, XM_084654, gen.NM_058193 gen.XM_084654 Figure 3423: PRO82074 Figure 3389: PRO82058 Figure 3424: DNA325575, XM_084522, Figure 3390: DNA272413, NM_003002, gen.XM_084522 gen.NM_003002 Figure 3425: PRO82075 Figure 3391: PRO60666 Figure 3426: DNA325576, XM_091786, Figure 3392: DNA271843, NM _004398, gen.XM_091786 gen.NM_004398 Figure 3427: DNA325577, XM_165390, Figure 3393: PRO60123 gen.XM_165390 Figure 3394: DNA325556, XM_017369, gen.XM_017369 Figure 3428: DNA325578, XM_084525, Figure 3395: DNA325557, NM _032299, gen.XM_084525 Figure 3429A-B: DNA325579, XM_010494, gen.NM_032299 gen.XM_010494 Figure 3396: PRO82060 Figure 3430A-B: DNA325580, NM_015064, Figure 3397: DNA325558, XM_055369, gen.NM_015064 gen.XM_055369 Figure 3431: PRO82078 Figure 3398: DNA325559, XM_051430, Figure 3432: DNA325581, NM_030775, gen.XM_051430 gen.NM_030775 Figure 3399: DNA325560, XM_006467, Figure 3433: PRO71031 gen.XM_006467 Figure 3434: DNA297398, NM_032642, Figure 3400: DNA325561, XM_113226, gen.NM_032642 gen.XM_113226 Figure 3435: PRO71031 Figure 3401: DNA325562, XM_165592, Figure 3436: DNA325582, XM_017080, gen.XM_165592 gen.XM_017080 Figure 3402: PRO82064 Figure 3437: DNA325583, XM_113739, Figure 3403: DNA325563, XM_166181, gen.XM_113739 gen.XM_166181 Figure 3438: PRO82080 Figure 3404: DNA325564, XM_052862, Figure 3439: DNA325584, NM_002014, gen.XM_052862

gen.NM_002046

Figure 3474: PRO36095 gen.NM_002014 Figure 3475A-B: DNA325602, XM_006958, Figure 3440: PRO59262 Figure 3441: DNA325585, XM_096661, gen.XM_006958 Figure 3476: DNA83180, NM_002342, gen.XM_096661 gen.NM_002342 Figure 3442: DNA325586, NM_018463, Figure 3477: PRO2622 gen.NM_018463 Figure 3478: DNA103514, NM_001038, Figure 3443: PRO82082 gen.NM_001038 Figure 3444: DNA325587, NM_021953, Figure 3479: PRO4841 gen.NM_021953 Figure 3480: DNA188396, NM_001065, Figure 3445: PRO82083 gen.NM_001065 Figure 3446: DNA325588, NM_031465, Figure 3481: PRO21924 gen.NM_031465 Figure 3447: PRO82084 Figure 3482A-C: DNA325603, XM .006947, Figure 3448: DNA325589, NM_005002, gen.XM_006947 Figure 3483A-B: DNA325604, XM_006936, gen.NM_005002 Figure 3449: PRO82085 gen.XM_006936 Figure 3484: PRO82097 Figure 3450: DNA325590, XM_033227, Figure 3485A-B: DNA325605, XM .006925, gen.XM_033227 gen.XM_006925 Figure 3451: DNA325591, XM_116926, Figure 3486: DNA325606, XM_096630, gen.XM_116926 Figure 3452: DNA88114, NM_001734, gen.XM_096630 Figure 3487: PRO82099 gen.NM_001734 Figure 3488: DNA325607, XM_084901, Figure 3453: PRO2660 Figure 3454: DNA325592, XM_058574, gen.XM_084901 Figure 3489: DNA226028, NM_002355, gen.XM_058574 gen.NM_002355 Figure 3455: DNA325593, NM_007273, Figure 3490: PRO36491 gen.NM_007273 Figure 3491: DNA325608, XM_031807, Figure 3456: PRO36970 gen.XM_031807 Figure 3457A-B: DNA325594, XM_032588, Figure 3492: PRO82101 gen.XM_032588 Figure 3493A-B: DNA325609, XM_049663, Figure 3458: DNA325595, NM_001975, gen.XM_049663 gen.NM_001975 Figure 3494: DNA325610, XM_012159, Figure 3459: PRO38010 gen.XM_012159 Figure 3460: DNA325596, NM_000365, Figure 3495: DNA325611, XM_084922, gen.NM_000365 gen.XM_084922 Figure 3461: PRO69549 Figure 3496: DNA325612, NM_031289, Figure 3462: DNA325597, XM_032614, gen.NM_031289 gen.XM_032614 Figure 3463: DNA325598, NM_002075, Figure 3497: PRO82104 Figure 3498: DNA226771, NM_003979, gen.NM_002075 gen.NM_003979 Figure 3464: PRO82091 Figure 3499: PRO37234 Figure 3465: DNA325599, XM_165910, Figure 3500: DNA325613, XM_084918, gen.XM_165910 gen.XM_084918 Figure 3466: DNA151827, NM_005439, Figure 3501: DNA325614, NM_007178, gen.NM_005439 gen.NM_007178 Figure 3467: PRO12902 Figure 3502: PRO82106 Figure 3468A-B: DNA254624, NM_001273, Figure 3503: DNA325615, XM_041100, gen.NM_001273 gen.XM_041100 Figure 3469: PRO49726 Figure 3504A-B: DNA325616, XM .058567, Figure 3470: DNA325600, NM_015438, gen.XM_058567 gen.NM_015438 Figure 3505: PRO82107 Figure 3471: PRO82093 Figure 3506A-B: DNA325617, XM_166605, Figure 3472: DNA325601, XM_033263, gen.XM_166605 gen.XM_033263 Figure 3507: DNA325618, XM_029805, Figure 3473: DNA225632, NM_002046,

gen.XM_029805

Figure 3508: PRO82109 Figure 3543: DNA325636, XM_012272, Figure 3509: DNA325619, NM _005889, gen.XM_012272 gen.NM_005889 Figure 3544: PRO82127 Figure 3545A-B: DNA325637, XM_056481, Figure 3510: PRO82110 Figure 3511: DNA256072, NM_001644, gen.XM_056481 Figure 3546: DNA325638, NM_006262, gen.NM_001644 gen.NM_006262 Figure 3512: PRO51121 Figure 3513: DNA325620, NM_018686, Figure 3547: PRO82129 Figure 3548: DNA325639, NM_018113, gen.NM_018686 gen.NM_018113 Figure 3514: PRO82111 Figure 3549: PRO82130 Figure 3515: DNA325621, XM_084770, Figure 3550: DNA271344, NM_001659, gen.XM_084770. Figure 3516: PRO82112 gen.NM_001659 Figure 3551: PRO59647 Figure 3517: DNA325622, NM_018048, Figure 3552: DNA325640, NM _017822, gen.NM_018048 gen.NM_017822 Figure 3518: PRO82113 Figure 3553: PRO82131 Figure 3519: DNA325623, XM_113730, Figure 3554A-E: DNA325641, XM_028760, gen.XM_113730 gen.XM_028760 Figure 3520: DNA150978, NM_007244, Figure 3555: DNA272379, NM_002733, gen.NM_007244 Figure 3521: PRO11601 gen.NM_002733 Figure 3556: PRO60634 Figure 3522: DNA325624, NM_006250, Figure 3557: DNA325642, XM_084866, gen.NM_006250 gen.XM_084866 Figure 3523: PRO82115 Figure 3524: DNA79313, NM _005042, Figure 3558: PRO82133 Figure 3559: DNA325643, XM_006826, gen.NM_005042 Figure 3525: PRO2555 gen.XM_006826 Figure 3560: DNA325644, XM_113719, Figure 3526: DNA150997, NM _004982, gen.XM_113719 gen.NM_004982 Figure 3561: DNA325645, XM_028662, Figure 3527: PRO12573 gen.XM_028662 Figure 3528: DNA325625, XM_050074, Figure 3562: DNA325646, XM_035497, gen.XM_050074 gen.XM_035497 Figure 3529: DNA325626, NM_024854, Figure 3563: PRO82137 gen.NM_024854 Figure 3564: DNA325647, XM_035490, Figure 3530: PRO82117 gen.XM_035490 Figure 3531: DNA325627, XM_084807, gen.XM_084807 Figure 3565: PRO82138 Figure 3532: DNA325628, XM_165906, Figure 3566: DNA325648, NM_013277, gen.NM_013277 gen.XM_165906 Figure 3567: PRO82139 Figure 3533A-B: DNA325629, XM_038659, Figure 3568: DNA325649, NM_003076, gen.XM_038659 gen.NM_003076 Figure 3534: PRO82120 Figure 3535: DNA325630, XM_006694, Figure 3569: PRO82140 Figure 3570: DNA325650, XM_115117, gen.XM_006694 Figure 3536: DNA325631, XM_006748, gen.XM_115117 Figure 3571: DNA325651, XM_035485, gen.XM_006748 Figure 3537: PRO82122 gen.XM_035485 Figure 3538: DNA325632, XM_016640, Figure 3572A-B: DNA325652, NM_016357, gen.XM.016640 gen.NM_016357 Figure 3539: DNA325633, XM_096146, Figure 3573: PRO82143 Figure 3574: DNA325653, NM_005171, gen.XM_096146 Figure 3540A-B: DNA325634, XM_084841, gen.NM_005171 Figure 3575: PRO60924 gen.XM_084841 Figure 3541: PRO82125 Figure 3576: DNA325654, NM_014033, Figure 3542: DNA325635, XM_090218, gen.NM_014033 gen.XM_090218 Figure 3577: PRO4348

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Figure 3578: DNA325655, XM_096620,	Figure 3611: PRO82162
gen.XM_096620	Figure 3612: DNA325674, NM_031157,
Figure 3579: DNA325656, XM_165905,	gen.NM_031157
gen.XM_165905	Figure 3613: PRO82163
Figure 3580: DNA325657, XM_015481,	Figure 3614: DNA325675, NM _004178,
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Figure 3581: DNA325658, XM_049148,	Figure 3615: PRO82164
gen.XM_049148	Figure 3616: DNA325676, NM_134323,
Figure 3582: DNA325659, XM_084885,	gen.NM_134323
gen.XM_084885	Figure 3617: PRO82165
Figure 3583: DNA325660, XM _084884,	Figure 3618: DNA325677, NM_134324,
gen.XM_084884	gen.NM_134324
Figure 3584: DNA325661, XM_113726,	Figure 3619: PRO82166
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Figure 3585: DNA325662, XM_015476,	gen.NM_005016
gen.XM_015476	Figure 3621: PRO70453
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gen.XM_049141	gen.NM_031989
Figure 3587: PRO82152	Figure 3623: PRO82167
Figure 3588: DNA227191, NM_021934,	Figure 3624: DNA325679, XM_028643,
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Figure 3589: PRO37654	Figure 3625: PRO82168
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Figure 3591: DNA270458, NM_002273,	Figure 3627: PRO82169
gen.NM_002273	Figure 3628: DNA227094, NM _005594,
Figure 3592: PRO58837	gen.NM_005594
Figure 3593: DNA227092, NM_000224,	Figure 3629: PRO37557
gen.NM_000224	Figure 3630: DNA325681, XM_084824,
Figure 3594: PRO37555	gen.XM_084824
Figure 3595: DNA325665, XM _029728,	Figure 3631: DNA304783, NM _014255,
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Figure 3596: DNA325666, XM_015468,	Figure 3632: PRO4426
gen.XM_015468	Figure 3633: DNA325682, XM_165903,
Figure 3597: PRO82155	gen.XM_165903
Figure 3598: DNA325667, XM_012162,	Figure 3634: DNA325683, XM_115140,
gen.XM_012162	gen.XM_115140
Figure 3599: DNA325668, XM_084789,	Figure 3635: DNA325684, XM_113712,
gen.XM_084789	
Figure 3600: DNA196351, NM _002178,	gen.XM_113712 Figure 3636: DNA325685, NM_006601,
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Figure 3601: PRO3449	Figure 3637: PRO82174
Figure 3602A-B: DNA325669, XM_029631,	Figure 3638: DNA325686, XM_012182,
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Figure 3603: PRO82158	Figure 3639: PRO82175
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gen.NM_015665	gen.XM_048943
Figure 3605: PRO82159	Figure 3641: DNA325688, XM_053164,
Figure 3606: DNA325671, NM_014311,	gen.XM_053164
gen.NM_014311	Figure 3642: DNA325689, XM_048991,
Figure 3607: PRO82160	gen.XM_048991
Figure 3608: DNA325672, XM _096606,	Figure 3643: DNA325690, NM_024068,
gen.XM_096606	gen.NM_024068
Figure 3609: PRO82161	Figure 3644: PRO82179
Figure 3610: DNA325673, NM_018457,	Figure 3645A-B: DNA325691, XM_056346,
gen.NM_018457	gen.XM_056346

gen.NM_005981 Figure 3646: DNA325692, NM_021019, gen.NM_021019 Figure 3682: PRO4666 Figure 3683: DNA325711, NM_000075, Figure 3647: PRO82181 gen.NM_000075 Figure 3648: DNA325693, NM_079423, Figure 3684: PRO4873 gen.NM_079423 Figure 3649: PRO82182 Figure 3685: DNA325712, NM_052984, Figure 3650: DNA325694, NM_079425, gen.NM_052984 Figure 3686: PRO82194 gen.NM_079425 Figure 3687: DNA325713, NM_000785, Figure 3651: PRO82183 Figure 3652: DNA325695, XM_049048, gen.NM_000785 Figure 3688: PRO58440 gen.XM_049048 Figure 3689: DNA325714, NM _005371, Figure 3653: PRO82184 gen.NM_005371 Figure 3654: DNA325696, NM_021104, Figure 3690: PRO82195 gen.NM_021104 Figure 3655: PRO11213 Figure 3691: DNA325715, NM_023032, gen.NM_023032 Figure 3656: DNA325697, NM -001029, Figure 3692: PRO82196 gen.NM_001029 Figure 3657: PRO10838 Figure 3693: DNA325716, NM .023033, gen.NM_023033 Figure 3658: DNA325698, XM_001482, gen.XM_001482 Figure 3694: PRO82197 Figure 3659: DNA325699, XM_049150, Figure 3695: DNA325717, NM_005726, gen.NM_005726 gen.XM_049150 Figure 3660: DNA325700, NM_006928, Figure 3696: PRO82198 Figure 3697: DNA325718, NM_006576, gen.NM_006928 Figure 3661: PRO2846 gen.NM_006576 Figure 3698: PRO82199 Figure 3662: DNA325701, XM_056353, Figure 3699A-B: DNA325719, XM_096038, gen.XM_056353 Figure 3663: DNA325702, NM_001780, gen.XM_096038 Figure 3700: DNA325720, XM_056681, gen.NM_001780 Figure 3664: PRO283 gen.XM_056681 Figure 3665: DNA325703, NM_031479, Figure 3701: PRO82201 Figure 3702: DNA325721, XM_084909, gen.NM_031479 gen.XM_084909 Figure 3666: PRO21773 Figure 3703: PRO82202 Figure 3667A-: DNA137231, NM_005269, Figure 3704: DNA325722, XM_004098, gen.NM_005269 Figure 3668: PRO9112 gen.XM_004098 Figure 3705: DNA325723, XM_084912, Figure 3669: DNA325704, NM_004990, gen.NM_004990 gen.XM_084912 Figure 3670: PRO82188 Figure 3706: PRO82204 Figure 3707: DNA325724, XM_040221, Figure 3671: DNA325705, XM_058528, gen.XM_058528 gen.XM_040221 Figure 3672: DNA325706, XM .084801, Figure 3708: DNA325725, XM _016605, gen.XM_016605 gen.XM_084801 Figure 3709: PRO82206 Figure 3673: PRO82190 Figure 3674: DNA325707, XM_048603, Figure 3710: DNA325726, XM_017508, gen.XM_017508 gen.XM_048603 Figure 3711: PRO82207 Figure 3675: PRO82191 Figure 3676: DNA325708, NM_133483, Figure 3712: DNA325727, NM_032338, gen.NM_133483 gen.NM_032338 Figure 3677: PRO82192 Figure 3713: PRO82208 Figure 3714A-B: DNA325728, XM_052460, Figure 3678: DNA79101, NM_006812, gen.XM_052460 gen.NM_006812 Figure 3679: PRO2549 Figure 3715: DNA325729, XM_083866, Figure 3680: DNA325709, XM_096566, gen.XM_083866 gen.XM_096566 Figure 3716: PRO82210 Figure 3681: DNA325710, NM_005981, Figure 3717: DNA304694, NM_020401,

Figure 3753: PRO9987 gen.NM_020401 Figure 3754: DNA325747, XM_167518, Figure 3718: PRO71120 gen.XM_167518 Figure 3719: DNA325730, XM _052474, Figure 3755: DNA325748, XM_052542, gen.XM_052474 Figure 3720: DNA227474, NM_015646, gen.XM_052542 Figure 3756: PRO82223 gen.NM_015646 Figure 3757: DNA325749, NM _003877, Figure 3721: PRO37937 Figure 3722: DNA325731, XM_053952, gen.NM_003877 Figure 3758: PRO12839 gen.XM_053952 Figure 3759: DNA325750, XM_012219, Figure 3723: PRO82212 gen.XM_012219 Figure 3724: DNA227171, NM _014515, Figure 3760: PRO69473 gen.NM_014515 Figure 3761: DNA325751, XM_012145, Figure 3725: PRO37634 gen.XM_012145 Figure 3726: DNA325732, XM_046041, Figure 3762: PRO82224 gen.XM_046041 Figure 3763: DNA274361, NM_000895, Figure 3727: DNA271492, NM _006530, gen.NM_000895 gen.NM_006530 Figure 3764: PRO62273 Figure 3728: PRO59785 Figure 3765: DNA325752, XM_006887, Figure 3729: DNA226014, NM_000239, gen.XM_006887 gen.NM_000239 Figure 3766: DNA325753, XM_006589, Figure 3730: PRO36477 gen.XM_006589 Figure 3731: DNA325733, XM _084645, Figure 3767: DNA325754, XM_090458, gen.XM_084645 gen.XM_090458 Figure 3732A-B: DNA325734, XM_039395, Figure 3768: PRO82227 gen.XM_039395 Figure 3769: DNA325755, XM_052641, Figure 3733: PRO82213 gen.XM_052641 Figure 3734: DNA325736, XM_040644, Figure 3770: PRO82228 gen.XM_040644 Figure 3771A-B: DNA325756, XM_049211, Figure 3735: PRO82214 gen.XM_049211 Figure 3736A-B: DNA325737, XM _006578, Figure 3772: DNA325757, XM _049201, gen.XM_006578 gen.XM_049201 Figure 3737: DNA325738, XM _038308, Figure 3773: DNA325758, XM_058556, gen.XM_038308 gen.XM_058556 Figure 3738: PRO82215 Figure 3774: DNA325759, XM_083864, Figure 3739: DNA325739, XM _096597, gen.XM_083864 gen.XM_096597 Figure 3775: DNA325760, XM_062437, Figure 3740: DNA325740, NM_001920, gen.XM_062437 gen.NM_001920 Figure 3776: PRO82232 Figure 3741: PRO2841 Figure 3777: DNA254777, NM_014325, Figure 3742: DNA325741, NM_133503, gen.NM_014325 gen.NM_133503 Figure 3778: PRO49875 Figure 3743: PRO2841 Figure 3779: DNA325761, XM_090413, Figure 3744: DNA325742, NM_133504, gen.XM_090413 gen.NM_133504 Figure 3780: PRO82233 Figure 3745: PRO82218 Figure 3781: DNA325762, NM_000970, Figure 3746: DNA325743, NM_133505, gen.NM_000970 gen.NM_133505 Figure 3782: PRO82234 Figure 3747: PRO82219 Figure 3783: DNA325763, XM_084800, Figure 3748: DNA325744, NM_133507, gen.XM_084800 gen.NM_133507 Figure 3784: PRO82235 Figure 3749: PRO82220 Figure 3785: DNA325764, NM_006817, Figure 3750: DNA325745, NM_133506, gen.NM_006817 gen.NM_133506 Figure 3786: PRO70694 Figure 3751: PRO82221 Figure 3787A-C: DNA325765, XM_083892, Figure 3752: DNA325746, NM _002345, gen.XM_083892 gen.NM_002345

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Figure 4167A-B: DNA325986, XM_007531. gen.XM_085204 gen.XM_007531 Figure 4133: DNA325967, XM_012398, Figure 4168: DNA325987, NM_014444, gen.XM_012398 gen.NM_014444 Figure 4134A-B: DNA325968, XM_036727, Figure 4169: PRO82431 gen.XM_036727 Figure 4170A-B: DNA227206, NM _005657, Figure 4135: DNA325969, XM_017240, gen.NM_005657 gen.XM_017240 Figure 4171: PRO37669 Figure 4136: DNA325970, NM_020149, Figure 4172: DNA325988, NM_020990, gen.NM_020149 gen.NM_020990 Figure 4137: PRO82415 Figure 4173: PRO82432 Figure 4138A-B: DNA325971, XM_031617, Figure 4174: DNA325989, NM_005313, gen.XM_031617 gen.NM_005313 Figure 4139A-B: DNA325972, NM _001211, Figure 4175: PRO2732 gen.NM_001211 Figure 4176: DNA325990, NM_005770, Figure 4140: PRO82417 gen.NM_005770 Figure 4141A-B: DNA151831, NM _004573, Figure 4177: PRO82433 gen.NM_004573 Figure 4178: DNA325991, NM_004048, Figure 4142: PRO12198 gen.NM_004048 Figure 4143: DNA325973, NM_130468, Figure 4179: PRO4379 gen.NM_130468 Figure 4180: DNA325992, XM_032403, Figure 4144: PRO82418 gen.XM_032403 Figure 4145: DNA325974, XM_031554, Figure 4181: PRO82434 gen.XM_031554 Figure 4182: DNA219233, NM _014335, Figure 4146: PRO82419 gen.NM_014335 Figure 4147: DNA325975, XM_031515, Figure 4183: PRO34557 gen.XM_031515 Figure 4184A-C: DNA325993, XM _034890, Figure 4148: DNA325976, NM_024111, gen.XM_034890 gen.NM_024111 Figure 4185: PRO82435 Figure 4149: PRO82421 Figure 4186: DNA325994, XM_058684, Figure 4150: DNA325977, NM_032196, gen.XM_058684 gen.NM_032196 Figure 4187: DNA325995, NM_003104, Figure 4151: PRO82422 gen.NM_003104 Figure 4152: DNA325978, NM_016359, Figure 4188: PRO82437 gen.NM_016359 Figure 4189: DNA325996, XM_007651, Figure 4153: PRO82423 gen.XM_007651 Figure 4154: DNA325979, NM_018454, Figure 4190: PRO82438 gen.NM_018454 Figure 4191: DNA325997, XM_090991, Figure 4155: PRO82424 gen.XM_090991 Figure 4156A-B: DNA325980, XM_007545, Figure 4192: PRO82439 gen.XM_007545 Figure 4193: DNA325998, NM_016304, Figure 4157: DNA325981, XM_091159, gen.NM_016304 gen.XM_091159 Figure 4194: PRO82440 Figure 4158: PRO82425 Figure 4195: DNA325999, NM_017610, Figure 4159: DNA325982, XM_031718, gen.NM_017610 gen.XM_031718 Figure 4196: PRO82441 Figure 4160: DNA325983, XM_085307, Figure 4197: DNA326000, NM_004701, gen.XM_085307 gen.NM_004701 Figure 4161: DNA227559, NM ..000070, Figure 4198: PRO82442 gen.NM_000070 Figure 4199A-B: DNA326001, XM _012418, Figure 4162: PRO38022 gen.XM_012418 Figure 4163A-B: DNA325984, XM_113823, Figure 4200: DNA326002, XM_039702, gen.XM_113823 gen.XM_039702 Figure 4164: PRO82428 Figure 4201: PRO82444 Figure 4165: DNA325985, XM_016713, Figure 4202: DNA326003, XM_113266, gen.XM_016713 gen.XM_113266 Figure 4166: PRO82429

Figure 4203: DNA326004, NM_001218, Figure 4238: PRO82460 gen.NM_001218 Figure 4239: DNA326022, XM_015366, Figure 4204: PRO54594 gen.XM_015366 Figure 4205: DNA326005, NM_015920, Figure 4240: PRO82461 gen.NM_015920 Figure 4241: DNA326023, XM_096060, Figure 4206: PRO82446 gen.XM_096060 Figure 4207: DNA326006, XM_113268, Figure 4242: DNA287331, NM_002654, gen.XM_113268 gen.NM_002654 Figure 4208: DNA255340, NM_017684, Figure 4243: PRO69595 gen.NM_017684 Figure 4244: DNA326024, XM_037778, Figure 4209: PRO50409 gen.XM_037778 Figure 4210: DNA326007, NM_002537, Figure 4245: DNA326025, XM_096842, gen.NM_002537 gen.XM_096842 Figure 4211: DNA326008, XM_085283, Figure 4246: DNA326026, NM_022369, gen.XM_085283 gen.NM_022369 Figure 4212: PRO82448 Figure 4247: PRO82465 Figure 4213: DNA326009, XM_016985, Figure 4248: DNA326027, NM_032907, gen.XM_016985 gen.NM_032907 Figure 4214: DNA234442, NM_014736, Figure 4249: PRO82466 gen.NM_014736 Figure 4250: DNA326028, XM_058699, Figure 4215: PRO38852 gen.XM_058699 Figure 4216: DNA326010, NM_022048, Figure 4251: DNA326029, XM_118637. gen.NM_022048 gen.XM_118637 Figure 4217: PRO82450 Figure 4252: DNA326030, XM_053585, Figure 4218: DNA326011, NM_000942, gen.XM_053585 gen.NM_000942 Figure 4253: PRO82469 Figure 4219: PRO2720 Figure 4254: DNA326031, XM_085239, Figure 4220: DNA326012, XM_050964, gen.XM_085239 gen.XM_050964 Figure 4255: PRO82470 Figure 4221: DNA326013, XM_007623, Figure 4256: DNA326032, XM_034897, gen.XM_007623 gen.XM_034897 Figure 4222A-B: DNA326014, NM_133375, Figure 4257A-B: DNA326033, XM_057020, gen.NM_133375 gen.XM_057020 Figure 4223: PRO82453 Figure 4258: PRO82472 Figure 4224: DNA226646, NM_017882, Figure 4259: DNA326034, NM_000743, gen.NM_017882 gen.NM_000743 Figure 4225: PRO37109 Figure 4260: PRO61219 Figure 4226: DNA326015, NM_015322, Figure 4261: DNA326035, NM_002789, gen.NM_015322 gen.NM_002789 Figure 4227: PRO82454 Figure 4262: PRO60499 Figure 4228: DNA326016, NM_001003, Figure 4263: DNA326036, XM_091100, gen.NM_001003 gen.XM_091100 Figure 4229: PRO82455 Figure 4264: PRO82473 Figure 4230A-B: DNA326017, XM_051463, Figure 4265: DNA255370, NM_012170, gen.XM_051463 gen.NM_012170 Figure 4231: PRO82456 Figure 4266: PRO50438 Figure 4232: DNA326018, NM_018357. Figure 4267: DNA273014, NM_000126, gen.NM_018357 gen.NM_000126 Figure 4233: PRO82457 Figure 4268: PRO61085 Figure 4234: DNA326019, XM_063639, Figure 4269: DNA326037, XM_044565, gen.XM_063639 gen.XM_044565 Figure 4235: PRO82458 Figure 4270: DNA326038, NM_025234, Figure 4236: DNA326020, XM_085249, gen.NM_025234 gen.XM_085249 Figure 4271: PRO82475 Figure 4237: DNA326021, XM_016076, Figure 4272: DNA326039, XM_044569, gen.XM_016076 gen.XM_044569

Figure 4273: DNA326040, NM_005724, Figure 4307A-B: DNA326060, XM_044533, gen.XM_044533 gen.NM_005724 Figure 4308: PRO82495 Figure 4274: PRO730 Figure 4275: DNA326041, XM_049354, Figure 4309A-C: DNA326061, XM_054900, gen.XM_049354 gen.XM_054900 Figure 4310: DNA326062, NM_032162, Figure 4276: PRO82477 Figure 4277: DNA326042, NM_007364, gen.NM_032162 Figure 4311A-B: DNA326063, XM_015835, gen.NM_007364 Figure 4278: DNA326043, XM_044593, gen.XM_015835 gen.XM_044593 Figure 4312: DNA326064, NM_018668, gen.NM_018668 Figure 4279: DNA326044, NM_006791, Figure 4313: PRO82499 gen.NM_006791 Figure 4314: DNA326065, XM_085262, Figure 4280: PRO82479 gen.XM_085262 Figure 4281: DNA326045, XM_060042, Figure 4315: DNA326066, NM _033544, gen.XM_060042 Figure 4282: DNA326046, XM_085215, gen.NM_033544 Figure 4316: PRO82501 gen.XM_085215 Figure 4317: DNA326067, XM_049372, Figure 4283: DNA326047, NM_001021, gen.XM_049372 gen.NM_001021 Figure 4318: PRO82502 Figure 4284: PRO82482 Figure 4319: DNA326068, XM_017971, Figure 4285: DNA326048, XM_031404, gen.XM_017971 gen.XM_031404 Figure 4320: DNA275181, NM_003090, Figure 4286: DNA326049, XM_096844, gen.NM_003090 gen.XM_096844 Figure 4321: PRO62882 Figure 4287: DNA326050, XM_045681, Figure 4322: DNA326069, XM_012462, gen.XM_045681 gen.XM_012462 Figure 4288: PRO82485 Figure 4323A-B: DNA326070, XM_085525, Figure 4289: DNA326051, XM_085280, gen.XM_085280 gen.XM_085525 Figure 4290: DNA326052, NM_022839, Figure 4324: PRO82505 gen.NM_022839 Figure 4325: DNA326071, XM_165923, Figure 4291: PRO82487 gen.XM_165923 Figure 4326: DNA326072, XM_113836, Figure 4292: DNA326053, XM_031354, gen.XM_113836 gen.XM_031354 Figure 4327: DNA326073, NM_017668, Figure 4293: DNA326054, NM.002168, gen.NM_017668 gen.NM_002168 Figure 4328: PRO82508 Figure 4294: PRO82489 Figure 4329: DNA326074, XM_027309, Figure 4295: DNA326055, XM_031292, gen.XM_027309 gen.XM_031292 Figure 4330: PRO82509 Figure 4296: DNA326056, NM_022566, Figure 4331: DNA326075, XM_018432, gen.NM_022566 Figure 4297: PRO82491 gen.XM_018432 Figure 4298A-B: DNA326057, XM_051860, Figure 4332: PRO82510 Figure 4333: DNA326076, XM_115352, gen.XM_051860 Figure 4299: PRO82492 gen.XM_115352 Figure 4334: DNA326077, XM _027365, Figure 4300: DNA275144, NM_000137, gen.XM_027365 gen.NM_000137 Figure 4335: DNA326078, NM_016641, Figure 4301: PRO62852 Figure 4302: DNA326058, NM_016645, gen.NM_016641 Figure 4336: PRO38464 gen.NM_016645 Figure 4337: DNA326079, XM_058796, Figure 4303: PRO82493 gen.XM_058796 Figure 4304: DNA326059, XM_044523, Figure 4338: DNA326080, XM_017984, gen.XM_044523 gen.XM_017984 Figure 4305: DNA150485, NM _006384, Figure 4339: PRO82513 gen.NM_006384 Figure 4340: DNA326081, NM .020677, Figure 4306: PRO12774

gen.NM_020677 Figure 4377: PRO82524 Figure 4341: PRO82514 Figure 4378: DNA326097, NM_023936, Figure 4342: DNA326082, XM_036680, gen.NM_023936 gen.XM_036680 Figure 4379: PRO82525 Figure 4343: PRO37961 Figure 4380: DNA326098, XM_034590, Figure 4344A-B: DNA326083, XM_048119, gen.XM_034590 gen.XM_048119 Figure 4381: PRO82526 Figure 4345: PRO82515 Figure 4382: DNA326099, NM_002952, Figure 4346: DNA326084, NM_024589, gen.NM_002952 gen.NM_024589 Figure 4383: PRO82527 Figure 4347: PRO82516 Figure 4384: DNA326100, NM_006453, Figure 4348: DNA326085, XM_050534, gen.NM_006453 gen.XM_050534 Figure 4385: PRO82528 Figure 4349: PRO82517 Figure 4386: DNA326101, NM_014353, Figure 4350: DNA326086, NM_024571, gen.NM_014353 gen.NM_024571 Figure 4387: PRO82529 Figure 4351: PRO82518 Figure 4388: DNA326102, NM_032271, Figure 4352: DNA326087, XM_027558, gen.NM_032271 gen.XM_027558 Figure 4389: PRO82530 Figure 4353: DNA326088, XM_008126, Figure 4390: DNA326103, XM_028848, gen.XM_008126 gen.XM_028848 Figure 4354: DNA326089, NM _000517. Figure 4391: PRO82531 gen.NM_000517 Figure 4392: DNA326104, NM_006711, Figure 4355: PRO3629 gen.NM_006711 Figure 4356: DNA326090, NM_000558, Figure 4393: PRO82532 gen.NM_000558 Figure 4394: DNA326105, NM_080594, Figure 4357: PRO3629 gen.NM_080594 Figure 4358: DNA326091, NM_018032, Figure 4395: PRO82533 gen.NM_018032 Figure 4396: DNA326106, NM_024339, Figure 4359: PRO38311 gen.NM_024339 Figure 4360: DNA273839, NM_006428, Figure 4397: PRO82534 gen.NM_006428 Figure 4398: DNA326107, NM_016639, Figure 4361: PRO61799 gen.NM_016639 Figure 4362A-B: DNA256844, NM_005632, Figure 4399: PRO12683 gen.NM_005632 Figure 4400: DNA326108, NM_021195, Figure 4363: PRO51775 gen.NM_021195 Figure 4364: DNA326092, XM_083939, Figure 4401: PRO82535 Figure 4402: DNA326109, NM_004203, gen.XM_083939 Figure 4365: PRO82521 gen.NM_004203 Figure 4366: DNA326093, NM_058192, Figure 4403: PRO82536 gen.NM_058192 Figure 4404: DNA326110, XM_058784, Figure 4367: PRO82522 gen.XM_058784 Figure 4368: DNA326094, XM_027412, Figure 4405: PRO82537 gen.XM_027412 Figure 4406: DNA326111, NM_024507, Figure 4369: PRO82523 gen.NM_024507 Figure 4370: DNA256886, NM_014587, Figure 4407: PRO82538 gen.NM_014587 Figure 4408: DNA326112, NM_006799, Figure 4371: PRO51815 gen.NM_006799 Figure 4372A-B: DNA326095, NM_001287, Figure 4409: PRO303 gen.NM_001287 Figure 4410A-C: DNA326113, XM_036528, Figure 4373: PRO38480 gen.XM_036528 Figure 4374: DNA254781, NM_016111, Figure 4411: DNA326114, NM_025108, gen.NM_016111 gen.NM_025108 Figure 4375: PRO49879 Figure 4412: PRO82540 Figure 4376: DNA326096, XM_034586, Figure 4413A-C: DNA326115, XM_165411, gen.XM_034586 gen.XM_165411

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gen.XM_017096	gen.XM_028417
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gen.XM_008557	Figure 4686: DNA326266, XM_008441,
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gen.NM_080822	Figure 4690: PRO82674
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Figure 4658: DNA326250, XM 2008509,	gen.XM_008679
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Figure 4797: PRO1204

Figure 4798: DNA326324, NM_000981, gen.NM_000981 Figure 4799: PRO4738 Figure 4800A-B: DNA326325, XM_008150, gen.XM_008150 Figure 4801: DNA326326, NM_000978, gen.NM_000978 Figure 4802: PRO82724 Figure 4803: DNA326327, XM_058830, gen.XM_058830 Figure 4804: PRO82725 Figure 4805: DNA270979, NM_002809, gen.NM_002809 Figure 4806: PRO59309 Figure 4807: DNA326328, NM_000422, gen.NM_000422 Figure 4808: PRO82726 Figure 4809: DNA326329, XM_008579, gen.XM_008579 Figure 4810: DNA326330, NM_002276, gen.NM_002276 Figure 4811: PRO82728 Figure 4812: DNA272889, NM _002275, gen.NM_002275 Figure 4813: PRO60979 Figure 4814: DNA326331, NM_002274, gen.NM_002274 Figure 4815: PRO82729 Figure 4816: DNA326332, NM .000526, gen.NM_000526 Figure 4817: PRO82730 Figure 4818: DNA326333, XM_049937, gen.XM_049937 Figure 4819A-B: DNA326334, XM_113334, gen.XM_113334 Figure 4820: DNA226389, NM _000964, gen.NM_000964 Figure 4821: PRO36852 Figure 4822: DNA326335, NM_006455, gen.NM_006455 Figure 4823: PRO82732 Figure 4824: DNA326336, XM_113938, gen.XM_113938 Figure 4825: DNA326337, XM _036465, gen.XM_036465 Figure 4826: DNA326338, XM_055061, gen.XM_055061 Figure 4827A-B: DNA326339, XM_036462, gen.XM_036462 Figure 4828: PRO82736 Figure 4829: DNA326340, XM _048654, gen.XM_048654 Figure 4830: DNA326341, NM_025197, gen.NM_025197 Figure 4831: PRO82737 Figure 4832: DNA326342, XM_054038,

Figure 4868: PRO82754 gen.XM_054038 Figure 4869: DNA326359, XM_008402, Figure 4833: PRO82738 gen.XM_008402 Figure 4834: DNA326343, NM _002265, Figure 4870: PRO82755 gen.NM_002265 Figure 4871: DNA326360, NM_017595, Figure 4835: PRO82739 gen.NM_017595 Figure 4836: DNA326344, XM_032201, Figure 4872: PRO82756 gen.XM_032201 Figure 4873: DNA326361, XM_085636, Figure 4837: PRO82740 gen.XM_085636 Figure 4838: DNA326345, NM_012138, Figure 4874: PRO82757 gen.NM_012138 Figure 4875: DNA326362, NM_006373, Figure 4839: PRO82741 gen.NM_006373 Figure 4840: DNA326346, XM_018534, Figure 4876: PRO82758 gen.XM_018534 Figure 4877: DNA196642, NM_005440, Figure 4841: DNA227873, NM_001050, gen.NM_005440 gen.NM_001050 Figure 4878: PRO25115 Figure 4842: PRO38336 Figure 4879A-B: DNA270901, NM_004247, Figure 4843: DNA270975, NM_000386, gen.NM_000386 gen.NM_004247 Figure 4880: DNA326363, XM_050159, Figure 4844: PRO59305 gen.XM_050159 Figure 4845: DNA88378, NM_002087, Figure 4881: DNA326364, XM_083983, gen.NM_002087 gen.XM.083983 Figure 4846: PRO2769 Figure 4882: PRO82760 Figure 4847: DNA326347, NM_016016, Figure 4883A-B: DNA326365, NM_021079, gen.NM_016016 gen.NM_021079 Figure 4848: PRO82743 Figure 4884: PRO82761 Figure 4849: DNA326348, XM_012642, Figure 4885: DNA326366, NM_133373, gen.XM_012642 gen.NM_133373 Figure 4850A-B: DNA326349, NM_005474, gen.NM_005474 Figure 4886: PRO82762 Figure 4887: DNA97290, NM_002512, Figure 4851: PRO82745 gen.NM_002512 Figure 4852: DNA326350, XM_045901, Figure 4888: PRO3637 gen.XM_045901 Figure 4889: DNA227071, NM_000269, Figure 4853: PRO82746 gen.NM_000269 Figure 4854: DNA257428, NM ..032376, Figure 4890: PRO37534 gen.NM_032376 Figure 4891: DNA227764, NM_003971, Figure 4855: PRO52010 gen.NM_003971 Figure 4856: DNA326351, XM_008351, Figure 4892: PRO38227 gen.XM_008351 Figure 4893A-B: DNA326367, NM_020038, Figure 4857: DNA326352, XM_032852, gen.NM_020038 gen.XM_032852 Figure 4894: PRO82763 Figure 4858: PRO82748 Figure 4895A-B: DNA326368, NM_020037, Figure 4859: DNA326353, NM _025233, gen.NM_025233 gen.NM_020037 Figure 4896: PRO82764 Figure 4860: PRO82749 Figure 4861: DNA326354, XM_032817, Figure 4897: DNA326369, XM_037971, gen.XM_037971 gen.XM_032817 Figure 4898: DNA254791, NM_018346, Figure 4862: PRO82750 gen.NM_018346 Figure 4863: DNA326355, XM_032813, Figure 4899: PRO49888 gen.XM_032813 Figure 4900: DNA287425, NM_018509, Figure 4864: DNA326356, XM_032766, gen.NM_018509 gen.XM_032766 Figure 4901: PRO69682 Figure 4865: DNA326357, NM_003766, Figure 4902A-B: DNA326370, XM_008432, gen.NM_003766 gen.XM_008432 Figure 4866: PRO82753 Figure 4903: DNA88554, NM_000250, Figure 4867: DNA326358, XM_008401, gen.NM_000250 gen.XM_008401

gen.XM_044367

Figure 4939: DNA227055, NM_002634, Figure 4904: PRO2839 gen.NM_002634 Figure 4905: DNA326371, XM_113919, Figure 4940: PRO37518 gen.XM_113919 Figure 4941: DNA326390, XM_011118, Figure 4906: DNA326372, NM_017777, gen.XM_011118 gen.NM_017777 Figure 4942: DNA326391, XM_055199, Figure 4907: PRO82768 gen.XM_055199 Figure 4908: DNA326373, NM .006924, Figure 4943A-B: DNA326392, XM_044372, gen.NM_006924 gen.XM_044372 Figure 4909: PRO82769 Figure 4944: DNA326393, XM_113315, Figure 4910: DNA326374, XM_115480, gen.XM_115480 gen.XM_113315 Figure 4911: DNA326375, NM_005831, Figure 4945: DNA326394, XM_012609, gen.XM_012609 gen.NM_005831 Figure 4946: DNA326395, NM_005220, Figure 4912: PRO59328 gen.NM_005220 Figure 4913: DNA326376, XM_117061, Figure 4947: PRO82787 gen.XM_117061 Figure 4948: DNA326396, XM .085589, Figure 4914: PRO82771 gen.XM_085589 Figure 4915: DNA326377, XM _008459, Figure 4949: PRO82788 gen.XM_008459 Figure 4950: DNA326397, XM_012634, Figure 4916A-B: DNA326378, XM_012651, gen.XM_012634 gen.XM_012651 Figure 4951: DNA326398, XM_085627, Figure 4917: DNA326379, NM_021626, gen.XM_085627 gen.NM_021626 Figure 4952: PRO82790 Figure 4918: PRO302 Figure 4953: DNA150814, NM_002086, Figure 4919: DNA287291, NM _021213, gen.NM_002086 gen.NM_021213 Figure 4954: PRO12806 Figure 4920: PRO69561 Figure 4955: DNA326399, NM_024844, Figure 4921A-B: DNA326380, NM_004859, gen.NM_024844 gen_NM_004859 Figure 4956: PRO82791 Figure 4922: PRO82774 Figure 4957: DNA326400, XM_041583, Figure 4923: DNA326381, XM_083966, gen.XM_041583 gen.XM_083966 Figure 4958: DNA326401, XM_046932, Figure 4924: DNA326382, XM_044426, gen.XM_046932 gen.XM_044426 Figure 4959: PRO82792 Figure 4925: PRO82776 Figure 4960: DNA326402, NM_004524, Figure 4926: DNA326383, XM_008253, gen.NM_004524 gen.XM_008253 Figure 4927: DNA326384, XM_044394, Figure 4961: PRO82793 Figure 4962A-B: DNA326403, XM_113951, gen.XM_044394 gen.XM_113951 Figure 4928: PRO10400 Figure 4963A-B: DNA88430, NM_000213, Figure 4929: DNA326385, NM_017647, gen.NM_000213 gen.NM_017647 Figure 4964: PRO2788 Figure 4930: PRO82778 Figure 4965A-B: DNA326404, XM_036104, Figure 4931: DNA326386, NM _007372, gen.XM_036104 gen.NM_007372 Figure 4966: PRO82794 Figure 4932: PRO82779 Figure 4967: DNA326405, NM_000154, Figure 4933: DNA326387, NM _002401, gen.NM_000154 gen.NM_002401 Figure 4968: PRO82795 Figure 4934: PRO37764 Figure 4969: DNA326406, NM_005324, Figure 4935: DNA326388, XM_044376, gen.NM_005324 gen.XM_044376 Figure 4936A-B: DNA150457, NM_006039, Figure 4970: PRO11403 Figure 4971A-B: DNA326407, XM_036115, gen.NM_006039 gen.XM_036115 Figure 4937: PRO12265 Figure 4972: PRO82796 Figure 4938: DNA326389, XM_044367, Figure 4973: DNA326408, XM_054344,

gen.NM_016286

Figure 5007: PRO82813 gen.XM_054344 Figure 5008: DNA326429, NM_004127, Figure 4974: PRO82797 gen.NM_004127 Figure 4975: DNA274755, NM_002766, Figure 5009: PRO82814 gen.NM_002766 Figure 5010A-C: DNA326430, XM_113943, Figure 4976: PRO70703 gen.XM_113943 Figure 4977A-B: DNA326409, XM_085531, Figure 5011: DNA326431, XM_113330, gen.XM_085531 gen.XM_113330 Figure 4978: DNA326410, XM_113892, Figure 5012: PRO82816 gen.XM_113892 Figure 5013: DNA326432, XM_113303, Figure 4979: PRO82799 Figure 4980: DNA326411, XM_017578, gen.XM_113303 Figure 5014: DNA287234, NM .031968, gen.XM_017578 gen.NM_031968 Figure 4981: PRO82800 Figure 5015: PRO69513 Figure 4982: DNA326412, XM_036785, Figure 5016: DNA326433, NM _022158, gen.XM_036785 gen.NM_022158 Figure 4983: PRO39201 Figure 5017: PRO82818 Figure 4984: DNA326413, XM_097043, Figure 5018: DNA326434, XM_038424, gen.XM_097043 gen.XM_038424 Figure 4985: DNA129504, NM_001168, Figure 5019: DNA326435, XM .085735, gen.NM_001168 gen.XM_085735 Figure 4986: PRO7143 Figure 5020: DNA326436, XM_046765, Figure 4987: DNA326414, XM_037196, gen.XM_046765 gen.XM_037196 Figure 5021: DNA326437, XM_046769, Figure 4988: DNA326415, XM_037195, gen.XM_046769 gen.XM_037195 Figure 5022: DNA326438, XM_046767, Figure 4989: DNA326416, XM_045104, gen.XM_046767 gen.XM_045104 Figure 5023: DNA273694, NM _006101, Figure 4990: PRO37540 gen.NM_006101 Figure 4991: DNA326417, XM_085563, Figure 5024: PRO61661 gen.XM_085563 Figure 5025A-B: DNA326439, XM_028744, Figure 4992A-B: DNA326418, XM_085716, gen.XM_028744 gen.XM_085716 Figure 5026: DNA326440, XM_165954, Figure 4993: PRO82805 gen.XM_165954 Figure 4994A-B: DNA326419, XM_049934, Figure 5027: DNA326441, XM_041678, gen.XM_049934 Figure 4995: DNA326420, XM_049931, gen.XM_041678 Figure 5028: DNA326442, XM_113343, gen.XM_049931 gen.XM_113343 Figure 4996A-B: DNA326421, XM_045581, Figure 5029: PRO82825 gen.XM_045581 Figure 5030: DNA326443, XM_067325, Figure 4997: PRO82807 gen.XM_067325 Figure 4998: DNA326422, XM_113945, Figure 5031: DNA326444, XM_012741, gen.XM_113945 gen.XM_012741 Figure 4999: DNA326423, XM_046481, Figure 5032: DNA326445, NM_014214, gen.XM_046481 gen.NM_014214 Figure 5000: DNA326424, XM_097195, Figure 5033: PRO82828 gen.XM_097195 Figure 5034A-B: DNA326446, XM_035640, Figure 5001: DNA326425, XM_097193, gen.XM_035640 gen.XM_097193 Figure 5035: PRO82829 Figure 5002: DNA326426, NM_004309, Figure 5036: DNA326447, XM_016382, gen.NM_004309 gen.XM_016382 Figure 5003: PRO61246 Figure 5037: DNA326448, NM _032933, Figure 5004: DNA326427, XM_046472, gen.NM_032933 gen.XM_046472 Figure 5038: PRO82831 Figure 5005: PRO82812 Figure 5039: DNA274690, NM _006938, Figure 5006: DNA326428, NM_016286, gen.NM_006938

Figure 5040A-B: DNA88457, NM_000227, Figure 5074: DNA326467, XM_006937, gen.NM_000227 gen.XM_006937 Figure 5075: DNA326468, XM _085779, Figure 5041: PRO2799 gen.XM_085779 Figure 5042: DNA326449, XM_085791, Figure 5076: DNA326469, XM_011089, gen.XM_085791 gen.XM_011089 Figure 5043: DNA326450, XM_085789, Figure 5077: PRO82850 gen.XM_085789 Figure 5078: DNA326470, XM_169540, Figure 5044: PRO82833 Figure 5045: DNA326451, XM_085790, gen.XM_169540 Figure 5079: PRO82851 gen.XM_085790 Figure 5046: DNA326452, XM_015755, Figure 5080: DNA326471, XM_167008, gen.XM_167008 gen.XM_015755 Figure 5081: PRO82852 Figure 5047: PRO82835 Figure 5082: DNA326472, XM_048471, Figure 5048: DNA326453, XM .097232, gen.XM_097232 gen.XM_048471 Figure 5083A-B: DNA326473, XM .008812, Figure 5049: DNA326454, XM_085788, gen.XM_008812 gen.XM _085788 Figure 5084A-B: DNA326474, XM_117096, Figure 5050: DNA88281, NM_001944, gen.XM_117096 gen.NM_001944 Figure 5085: PRO82855 Figure 5051: PRO2267 Figure 5086: DNA326475, NM_002385, Figure 5052: DNA271841, NM_003787, gen.NM_002385 gen.NM_003787 Figure 5087: PRO82856 Figure 5053: PRO60121 Figure 5088: DNA326476, XM_015241, Figure 5054: DNA326455, XM_008723, gen.XM_015241 gen.XM_008723 Figure 5089A-B: DNA326477, XM_008695, Figure 5055: DNA326456, XM_084007, gen.XM_008695 gen.XM_084007 Figure 5090A-B: DNA326478, XM_041872, Figure 5056: DNA256813, NM_018255, gen.NM_018255 gen.XM_041872 Figure 5091: PRO82859 Figure 5057: PRO51744 Figure 5092: DNA326479, XM_051586, Figure 5058: DNA326457, XM_085775, gen.XM_051586 gen.XM_085775 Figure 5093: DNA326480, NM_003712, Figure 5059: PRO82840 gen.NM_003712 Figure 5060: DNA326458, NM_138443, Figure 5094: PRO1077 gen.NM_138443 Figure 5095: DNA326481, XM_042018, Figure 5061: PRO82841 gen.XM_042018 Figure 5062: DNA326459, XM_038872, Figure 5096: PRO2560 gen.XM_038872 Figure 5097: DNA326482, XM_114018, Figure 5063: PRO82842 gen.XM_114018 Figure 5064: DNA326460, XM_086779, Figure 5098: DNA326483, NM_017876, gen.XM_086779 Figure 5065: DNA326461, XM_167363, gen.NM_017876 Figure 5099: PRO82861 gen.XM_167363 Figure 5100: DNA326484, NM_031990, Figure 5066: DNA326462, XM_031944, gen.XM_031944 gen.NM_031990 Figure 5101: PRO82862 Figure 5067: DNA326463, NM _000985, Figure 5102: DNA326485, NM_002819, gen.NM_000985 gen.NM_002819 Figure 5068: PRO82846 Figure 5103: PRO62899 Figure 5069: DNA326464, NM_002396, Figure 5104: DNA326486, NM_005224, gen.NM_002396 gen.NM_005224 Figure 5070: PRO61113 Figure 5105: PRO82863 Figure 5071: DNA326465, XM_166288, Figure 5106: DNA326487, XM_037565, gen.XM_166288 gen.XM_037565 Figure 5072: DNA326466, NM_004539, Figure 5107: PRO82864 gen.NM_004539 Figure 5108: DNA326488, XM_092042, Figure 5073: PRO60800

gen.NM_001319

Figure 5142: PRO82881 gen.XM_092042 Figure 5143: DNA326510, NM_017797, Figure 5109: DNA326489, XM_037572, gen.NM_017797 gen.XM_037572 Figure 5144: PRO82882 Figure 5110: DNA326490, XM_009279, Figure 5145: DNA326511, XM_030714, gen.XM_009279 gen.XM_030714 Figure 5111: PRO82867 Figure 5146: DNA256555, NM_017572, Figure 5112: DNA326491, NM_002085, gen.NM_017572 gen.NM_002085 Figure 5147: PRO51586 Figure 5113A-B: DNA326492, XM .009277, Figure 5148A-B: DNA326512, NM_003938, gen.XM_009277 gen.NM_003938 Figure 5114: DNA326493, XM_012913, Figure 5149: PRO82884 gen.XM_012913 Figure 5150A-B: DNA326513, XM_046822, Figure 5115: DNA274101, NM_001687, gen.XM_046822 gen.NM_001687 Figure 5151: PRO82885 Figure 5116: PRO62039 Figure 5152: DNA326514, NM_007165, Figure 5117: DNA326494, XM_028067, gen.NM_007165 gen.XM_028067 Figure 5153: PRO82886 Figure 5118: PRO82871 Figure 5154: DNA287636, NM_004152, Figure 5119: DNA326495, XM_028064, gen.NM_004152 gen.XM_028064 Figure 5155: DNA326515, NM_012458, Figure 5120: DNA326496, NM_024407, gen.NM_012458 gen.NM_024407 Figure 5156: PRO82887 Figure 5121: PRO82872 Figure 5157: DNA326516, NM_032737, Figure 5122: DNA326497, NM_000156, gen.NM_032737 gen.NM_000156 Figure 5158: PRO82888 Figure 5123: PRO58046 Figure 5159: DNA326517, XM_030485, Figure 5124: DNA326498, NM_138924, gen.XM_030485 gen.NM_138924 Figure 5160: DNA326518, XM_046934, Figure 5125: PRO82873 gen.XM_046934 Figure 5126: DNA326499, NM_001018, Figure 5161: DNA326519, NM_003021, gen.NM_001018 gen.NM_003021 Figure 5127: PRO10485 Figure 5162: PRO62302 Figure 5128: DNA326500, XM_086101, Figure 5163: DNA326520, XM_055686, gen.XM_086101 gen.XM_055686 Figure 5129: PRO82874 Figure 5164: PRO37951 Figure 5130: DNA326501, XM_086102, Figure 5165: DNA326521, XM_009222, gen.XM_086102 gen.XM_009222 Figure 5131: DNA326502, XM_047584, Figure 5166: DNA326522, XM_052635, gen.XM_047584 gen.XM_052635 Figure 5132A-B: DNA326503, XM_047600, Figure 5167: PRO82892 gen.XM_047600 Figure 5168: DNA326523, XM_052661, Figure 5133: PRO38496 gen.XM_052661 Figure 5134: DNA326504, XM_097420, Figure 5169: DNA326524, NM_016263, gen.XM_097420 gen.NM_016263 Figure 5135A-B: DNA326505, XM_030721, Figure 5170: PRO82893 gen.XM_030721 Figure 5171: DNA326525, NM_006339, Figure 5136: PRO82877 gen.NM_006339 Figure 5137: DNA326506, XM_030720, Figure 5172: PRO82894 gen.XM_030720 Figure 5173: DNA326526, NM_032753, Figure 5138: DNA326507, NM .031213, gen.NM_032753 gen.NM_031213 Figure 5174: PRO82895 Figure 5139: PRO82879 Figure 5175: DNA326527, XM_056421, Figure 5140: DNA326508, XM_039723, gen.XM_056421 gen.XM_039723 Figure 5176A-B: DNA326528, XM_031917, Figure 5141: DNA326509, NM_001319,

gen.XM_031917

Figure 5177: PRO82897 gen.XM_012798 Figure 5213: DNA326548, XM _044608, Figure 5178: DNA326529, NM_001961, gen.XM_044608 gen.NM_001961 Figure 5214: DNA326549, NM_003624, Figure 5179: PRO62225 Figure 5180: DNA326530, XM_016871, gen.NM_003624 gen.XM_016871 Figure 5215: PRO82915 Figure 5181: DNA326531, NM 016539, Figure 5216: DNA326550, NM_016579, gen.NM_016539 gen.NM_016579 Figure 5217: PRO224 Figure 5182: PRO82899 Figure 5218A-B: DNA326551, XM_048351, Figure 5183: DNA326532, XM_117122, gen.XM_048351 gen.XM_117122 Figure 5219: DNA326552, XM_048364, Figure 5184: DNA326533, XM_031857, gen.XM_048364 gen.XM_031857 Figure 5220: PRO82917 Figure 5185: PRO82901 Figure 5221: DNA326553, XM_091938, Figure 5186: DNA326534, NM_024333, gen.XM_091938 gen.NM_024333 Figure 5187: PRO82902 Figure 5222: DNA326554, XM_097300, gen.XM_097300 Figure 5188: DNA326535, NM_003025, Figure 5223: DNA326555, XM_049282, gen.NM_003025 gen.XM_049282 Figure 5189: PRO82903 Figure 5224: PRO82920 Figure 5190: DNA326536, NM_025241, Figure 5225: DNA326556, XM_058232, gen.NM_025241 gen.XM_058232 Figure 5191: PRO82904 Figure 5226: DNA326557, XM_045151, Figure 5192: DNA326537, XM_035638, gen.XM_035638 gen.XM_045151 Figure 5227A-B: DNA326558, XM_050435, Figure 5193: PRO82905 Figure 5194A-B: DNA326538, XM_035636, gen.XM_050435 Figure 5228: PRO82923 gen.XM_035636 Figure 5229: DNA326559, XM_113988, Figure 5195: DNA326539, XM_012862, gen.XM_113988 gen.XM_012862 Figure 5230: DNA326560, NM_058164, Figure 5196A-B: DNA326540, XM_035627, gen.NM_058164 gen.XM_035627 Figure 5197A-B: DNA326541, XM_035625, Figure 5231: PRO82925 Figure 5232: DNA227280, NM_020230, gen.XM_035625 gen.NM_020230 Figure 5198: PRO82909 Figure 5233: PRO37743 Figure 5199: DNA274761, NM_014649, Figure 5234: DNA270621, NM_003755, gen.NM_014649 gen.NM_003755 Figure 5200: PRO62531 Figure 5235: PRO58991 Figure 5201: DNA272421, NM .006012, Figure 5236: DNA326561, XM_049502, gen.NM_006012 Figure 5202: PRO60674 gen.XM_049502 Figure 5237: DNA326562, NM_007065, Figure 5203: DNA326542, NM_003685, gen.NM_007065 gen.NM_003685 Figure 5238: PRO63226 Figure 5204: PRO82910 Figure 5239: DNA326563, XM_049561, Figure 5205A-B: DNA326543, XM_009010, gen.XM_049561 gen.XM_009010 Figure 5240: DNA326564, XM_017204, Figure 5206: DNA270315, NM_004240, gen.NM_004240 gen.XM_017204 Figure 5207: PRO58702 Figure 5241: DNA326565, NM_005498, Figure 5208: DNA326544, NM_005490, gen.NM_005498 Figure 5242: PRO62112 gen.NM_005490 Figure 5243: DNA326566, XM .008887, Figure 5209: PRO201 gen.XM_008887 Figure 5210: DNA326546, XM _044619, Figure 5244: DNA326567, XM_085862, gen.XM_044619 gen.XM_085862 Figure 5211: PRO82912 Figure 5245: PRO82930 Figure 5212: DNA326547, XM_012798,

Figure 5246: DNA326568, XM_084014, gen.XM_084014 Figure 5247A-B: DNA326569, XM_032710, gen.XM_032710 Figure 5248: DNA326570, XM_032719, gen.XM_032719 Figure 5249: PRO82933 Figure 5250: DNA326571, NM_024029, gen.NM_024029 Figure 5251: PRO23794 Figure 5252: DNA326572, XM_032724, gen.XM_032724 Figure 5253: PRO82934 Figure 5254A-B: DNA326573, NM .003072, gen.NM_003072 Figure 5255: PRO82935 Figure 5256A-B: DNA326574, XM _009082, gen.XM_009082 Figure 5257: DNA326575, XM_032774, gen.XM_032774 Figure 5258: DNA218271, NM_000121, gen.NM_000121 Figure 5259: PRO34323 Figure 5260: DNA326576, XM_057074, gen.XM_057074 Figure 5261: DNA326577, XM_032782, gen.XM_032782 Figure 5262: DNA326578, NM_032377, gen.NM_032377 Figure 5263: PRO82939 Figure 5264: DNA326579, XM_015697, gen.XM_015697 Figure 5265: PRO82940 Figure 5266: DNA326580, XM_010156, gen.XM_010156 Figure 5267: DNA326581, NM _001930, gen.NM_001930 Figure 5268: PRO58446 Figure 5269: DNA326582, NM 013406, gen.NM_013406 Figure 5270: DNA326583, NM_013407, gen.NM_013407 Figure 5271: PRO82943 Figure 5272: DNA103320, NM_002229, gen.NM_002229 Figure 5273: PRO4650 Figure 5274: DNA326584, XM_009063, gen.XM_009063 Figure 5275: PRO82944 Figure 5276: DNA326585, XM_085917, gen.XM_085917 Figure 5277: DNA274034, NM_006397, gen.NM_006397 Figure 5278: PRO61977 Figure 5279: DNA287243, NM_004461,

gen.NM_004461

Figure 5280: PRO69518 Figure 5281: DNA326586, XM_032020, gen.XM_032020 Figure 5282: PRO2718 Figure 5283: DNA326587, NM_005053, gen.NM_005053 Figure 5284: PRO22613 Figure 5285: DNA326588, XM_085916, gen.XM_085916 Figure 5286: DNA326589, NM_017722, gen.NM_017722 Figure 5287: PRO82947 Figure 5288: DNA326590, NM _003765, gen.NM_003765 Figure 5289: PRO82948 Figure 5290: DNA326591, XM_051364, gen.XM_051364 Figure 5291: PRO82949 Figure 5292: DNA326592, XM_031345, gen.XM_031345 Figure 5293: PRO82950 Figure 5294: DNA326593, XM_113352, gen.XM_113352 Figure 5295: DNA326594, XM_058967, gen.XM_058967 Figure 5296: PRO82952 Figure 5297: DNA326595, XM_085909, gen.XM_085909 Figure 5298: DNA269894, NM_002730, gen.NM_002730 Figure 5299: PRO58292 Figure 5300: DNA326596, NM_018154, gen.NM_018154 Figure 5301: PRO82954 Figure 5302: DNA326597, XM_031276, gen.XM_031276 Figure 5303: DNA326598, XM_031273, gen.XM_031273 Figure 5304: PRO82956 Figure 5305: DNA326599, XM 2031263, gen.XM_031263 Figure 5306: PRO82957 Figure 5307: DNA326600, XM_031251, gen.XM_031251 Figure 5308: DNA326601, NM_006844, gen.NM_006844 Figure 5309: PRO82958 Figure 5310A-C: DNA326602, XM _009303, gen.XM_009303 Figure 5311: DNA326603, XM_086074, gen.XM_086074 Figure 5312: DNA269630, NM _003290, gen.NM_003290 Figure 5313: PRO58042 Figure 5314: DNA326604, NM _005370, gen.NM_005370

gen.XM_114004

Figure 5350: DNA326625, NM_012181, Figure 5315: PRO12130 gen.NM_012181 Figure 5316: DNA326605, XM_113348, Figure 5351: PRO82980 gen.XM_113348 Figure 5352: DNA227249, NM_007263, Figure 5317: DNA326606, NM_032207, gen.NM_007263 gen.NM_032207 Figure 5353: PRO37712 Figure 5318: PRO82962 Figure 5354: DNA326626, XM_018515, Figure 5319A-B: DNA326607, NM .006387, gen.XM_018515 gen.NM_006387 Figure 5355: DNA326627, NM_033415, Figure 5320: PRO82963 gen.NM_033415 Figure 5321: DNA326608, NM_024881, Figure 5356: PRO82982 gen.NM_024881 Figure 5357: DNA326628, XM _009330, Figure 5322: PRO82964 gen.XM_009330 Figure 5323: DNA326609, NM_024104, Figure 5358: DNA326629, NM_134440, gen.NM_024104 gen.NM_134440 Figure 5324: PRO82965 Figure 5359: PRO82983 Figure 5325A-C: DNA326610, XM .008854, Figure 5360: DNA326630, NM _003721, gen.XM_008854 gen.NM_003721 Figure 5326: DNA326611, NM_014173, Figure 5361: PRO59220 gen.NM_014173 Figure 5362: DNA326631, NM_015965, Figure 5327: PRO82967 gen.NM_015965 Figure 5328: DNA287240, NM_004335, Figure 5363: PRO82984 gen.NM_004335 Figure 5364: DNA326632, XM_016378, Figure 5329: PRO29371 gen.XM_016378 Figure 5330: DNA326612, XM_050660, Figure 5365: PRO82985 gen.XM_050660 Figure 5366: DNA326633, XM_114027, Figure 5331: DNA326613, XM_086116, gen.XM_114027 gen.XM_086116 Figure 5367: DNA326634, XM_165963, Figure 5332: DNA326614, NM_018174, gen.XM_165963 gen.NM_018174 Figure 5368: PRO82987 Figure 5333: PRO82970 Figure 5369: DNA326635, XM_015769, Figure 5334: DNA326615, NM_000980, gen.XM_015769 gen.NM_000980 Figure 5370: DNA326636, XM_012812, Figure 5335: PRO82971 gen.XM_012812 Figure 5336: DNA326616, XM_055230, Figure 5371: DNA326637, XM _085971, gen.XM_055230 gen.XM_085971 Figure 5337: DNA326617, XM_012179, Figure 5372: DNA326638, XM_037662, gen.XM_012179 gen.XM_037662 Figure 5338A-B: DNA326618, XM_009293, Figure 5373: PRO82991 gen.XM_009293 Figure 5374: DNA326639, NM_001238, Figure 5339: DNA326619, XM_038146, gen.NM_001238 gen.XM_038146 Figure 5375: PRO82992 Figure 5340: PRO82975 Figure 5376: DNA326640, NM_057182, Figure 5341: DNA326620, XM_092046, gen.NM_057182 gen.XM_092046 Figure 5377: PRO4756 Figure 5342: PRO82976 Figure 5343: DNA326621, XM_038098, Figure 5378: DNA326641, XM _009180, gen.XM_009180 gen.XM_038098 Figure 5379: DNA326642, XM_117118, Figure 5344: PRO82977 Figure 5345: DNA326622, NM_032627, gen.XM_117118 Figure 5380: DNA326643, XM _092049, gen.NM_032627 gen.XM_092049 Figure 5346: PRO82978 Figure 5381: PRO82995 Figure 5347: DNA326623, XM_165960, Figure 5382: DNA326644, XM _028672, gen.XM_165960 gen.XM_028672 Figure 5348: PRO82979 Figure 5383: DNA326645, XM_028666, Figure 5349: DNA326624, XM_114004, gen.XM_028666

Figure 5384: DNA326646, XM_009338, gen.XM_009338 Figure 5385: DNA326647, XM_048258, gen.XM_048258 Figure 5386: PRO82998 Figure 5387: DNA256836, NM_018468, gen.NM_018468 Figure 5388: PRO51767 Figure 5389: DNA326648, NM_024321, gen.NM_024321 Figure 5390: PRO82999 Figure 5391A-B: DNA326649, XM_049237, gen.XM_049237 Figure 5392: PRO83000 Figure 5393: DNA326650, NM_032635, gen.NM_032635 Figure 5394: PRO23845 Figure 5395: DNA326651, XM_115615, gen.XM_115615 Figure 5396A-B: DNA326652, XM_091984, gen.XM_091984 Figure 5397: PRO83002 Figure 5398: DNA326653, XM_085986, gen.XM_085986 Figure 5399: DNA326654, XM_032285, gen.XM_032285 Figure 5400: PRO83004 Figure 5401: DNA326655, NM_002812, gen.NM_002812 Figure 5402: PRO83005 Figure 5403A-E: DNA326656, XM _029455, gen.XM_029455 Figure 5404: DNA326657, XM_029450, gen.XM_029450 Figure 5405: PRO83007 Figure 5406: DNA326658, XM_009149, gen.XM_009149 Figure 5407: PRO62500 Figure 5408: DNA326659, XM_056602, gen.XM_056602 Figure 5409: DNA326660, NM_012237, gen.NM_012237 Figure 5410: PRO83008 Figure 5411: DNA326661, NM_030593, gen.NM_030593 Figure 5412: PRO83009 Figure 5413: DNA326662, NM_017827, gen.NM_017827 Figure 5414: PRO83010 Figure 5415: DNA326663, NM_021107, gen.NM_021107 Figure 5416: PRO83011 Figure 5417: DNA326664, NM_033363, gen.NM_033363 Figure 5418: PRO83012

Figure 5419: DNA326665, XM_059045,

gen.XM_059045 Figure 5420: PRO83013 Figure 5421: DNA273474, NM_005884, gen.NM_005884 Figure 5422: PRO61458 Figure 5423: DNA326666, XM_046090, gen.XM_046090 Figure 5424: PRO83014 Figure 5425: DNA326667, XM _086004, gen.XM_086004 Figure 5426: DNA272347, NM_001020, gen.NM_001020 Figure 5427: PRO60603 Figure 5428A-B: DNA326668, NM_003169, gen.NM_003169 Figure 5429: PRO12822 Figure 5430: DNA326669, XM_053074, gen.XM_053074 Figure 5431: PRO83016 Figure 5432: DNA326670, NM_016941, gen.NM_016941 Figure 5433: PRO83017 Figure 5434: DNA256840, NM_004714, gen.NM_004714 Figure 5435: PRO51771 Figure 5436: DNA326671, NM_001436, gen.NM_001436 Figure 5437: PRO83018 Figure 5438: DNA326672, XM_016410, gen.XM_016410 Figure 5439: DNA326673, XM_012860, gen.XM_012860 Figure 5440: DNA326674, XM_097365, gen.XM_097365 Figure 5441: DNA274139, NM_006503, gen.NM_006503 Figure 5442: PRO62075 Figure 5443: DNA326675, XM_009203, gen.XM_009203 Figure 5444: DNA326676, XM_047409, gen.XM_047409 Figure 5445: DNA326677, XM_047376, gen.XM_047376 Figure 5446A-B: DNA326678, XM_047374, gen.XM_047374 Figure 5447: DNA326679, XM_059052, gen.XM_059052 Figure 5448: DNA273600, NM .004596, gen.NM_004596 Figure 5449: PRO61575 Figure 5450: DNA326680, XM_030914, gen.XM_030914 Figure 5451: DNA326681, NM_052848, gen.NM_052848 Figure 5452: PRO83027 Figure 5453: DNA326682, XM_008912,

gen.XM_008912	gen.XM_085950
Figure 5454: DNA326683, NM_020158,	Figure 5488: DNA326704, XM_028263,
gen.NM_020158	gen.XM_028263
Figure 5455: PRO83029	Figure 5489: DNA326705, XM_085928,
Figure 5456: DNA326684, XM_030901,	gen.XM_085928
gen.XM_030901	Figure 5490: PRO36963
Figure 5457: PRO83030	Figure 5491: DNA326706, XM_028267,
Figure 5458: DNA326685, NM_018035,	gen.XM_028267
gen.NM_018035	Figure 5492: DNA326707, NM_013403,
Figure 5459: PRO83031	gen.NM_013403
Figure 5460: DNA326686, XM_085874,	Figure 5493: PRO83050
gen.XM_085874	Figure 5494: DNA103580, NM_001743,
Figure 5461: DNA326687, XM_085875,	gen.NM_001743
gen.XM_085875	Figure 5495: PRO4904
Figure 5462: DNA326688, XM_085876,	Figure 5496: DNA326708, XM_009126,
gen.XM_085876	gen.XM_009126
Figure 5463: DNA326689, XM_058949,	Figure 5497: DNA326709, NM_006247,
gen.XM_058949	gen.NM_006247
Figure 5464: PRO83035	Figure 5498: PRO25881
Figure 5465: DNA326690, XM_030895,	Figure 5499: DNA326710, NM_003370,
gen.XM_030895	gen.NM_003370
Figure 5466: DNA326691, XM_115603,	Figure 5500: PRO83052
gen.XM_115603	Figure 5501: DNA326711, XM_085856,
Figure 5467: PRO83037	gen.XM_085856
Figure 5468: DNA326692, NM_001022,	Figure 5502: DNA150784, NM_001983,
gen.NM_001022	gen.NM_001983
Figure 5469: PRO83038	Figure 5503: PRO12800
Figure 5470: DNA326693, NM_004706,	Figure 5504: DNA270931, NM_012099,
gen.NM_004706	gen.NM_012099
Figure 5471: PRO83039	Figure 5505: PRO59264
Figure 5472: DNA326694, XM _008878,	Figure 5506A-B: DNA257531, NM_031417,
gen.XM_008878	gen.NM_031417
Figure 5473: PRO83040	Figure 5507: PRO52101
Figure 5474: DNA326695, NM_022752,	Figure 5508: DNA326712, NM_001294,
gen.NM_022752	gen.NM_001294
Figure 5475: PRO83041	Figure 5509: PRO83054
Figure 5476: DNA151808, NM_006494,	Figure 5510: DNA326713, XM _097274,
gen.NM_006494	gen.XM_097274
Figure 5477: PRO12892	Figure 5511: DNA88084, NM .000041,
Figure 5478: DNA326696, NM_001816,	gen.NM_000041
gen.NM_001816	Figure 5512: PRO2644
Figure 5479: PRO34151	Figure 5513: DNA256533, NM_006114,
Figure 5480: DNA326697, NM_000554,	gen.NM_006114
gen.NM_000554	Figure 5514: PRO51565
Figure 5481: PRO83042	Figure 5515: DNA251057, NM _002856,
Figure 5482: DNA326698, XM_049920,	gen.NM_002856
gen.XM_049920	Figure 5516: PRO47354
Figure 5483: DNA326699, XM_055859,	Figure 5517: DNA226011, NM_005581,
gen.XM_055859	gen.NM_005581
Figure 5484A-B: DNA326700, XM_009125,	Figure 5518: PRO36474
gen.XM_009125	Figure 5519: DNA326714, NM_012116,
Figure 5485: DNA326701, XM_008860,	gen.NM_012116
gen.XM_008860	Figure 5520: PRO83056
Figure 5486: DNA326702, XM_009036,	Figure 5521: DNA326715, XM .097275,
gen.XM_009036	gen.XM_097275
Figure 5487: DNA326703, XM _085950,	Figure 5522: DNA326716, XM_008851,
	HIGHITE 3322: DNA320/10. AMI_000031.

gen.XM_008851	gen.NM_003598
Figure 5523: DNA274289, NM_016440,	Figure 5557: PRO83075
gen.NM_016440	Figure 5558: DNA326736, NM_006666,
Figure 5524: PRO62212	gen.NM_006666
Figure 5525: DNA326717, NM_012068,	Figure 5559: PRO83076
gen.NM_012068	Figure 5560: DNA326737, XM_114024,
Figure 5526: PRO83059	gen.XM_114024
Figure 5527: DNA326718, XM_085927,	Figure 5561: PRO83077
gen.XM_085927	Figure 5562: DNA304658, NM_000146,
Figure 5528: DNA326719, XM_084023,	gen.NM_000146
gen.XM_084023	Figure 5563: PRO71085
Figure 5529: DNA326720, XM_167530,	Figure 5564: DNA326738, NM_004324,
gen.XM_167530	gen.NM_004324
Figure 5530: DNA326721, XM_114025,	Figure 5565: PRO38101
gen.XM_114025	Figure 5566: DNA326739, NM_006184,
Figure 5531: DNA326722, XM_008985,	gen.NM_006184
gen.XM_008985	Figure 5567: PRO83078
Figure 5532: DNA326723, NM_030973,	Figure 5568: DNA273066, NM_001190,
gen.NM_030973	gen.NM_001190
Figure 5533: PRO83065	Figure 5569: PRO61129
Figure 5534: DNA326724, NM_025129,	Figure 5570: DNA326740, XM_058987,
gen.NM_025129	gen.XM_058987
Figure 5535: PRO83066	Figure 5571: DNA326741, NM_000979,
Figure 5536: DNA326725, NM_014203,	gen.NM_000979
gen.NM_014203	Figure 5572: PRO83080
Figure 5537: DNA326726, XM_085934,	Figure 5573: DNA326742, XM_085935,
gen.XM_085934	gen.XM_085935
Figure 5538: PRO83068	Figure 5574: DNA326743, NM_031485,
Figure 5539: DNA326727, NM _001536,	gen.NM_031485
gen.NM_001536	Figure 5575: PRO61308
Figure 5540: PRO83069	Figure 5576: DNA103239, NM_006801,
Figure 5541: DNA326728, XM_165432,	gen.NM_006801
gen.XM_165432	Figure 5577: PRO4569
Figure 5542: DNA274823, NM _001571,	Figure 5578: DNA326744, XM_046419,
gen.NM_001571	gen.XM_046419
Figure 5543: PRO62582	Figure 5579: PRO83082
Figure 5544A-B: DNA326729, XM_046313,	Figure 5580: DNA326745, NM .002691,
gen.XM_046313	gen.NM_002691
Figure 5545: PRO83071	Figure 5581: PRO83083
Figure 5546: DNA326730, NM_015953,	Figure 5582: DNA326746, XM_056286,
gen.NM_015953	gen.XM_056286
Figure 5547: PRO83072	Figure 5583: PRO83084
Figure 5548: DNA326731, XM_027904,	Figure 5584: DNA326747, XM_058990,
gen.XM_027904	gen.XM_058990
Figure 5549: DNA326732, XM_084026,	Figure 5585: PRO83085
gen.XM_084026	Figure 5586: DNA326748, XM_091981,
Figure 5550: DNA290260, NM_012423,	gen.XM_091981
gen.NM_012423	Figure 5587: PRO83086
Figure 5551: PRO70385	Figure 5588: DNA326749, NM_032712,
Figure 5552: DNA326733, XM_058991,	gen.NM_032712
gen.XM_058991	Figure 5589: PRO23238
Figure 5553: PRO83073	Figure 5590: DNA83154, NM .001648,
Figure 5554: DNA326734, NM_017916,	gen.NM_001648
gen.NM_017916	Figure 5591: PRO2109
Figure 5555: PRO83074	Figure 5592: DNA326750, XM_055658,
Figure 5556: DNA326735, NM_003598,	gen.XM_055658

Figure 5593: DNA269481, NM_001985, gen.NM_001985 Figure 5594: PRO57901 Figure 5595: DNA326751, XM_091886, gen.XM_091886 Figure 5596: PRO83087 Figure 5597: DNA326752, XM_008830, gen.XM_008830 Figure 5598: DNA326753, XM_039908, gen.XM_039908 Figure 5599: PRO83089 Figure 5600: DNA326754, NM_015629, gen.NM_015629 Figure 5601: PRO83090 Figure 5602: DNA326755, XM_050236, gen.XM_050236 Figure 5603: DNA326756, XM_050589, gen.XM_050589 Figure 5604: PRO83092 Figure 5605: DNA326757, XM_117128, gen.XM_117128 Figure 5606: PRO83093 Figure 5607: DNA326758, XM_059321, gen.XM_059321 Figure 5608: DNA326759, NM_003283, gen.NM_003283 Figure 5609: PRO83095 Figure 5610A-B: DNA326760, NM_014931, gen.NM_014931 Figure 5611: PRO83096 Figure 5612: DNA326761, XM_035919, gen.XM_035919 Figure 5613: DNA326762, NM_000991, gen.NM_000991 Figure 5614: PRO83098 Figure 5615: DNA273346, NM_014501, gen.NM_014501 Figure 5616: PRO61349 Figure 5617: DNA326763, NM_013333, gen.NM_013333 Figure 5618: PRO83099 Figure 5619: DNA326764, NM .007279, gen.NM_007279 Figure 5620: PRO83100 Figure 5621: DNA326765, NM_016202, gen.NM_016202 Figure 5622: PRO83101 Figure 5623: DNA326766, XM _034377, gen.XM_034377 Figure 5624: PRO83102 Figure 5625: DNA272062, NM_014453, gen.NM_014453 Figure 5626: PRO60333

Figure 5627: DNA254548, NM _005762,

gen.NM_005762

Figure 5628: PRO49653

Figure 5629: DNA326767, XM_085972, gen.XM_085972 Figure 5630: PRO83103 Figure 5631: DNA326768, NM_032792, gen.NM_032792 Figure 5632: PRO83104 Figure 5633: DNA326769, NM .001009, gen.NM_001009 Figure 5634: PRO83105 Figure 5635: DNA326770, XM_058125, gen.XM_058125 Figure 5636: DNA326771, NM_024691, gen.NM_024691 Figure 5637: PRO83107 Figure 5638: DNA297288, NM_021158, gen.NM_021158 Figure 5639: PRO70810 Figure 5640: DNA304662, NM_031229, gen.NM_031229 Figure 5641: PRO71089 Figure 5642: DNA326772, NM_031228, gen.NM_031228 Figure 5643: PRO83108 Figure 5644: DNA326773, XM_097749, gen.XM_097749 Figure 5645: PRO83109 Figure 5646: DNA326774, XM_055993, gen.XM_055993 Figure 5647: DNA326775, XM_009622, gen.XM_009622 Figure 5648: DNA326776, NM_000801, gen.NM_000801 Figure 5649: PRO59142 Figure 5650: DNA326777, NM_054014, gen.NM_054014 Figure 5651: PRO59142 Figure 5652: DNA326778, NM_016143, gen.NM_016143 Figure 5653: PRO83112 Figure 5654: DNA287270, NM_003091, gen.NM_003091 Figure 5655: PRO69541 Figure 5656: DNA326779, NM_052881, gen.NM_052881 Figure 5657: PRO83113 Figure 5658: DNA326780, XM_044914, gen.XM_044914 Figure 5659: PRO83114 Figure 5660: DNA326781, XM_044915, gen.XM_044915 Figure 5661: DNA326782, NM _006899, gen.NM_006899 Figure 5662: PRO83116 Figure 5663: DNA326783, NM_019609, gen.NM_019609 Figure 5664: PRO83117

Figure 5665: DNA326784, NM_021826, gen.NM_021826 Figure 5666: PRO83118 Figure 5667: DNA326785, XM_045418, gen.XM_045418 Figure 5668: DNA287261, NM .017874, gen.NM_017874 Figure 5669: PRO69533 Figure 5670: DNA326786, XM_086710, gen.XM_086710 Figure 5671: DNA326787, XM_045451, gen.XM_045451 Figure 5672: PRO83121 Figure 5673: DNA326788, XM_114174, gen.XM_114174 Figure 5674: DNA326789, XM_045460, gen.XM_045460 Figure 5675: DNA326790, XM_059268, gen.XM_059268 Figure 5676A-B: DNA271010, NM_014737, gen.NM_014737 Figure 5677: PRO59339 Figure 5678: DNA326791, XM_056035, gen.XM_056035 Figure 5679: DNA83170, NM_001819, gen.NM_001819 Figure 5680: PRO2615 Figure 5681: DNA227348, NM_019095, gen.NM_019095 Figure 5682: PRO37811 Figure 5683: DNA326792, NM_003092, gen.NM_003092 Figure 5684: PRO83125 Figure 5685: DNA287290, NM_014426, gen.NM_014426 Figure 5686: PRO69560 Figure 5687: DNA326793, XM_086701, gen.XM_086701 Figure 5688: DNA326794, XM_117209, gen.XM_117209 Figure 5689A-B: DNA326795, XM_046520, gen.XM_046520 Figure 5690: PRO83128 Figure 5691: DNA326796, XM_115846, gen.XM_115846 Figure 5692: PRO83129 Figure 5693: DNA326797, NM_080820, gen.NM_080820 Figure 5694: PRO83130 Figure 5695: DNA326798, XM_086715, gen.XM.086715 Figure 5696: DNA326799, XM_092760, gen.XM_092760 Figure 5697: PRO83132

Figure 5698: DNA326800, NM_012255,

gen.NM_012255

Figure 5699: PRO83133 Figure 5700: DNA326801, XM_012970, gen.XM_012970 Figure 5701: DNA326802, XM_042765, gen.XM_042765 Figure 5702: PRO83135 Figure 5703: DNA150548, NM_001247, gen.NM_001247 Figure 5704: PRO12324 Figure 5705A-B: DNA326803, XM_009436, gen.XM_009436 Figure 5706: DNA326804, XM_114178, gen.XM_114178 Figure 5707: PRO83137 Figure 5708: DNA326805, XM_046160, gen.XM_046160 Figure 5709: PRO83138 Figure 5710: DNA326806, XM_046179, gen.XM_046179 Figure 5711: PRO83139 Figure 5712: DNA326807, XM_086745, gen.XM_086745 Figure 5713: DNA326808, NM_138578, gen.NM_138578 Figure 5714: PRO83141 Figure 5715: DNA326809, NM_012112, gen.NM_012112 Figure 5716: PRO83142 Figure 5717: DNA326810, XM_086736, gen.XM_086736 Figure 5718: PRO83143 Figure 5719: DNA326811, NM_030815, gen.NM_030815 Figure 5720: PRO83144 Figure 5721A-B: DNA150767, NM_014742, gen.NM_014742 Figure 5722: PRO12460 Figure 5723A-B: DNA326812, XM_047007, gen.XM_047007 Figure 5724: PRO83145 Figure 5725A-B: DNA326813, XM_047011, gen.XM_047011 Figure 5726: PRO83146 Figure 5727A-B: DNA326814, XM_047018, gen.XM_047018 Figure 5728: DNA326815, XM_009450, gen.XM_009450 Figure 5729: DNA326816, NM_033197, gen.NM_033197 Figure 5730: PRO83149 Figure 5731: DNA326817, XM_097772, gen.XM_097772 Figure 5732: PRO83150 Figure 5733: DNA326818, NM_016732, gen.NM_016732 Figure 5734: DNA97298, NM_003908,

gen.NM_024855 gen.NM_003908 Figure 5770: PRO83165 Figure 5735: PRO3645 Figure 5771A-B: DNA227472, NM_002660, Figure 5736: DNA326819, NM_000687, gen.NM_002660 gen.NM_000687 Figure 5772: PRO37935 Figure 5737: PRO83152 Figure 5738: DNA273517, NM_000178, Figure 5773: DNA326836, XM_097727, gen.XM_097727 gen.NM_000178 Figure 5774: DNA103525, NM_002466, Figure 5739: PRO61498 gen.NM_002466 Figure 5740: DNA326820, NM_018217, Figure 5775: PRO4852 gen.NM_018217 Figure 5776: DNA326837, XM_029810, Figure 5741: PRO83153 gen.XM_029810 Figure 5742: DNA326821, NM _002212, Figure 5777: PRO83167 gen.NM_002212 Figure 5743: PRO60945 Figure 5778: DNA326838, XM_029822, gen.XM_029822 Figure 5744A-C: DNA326822, NM -007186, Figure 5779: DNA326839, NM_002638, gen.NM_007186 gen.NM_002638 Figure 5745: DNA226758, NM _015966, Figure 5780: PRO2065 gen.NM_015966 Figure 5781: DNA326840, NM_003064, Figure 5746: PRO37221 gen.NM_003064 Figure 5747: DNA194701, NM _003915, Figure 5782: PRO1720 gen.NM_003915 Figure 5783: DNA326841, NM_015937, Figure 5748: PRO24002 gen.NM_015937 Figure 5749: DNA326823, XM_113380, Figure 5784: PRO83169 gen.XM_113380 Figure 5785: DNA273320, NM_007019, Figure 5750: DNA326824, NM_016558, gen.NM_007019 gen.NM_016558 Figure 5786: PRO61327 Figure 5751: PRO83155 Figure 5787: DNA326842, NM_033421, Figure 5752: DNA326825, NM_015511, gen.NM_033421 gen.NM_015511 Figure 5788: PRO83170 Figure 5753: PRO83156 Figure 5789: DNA88569, NM_006227, Figure 5754: DNA326826, XM_009501, gen.NM_006227 gen.XM_009501 Figure 5790: PRO2420 Figure 5755: PRO83157 Figure 5791: DNA88239, NM_004994, Figure 5756: DNA326827, XM_057236, gen.NM_004994 gen.XM_057236 Figure 5792: PRO2711 Figure 5757: DNA326828, NM_024918, Figure 5793: DNA326843, XM_057374, gen.NM_024918 gen.XM_057374 Figure 5758: PRO83159 Figure 5794: DNA326844, XM_114163, Figure 5759: DNA326829, XM_009642, gen.XM_114163 gen.XM_009642 Figure 5795A-B: DNA326845, XM_097731, Figure 5760: DNA194807, NM_006698, gen.XM_097731 gen.NM_006698 Figure 5796A-B: DNA326846, XM _030044, Figure 5761: PRO24077 Figure 5762: DNA326830, XM_009686, gen.XM_030044 Figure 5797: PRO83174 gen.XM_009686 Figure 5798: DNA326847, NM_017895, Figure 5763: DNA326831, NM_030877, gen.NM_017895 gen.NM_030877 Figure 5799: PRO83175 Figure 5764: PRO83161 Figure 5800: DNA326848, XM_097713, Figure 5765: DNA326832, XM_028806, gen.XM_097713 gen.XM_028806 Figure 5801: PRO83176 Figure 5766A-B: DNA326833, XM_028810, Figure 5802: DNA326849, NM_005985, gen.XM_028810 gen.NM_005985 Figure 5767: PRO83163 Figure 5768: DNA326834, XM_012931, Figure 5803: PRO83177 Figure 5804: DNA326850, NM_003349, gen.XM_012931 gen.NM_003349 Figure 5769: DNA326835, NM_024855,

gen.XM_037202

Figure 5841: PRO83190 Figure 5805: PRO83178 Figure 5806: DNA326851, NM_022442, Figure 5842: DNA326868, XM _037206, gen.XM_037206 gen.NM_022442 Figure 5843: PRO83191 Figure 5807: PRO83179 Figure 5844: DNA103486, NM_007002, Figure 5808: DNA326852, NM_005194, gen.NM_007002 gen.NM_005194 Figure 5845: PRO4813 Figure 5809: DNA326853, NM_002827, Figure 5846A-D: DNA326869, XM _037217, gen.NM_002827 gen.XM_037217 Figure 5810: PRO38066 Figure 5847: DNA326870, NM_001024, Figure 5811: DNA326854, NM_003859, gen.NM_001024 gen.NM_003859 Figure 5848: PRO83193 Figure 5812: PRO83180 Figure 5849: DNA326871, NM _018270, Figure 5813: DNA326855, XM_114165, gen.NM_018270 gen.XM_114165 Figure 5850: PRO83194 Figure 5814: DNA269526, NM_001324, Figure 5851: DNA326872, XM_028783, gen.NM_001324 gen.XM_028783 Figure 5815: PRO57942 Figure 5852: PRO83195 Figure 5816: DNA326856, XM_009549, gen.XM_009549 Figure 5853: DNA326873, NM_001853, gen.NM_001853 Figure 5817: PRO83182 Figure 5854: PRO83196 Figure 5818: DNA326857, XM_030621, Figure 5855: DNA326874, NM _080796, gen.XM_030621 gen.NM_080796 Figure 5819: DNA326858, XM _086648, Figure 5856: PRO83197 gen.XM_086648 Figure 5857: DNA326875, NM _022105, Figure 5820: PRO83183 gen.NM_022105 Figure 5821: DNA326859, XM_009672, Figure 5858: PRO83198 gen.XM_009672 Figure 5859: DNA326876, NM_080797, Figure 5822: PRO83184 Figure 5823A-B: DNA326860, XM_009671, gen.NM_080797 Figure 5860: PRO83199 gen.XM_009671 Figure 5861: DNA326877, NM_018209, Figure 5824: DNA326861, NM_004738, gen.NM_018209 gen.NM_004738 Figure 5862: PRO83200 Figure 5825: PRO983 Figure 5863A-C: DNA326878, XM_028834, Figure 5826: DNA326862, NM_016592, gen.XM_028834 gen.NM_016592 Figure 5864: PRO83201 Figure 5827: PRO83185 Figure 5865: DNA326879, NM_024299, Figure 5828: DNA326863, NM_080425, gen.NM_024299 gen.NM_080425 Figure 5866: PRO83202 Figure 5829: PRO83186 Figure 5867A-C: DNA326880, XM_028918, Figure 5830: DNA304670, NM .000516, gen.NM_000516 gen.XM_028918 Figure 5868: PRO83203 Figure 5831: PRO71097 Figure 5832: DNA326864, NM_080426, Figure 5869: DNA326881, NM_032527, gen.NM_032527 gen.NM_080426 Figure 5833: PRO83187 Figure 5870: PRO83204 Figure 5871A-B: DNA326882, XM_028966, Figure 5834: DNA326865, XM_030699, gen.XM_028966 gen.XM_030699 Figure 5872: PRO83205 Figure 5835: PRO83188 Figure 5873: DNA269746, NM_012469, Figure 5836: DNA188229, NM_000114, gen.NM_012469 gen.NM_000114 Figure 5874: PRO58155 Figure 5837: PRO21728 Figure 5875: DNA326883, XM_114154, Figure 5838: DNA326866, NM .002792, gen.XM_114154 gen.NM_002792 Figure 5876: DNA326884, XM_072173, Figure 5839: PRO83189 gen.XM_072173 Figure 5840A-B: DNA326867, XM_037202,

Figure 5877: DNA326885, XM_086759,

Figure 5912: PRO83219

Figure 5913: DNA326901, XM_036042, gen.XM_086759 Figure 5878: DNA326886, XM_086760, gen.XM_036042 Figure 5914: DNA326902, XM_086770, gen.XM_086760 gen.XM_086770 Figure 5879: DNA326887, NM_021219, Figure 5915: DNA326903, NM_004928, gen.NM_021219 gen.NM_004928 Figure 5880: PRO28687 Figure 5916: PRO83222 Figure 5881: DNA188732, NM_000484, Figure 5917: DNA326904, XM_036087, gen.NM_000484 gen.XM_036087 Figure 5882: PRO25302 Figure 5918: PRO83223 Figure 5883: DNA326888, NM_016940, Figure 5919: DNA326905, XM_009805, gen.NM_016940 gen.XM_009805 Figure 5884: PRO83210 Figure 5920: PRO83224 Figure 5885: DNA254572, NM_006585, Figure 5921: DNA226409, NM_004339, gen.NM_006585 gen.NM_004339 Figure 5886: PRO49675 Figure 5922: PRO36872 Figure 5887: DNA326889, NM_005806, Figure 5923: DNA326906, XM_036107, gen.NM_005806 gen.XM_036107 Figure 5888: PRO83211 Figure 5924A-B: DNA326907, XM_036175, Figure 5889: DNA326890, XM_114185, gen.XM_036175 gen.XM_114185 Figure 5925: DNA326908, XM _097817, Figure 5890: DNA254994, NM_017613, gen.XM_097817 gen.NM_017613 Figure 5926A-B: DNA326909, XM_054566, Figure 5891: PRO50083 gen.XM_054566 Figure 5892: DNA274129, NM_001697, Figure 5927: DNA326910, XM_036755, gen.NM_001697 gen.XM_036755 Figure 5893: PRO62065 Figure 5928: DNA326911, XM_086773, Figure 5894: DNA326891, NM_001757, gen.XM_086773 gen.NM_001757 Figure 5929: DNA326912, XM_097807, Figure 5895: PRO83212 gen.XM_097807 Figure 5896A-C: DNA151898, NM_003316, Figure 5930: DNA326913, XM_086777, gen.NM_003316 gen.XM_086777 Figure 5897: PRO12135 Figure 5931: DNA326914, NM_002340, Figure 5898: DNA326892, NM_003720, gen.NM_002340 gen.NM_003720 Figure 5932: PRO83233 Figure 5899: PRO83213 Figure 5933A-B: DNA326915, NM_003906, Figure 5900: DNA326893, NM_002606, gen.NM_003906 gen.NM_002606 Figure 5934: PRO83234 Figure 5901: PRO83214 Figure 5935: DNA226617, NM .006272, Figure 5902: DNA326894, XM_033015, gen.NM_006272 gen.XM_033015 Figure 5936: PRO37080 Figure 5903: DNA326895, XM_033016, Figure 5937: DNA326916, NM_033070, gen.XM_033016 gen.NM_033070 Figure 5904: PRO59669 Figure 5938: PRO83235 Figure 5905: DNA326896, NM .003681, Figure 5939: DNA255046, NM_017829, gen.NM_003681 gen.NM_017829 Figure 5906: PRO69486 Figure 5940: PRO50134 Figure 5907: DNA326897, XM_035999, Figure 5941: DNA326917, NM_001696, gen.XM_035999 gen.NM_001696 Figure 5908: DNA326898, NM_020132, Figure 5942: PRO83236 gen.NM_020132 Figure 5943A-B: DNA326918, XM_032996, Figure 5909: PRO83217 gen.XM_032996 Figure 5910: DNA326899, XM_036011, Figure 5944: PRO83237 gen.XM_036011 Figure 5945: DNA326919, XM_167538, Figure 5911: DNA326900, NM_013369, gen.XM_167538 gen.NM_013369 Figure 5946: DNA326920, XM_033090,

gen.XM_033090	Figure 5981A-B: DNA326938, XM_037797,
Figure 5947: DNA225954, NM_000407,	gen.XM_037797
gen.NM _000407	Figure 5982: PRO83256
Figure 5948: PRO36417	Figure 5983: DNA326939, NM_004175,
Figure 5949: DNA326921, XM_058918,	gen.NM_004175
gen.XM_058918	Figure 5984: PRO83257
Figure 5950: DNA326922, XM_097833,	Figure 5985: DNA326940, XM_086821,
gen.XM_097833	gen.XM_086821
Figure 5951: DNA326923, NM_024627,	Figure 5986: DNA326941, XM_092888,
gen.NM_024627	gen.XM_092888
Figure 5952: PRO83242	Figure 5987: DNA326942, NM_005080,
Figure 5953: DNA326924, XM_086809,	gen.NM_005080
gen.XM_086809	Figure 5988: PRO83260
Figure 5954: DNA326925, NM_006440,	Figure 5989: DNA269830, NM_005243,
gen.NM_006440	gen.NM_005243
Figure 5955: PRO83244	Figure 5990: PRO58232
Figure 5956: DNA226561, NM .000754,	Figure 5991: DNA326943, NM_006478,
gen.NM_000754	gen.NM_006478
Figure 5957: PRO37024	Figure 5992: PRO83261
Figure 5958: DNA326926, NM_007310,	Figure 5993A-B: DNA326944, XM _037945,
gen.NM_007310	gen.XM_037945
Figure 5959: PRO83245	Figure 5994: DNA103462, NM_000268,
Figure 5960A-B: DNA326927, XM .033813,	gen.NM_000268
gen.XM_033813	Figure 5995: PRO4789
Figure 5961: DNA326928, NM_022727,	Figure 5996: DNA326945, NM_032204,
gen.NM_022727	gen.NM_032204
Figure 5962: PRO83247	Figure 5997: PRO83263
Figure 5963: DNA326929, XM_086805,	Figure 5998: DNA326946, XM_066291,
gen.XM_086805	gen.XM_066291
Figure 5964: DNA326930, XM_086873,	Figure 5999: DNA326947, NM_005877,
gen.XM_086873	gen.NM_005877
Figure 5965: DNA257549, NM _030573,	Figure 6000: PRO62328
gen.NM_030573	Figure 6001: DNA326948, NM_016498,
Figure 5966: PRO52119	gen.NM_016498
Figure 5967: DNA326931, XM .096155,	Figure 6002: PRO83265
gen.XM_096155	Figure 6003: DNA254141, NM_014303,
Figure 5968: DNA326932, XM_096156,	gen.NM_014303
gen.XM.096156	Figure 6004: PRO49256
Figure 5969A-B: DNA326933, XM_036937,	Figure 6005A-B: DNA151882, NM_014941,
gen.XM_036937	gen.NM_014941
Figure 5970: PRO83252	Figure 6006: PRO12134 Figure 6007: DNA326949, NM_006932,
Figure 5971: DNA326934, XM_097886,	gen.NM_006932
gen.XM_097886 Figure 5972: PRO83253	Figure 6008: PRO83266
	Figure 6009: DNA326950, NM_134269,
Figure 5973: DNA304835, NM_022044,	gen.NM_134269
gen.NM_022044 Figure 5974: PRO71242	Figure 6010: PRO83267
Figure 5975: DNA326935, NM_006115,	Figure 6011: DNA270697, NM_004147,
gen.NM_006115	gen.NM_004147
Figure 5976: PRO37012	Figure 6012: PRO59061
Figure 5970: PNO37012 Figure 5977: DNA326936, XM _037682,	Figure 6013: DNA326951, XM_059335,
gen.XM_037682	gen.XM_059335
Figure 5978: PRO83254	Figure 6014: DNA326952, XM_018539,
Figure 5979: DNA326937, NM_002415,	gen.XM_018539
gen.NM_002415	Figure 6015: DNA326953, NM_014306,
Figure 5980: PRO83255	gen.NM_014306
1 18u10 2700, 1 NO03233	Portrational .

Figure 6016: PRO83270 Figure 6017: DNA326954, NM_012179, gen.NM_012179 Figure 6018: PRO83271 Figure 6019A-B: DNA326955, XM 038584, gen.XM_038584 Figure 6020: DNA151752, NM_002133, gen.NM_002133 Figure 6021: PRO12886 Figure 6022: DNA326956, XM_009947, gen.XM_009947 Figure 6023: PRO12845 Figure 6024: DNA326957, XM_114209, gen.XM_114209 Figure 6025A-B: DNA326958, NM_002473, gen.NM_002473 Figure 6026: PRO83273 Figure 6027: DNA188740, NM_003753, gen.NM_003753 Figure 6028: PRO22481 Figure 6029: DNA326959, NM_021126, gen.NM_021126 Figure 6030: PRO70331 Figure 6031: DNA326960, XM_009967, gen.XM_009967 Figure 6032: DNA326961, NM_013365, gen.NM_013365 Figure 6033: PRO83274 Figure 6034: DNA290259, NM_018957, gen.NM_018957 Figure 6035: PRO70383 Figure 6036: DNA326962, NM_020315, gen.NM_020315 Figure 6037: PRO83275 Figure 6038: DNA304719, NM_002305, gen.NM_002305 Figure 6039: PRO71145 Figure 6040: DNA326963, NM _007032, gen.NM_007032 Figure 6041: PRO83276 Figure 6042: DNA326964, XM_009973, gen.XM_009973 Figure 6043: DNA326965, XM_086830, gen.XM_086830 Figure 6044: PRO83278 Figure 6045: DNA254240, NM_016091, gen.NM_016091 Figure 6046: PRO49352 Figure 6047A-B: DNA326966, XM_039236, gen.XM_039236 Figure 6048: PRO83279 Figure 6049: DNA326967, NM_006941, gen.NM_006941

Figure 6050: PRO83280

gen.XM_039248

Figure 6051: DNA326968, XM_039248,

Figure 6052: DNA326969, NM_012323, gen.NM_012323 Figure 6053: PRO83282 Figure 6054: DNA326970, NM_012264, gen.NM_012264 Figure 6055: PRO12490 Figure 6056: DNA326971, NM_015373, gen.NM_015373 Figure 6057: PRO83283 Figure 6058: DNA326972, NM_020243, gen.NM_020243 Figure 6059: PRO23231 Figure 6060: DNA326973, XM_039339, gen.XM_039339 Figure 6061: DNA326974, NM_000967, gen.NM_000967 Figure 6062: PRO83285 Figure 6063: DNA326975, XM _010000, gen.XM_010000 Figure 6064: DNA326976, XM_010002, gen.XM_010002 Figure 6065: DNA326977, XM_039372, gen.XM_039372 Figure 6066: DNA326978, XM_013010, gen.XM_013010 Figure 6067: PRO83288 Figure 6068: DNA254165, NM_000026, gen.NM_000026 Figure 6069: PRO49278 Figure 6070: DNA326979, NM_003932, gen.NM_003932 Figure 6071: PRO4586 Figure 6072: DNA326980, NM_014248, gen.NM_014248 Figure 6073: PRO83289 Figure 6074: DNA326981, XM_086844, gen.XM_086844 Figure 6075: DNA219225, NM_002883, gen.NM_002883 Figure 6076: PRO34531 Figure 6077: DNA326982, NM_003216, gen.NM_003216 Figure 6078: PRO83291 Figure 6079: DNA270954, NM _001098, gen.NM_001098 Figure 6080: PRO59285 Figure 6081: DNA326983, NM_001469, gen.NM_001469 Figure 6082: PRO4872 Figure 6083: DNA326984, NM_005008, gen.NM_005008 Figure 6084: PRO83292 Figure 6085A-B: DNA326985, NM_004599, gen.NM_004599 Figure 6086: PRO83293

Figure 6087A-B: DNA326986, XM_010024,

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DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

I. <u>Definitions</u>

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The terms "TAT polypeptide" and "TAT" as used herein and when immediately followed by a numerical designation, refer to various polypeptides, wherein the complete designation (i.e., TAT/number) refers to specific polypeptide sequences as described herein. The terms "TAT/number polypeptide" and "TAT/number" wherein the term "number" is provided as an actual numerical designation as used herein encompass native sequence polypeptides, polypeptide variants and fragments of native sequence polypeptides and polypeptide variants (which are further defined herein). The TAT polypeptides described herein may be isolated from a variety of sources, such as from human tissue types or from another source, or prepared by recombinant or synthetic methods. The term "TAT polypeptide" refers to each individual TAT/number polypeptide disclosed herein. All disclosures in this specification which refer to the "TAT polypeptide" refer to each of the polypeptides individually as well as jointly. For example, descriptions of the preparation of, purification of, derivation of, formation of antibodies to or against, formation of TAT binding oligopeptides to or against, formation of TAT binding organic molecules to or against, administration of, compositions containing, treatment of a disease with, etc., pertain to each polypeptide of the invention individually. The term "TAT polypeptide" also includes variants of the TAT/number polypeptides disclosed herein.

A "native sequence TAT polypeptide" comprises a polypeptide having the same amino acid sequence as the corresponding TAT polypeptide derived from nature. Such native sequence TAT polypeptides can be isolated from nature or can be produced by recombinant or synthetic means. The term "native sequence TAT polypeptide" specifically encompasses naturally-occurring truncated or secreted forms of the specific TAT polypeptide (e.g., an extracellular domain sequence), naturally-occurring variant forms (e.g., alternatively spliced forms) and naturally-occurring allelic variants of the polypeptide. In certain embodiments of the invention, the native sequence TAT polypeptides disclosed herein are mature or full-length native sequence polypeptides comprising the full-length amino acids sequences shown in the accompanying figures. Start and stop codons (if indicated) are shown in bold font and underlined in the figures. Nucleic acid residues indicated as "N" in the accompanying figures are any nucleic acid residue. However, while the TAT polypeptides disclosed in the accompanying figures are shown to begin with methionine residues designated herein as amino acid position 1 in the figures, it is conceivable and possible that other methionine residues located either upstream or downstream from the amino acid position 1 in the figures may be employed as the starting amino acid residue for the TAT polypeptides.

The TAT polypeptide "extracellular domain" or "ECD" refers to a form of the TAT polypeptide which is essentially free of the transmembrane and cytoplasmic domains. Ordinarily, a TAT polypeptide ECD will have less than 1% of such transmembrane and/or cytoplasmic domains and preferably, will have less than 0.5% of such domains. It will be understood that any transmembrane domains identified for the TAT polypeptides of the present invention are identified pursuant to criteria routinely employed in the art for identifying that type of hydrophobic domain. The exact boundaries of a transmembrane domain may vary but most likely by no more than about 5 amino acids at either end of the domain as initially identified herein. Optionally, therefore, an

extracellular domain of a TAT polypeptide may contain from about 5 or fewer amino acids on either side of the transmembrane domain/extracellular domain boundary as identified in the Examples or specification and such polypeptides, with or without the associated signal peptide, and nucleic acid encoding them, are contemplated by the present invention.

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The approximate location of the "signal peptides" of the various TAT polypeptides disclosed herein may be shown in the present specification and/or the accompanying figures. It is noted, however, that the C-terminal boundary of a signal peptide may vary, but most likely by no more than about 5 amino acids on either side of the signal peptide C-terminal boundary as initially identified herein, wherein the C-terminal boundary of the signal peptide may be identified pursuant to criteria routinely employed in the art for identifying that type of amino acid sequence element (e.g., Nielsen et al., Prot. Eng. 10:1-6 (1997) and von Heinje et al., Nucl. Acids. Res. 14:4683-4690 (1986)). Moreover, it is also recognized that, in some cases, cleavage of a signal sequence from a secreted polypeptide is not entirely uniform, resulting in more than one secreted species. These mature polypeptides, where the signal peptide is cleaved within no more than about 5 amino acids on either side of the C-terminal boundary of the signal peptide as identified herein, and the polynucleotides encoding them, are contemplated by the present invention.

"TAT polypeptide variant" means a TAT polypeptide, preferably an active TAT polypeptide, as defined herein having at least about 80% amino acid sequence identity with a full-length native sequence TAT polypeptide sequence as disclosed herein, a TAT polypeptide sequence lacking the signal peptide as disclosed herein, an extracellular domain of a TAT polypeptide, with or without the signal peptide, as disclosed herein or any other fragment of a full-length TAT polypeptide sequence as disclosed herein (such as those encoded by a nucleic acid that represents only a portion of the complete coding sequence for a full-length TAT polypeptide). Such TAT polypeptide variants include, for instance, TAT polypeptides wherein one or more amino acid residues are added, or deleted, at the N- or C-terminus of the full-length native amino acid sequence. Ordinarily, a TAT polypeptide variant will have at least about 80% amino acid sequence identity, alternatively at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% amino acid sequence identity, to a full-length native sequence TAT polypeptide sequence as disclosed herein, a TAT polypeptide sequence lacking the signal peptide as disclosed herein, an extracellular domain of a TAT polypeptide, with or without the signal peptide, as disclosed herein or any other specifically defined fragment of a full-length TAT polypeptide sequence as disclosed herein. Ordinarily, TAT variant polypeptides are at least about 10 amino acids in length, alternatively at least about 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600 amino acids in length, or more. Optionally, TAT variant polypeptides will have no more than one conservative amino acid substitution as compared to the native TAT polypeptide sequence, alternatively no more than 2, 3, 4, 5, 6, 7, 8, 9, or 10 conservative amino acid substitution as compared to the native TAT polypeptide sequence.

"Percent (%) amino acid sequence identity" with respect to the TAT polypeptide sequences identified

herein is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in the specific TAT polypeptide sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared. For purposes herein, however, % amino acid sequence identity values are generated using the sequence comparison computer program ALIGN-2, wherein the complete source code for the ALIGN-2 program is provided in Table 1 below. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc. and the source code shown in Table 1 below has been filed with user documentation in the U.S. Copyright Office, Washington D.C., 20559, where it is registered under U.S. Copyright Registration No. TXU510087. The ALIGN-2 program is publicly available through Genentech, Inc., South San Francisco, California or may be compiled from the source code provided in Table 1 below. The ALIGN-2 program should be compiled for use on a UNIX operating system, preferably digital UNIX V4.0D. All sequence comparison parameters are set by the ALIGN-2 program and do not vary.

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In situations where ALIGN-2 is employed for amino acid sequence comparisons, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) is calculated as follows:

100 times the fraction X/Y

where X is the number of amino acid residues scored as identical matches by the sequence alignment program ALIGN-2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A. As examples of % amino acid sequence identity calculations using this method, Tables 2 and 3 demonstrate how to calculate the % amino acid sequence identity of the amino acid sequence designated "Comparison Protein" to the amino acid sequence designated "TAT", wherein "TAT" represents the amino acid sequence of a hypothetical TAT polypeptide of interest, "Comparison Protein" represents the amino acid sequence of a polypeptide against which the "TAT" polypeptide of interest is being compared, and "X, "Y" and "Z" each represent different hypothetical amino acid residues. Unless specifically stated otherwise, all % amino acid sequence identity values used herein are obtained as described in the immediately preceding paragraph using the ALIGN-2 computer program.

"TAT variant polynucleotide" or "TAT variant nucleic acid sequence" means a nucleic acid molecule

which encodes a TAT polypeptide, preferably an active TAT polypeptide, as defined herein and which has at least about 80% nucleic acid sequence identity with a nucleotide acid sequence encoding a full-length native sequence TAT polypeptide sequence as disclosed herein, a full-length native sequence TAT polypeptide sequence lacking the signal peptide as disclosed herein, an extracellular domain of a TAT polypeptide, with or without the signal peptide, as disclosed herein or any other fragment of a full-length TAT polypeptide sequence as disclosed herein (such as those encoded by a nucleic acid that represents only a portion of the complete coding sequence for a full-length TAT polypeptide). Ordinarily, a TAT variant polynucleotide will have at least about 80% nucleic acid sequence identity, alternatively at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% nucleic acid sequence identity with a nucleic acid sequence encoding a full-length native sequence TAT polypeptide sequence as disclosed herein, an extracellular domain of a TAT polypeptide, with or without the signal peptide as disclosed herein or any other fragment of a full-length TAT polypeptide, with or without the signal sequence, as disclosed herein or any other fragment of a full-length TAT polypeptide sequence as disclosed herein. Variants do not encompass the native nucleotide sequence.

Ordinarily, TAT variant polynucleotides are at least about 5 nucleotides in length, alternatively at least about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600, 610, 620, 630, 640, 650, 660, 670, 680, 690, 700, 710, 720, 730, 740, 750, 760, 770, 780, 790, 800, 810, 820, 830, 840, 850, 860, 870, 880, 890, 900, 910, 920, 930, 940, 950, 960, 970, 980, 990, or 1000 nucleotides in length, wherein in this context the term "about" means the referenced nucleotide sequence length plus or minus 10% of that referenced length.

"Percent (%) nucleic acid sequence identity" with respect to TAT-encoding nucleic acid sequences identified herein is defined as the percentage of nucleotides in a candidate sequence that are identical with the nucleotides in the TAT nucleic acid sequence of interest, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity. Alignment for purposes of determining percent nucleic acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN or Megalign (DNASTAR) software. For purposes herein, however, % nucleic acid sequence identity values are generated using the sequence comparison computer program ALIGN-2, wherein the complete source code for the ALIGN-2 program is provided in Table 1 below. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc. and the source code shown in Table 1 below has been filed with user documentation in the U.S. Copyright Office, Washington D.C., 20559, where it is registered under U.S. Copyright Registration No. TXU510087. The ALIGN-2 program is publicly available through Genentech, Inc., South San Francisco, California or may be compiled from the source code provided in Table 1 below. The ALIGN-2 program should be compiled for use on a UNIX operating system, preferably digital UNIX V4.0D. All sequence comparison

parameters are set by the ALIGN-2 program and do not vary.

In situations where ALIGN-2 is employed for nucleic acid sequence comparisons, the % nucleic acid sequence identity of a given nucleic acid sequence C to, with, or against a given nucleic acid sequence D (which can alternatively be phrased as a given nucleic acid sequence C that has or comprises a certain % nucleic acid sequence identity to, with, or against a given nucleic acid sequence D) is calculated as follows:

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100 times the fraction W/Z

where W is the number of nucleotides scored as identical matches by the sequence alignment program ALIGN-2 in that program's alignment of C and D, and where Z is the total number of nucleotides in D. It will be appreciated that where the length of nucleic acid sequence C is not equal to the length of nucleic acid sequence D, the % nucleic acid sequence identity of C to D will not equal the % nucleic acid sequence identity of D to C. As examples of % nucleic acid sequence identity calculations, Tables 4 and 5, demonstrate how to calculate the % nucleic acid sequence identity of the nucleic acid sequence designated "Comparison DNA" to the nucleic acid sequence designated "TAT-DNA", wherein "TAT-DNA" represents a hypothetical TAT-encoding nucleic acid sequence of interest, "Comparison DNA" represents the nucleotide sequence of a nucleic acid molecule against which the "TAT-DNA" nucleic acid molecule of interest is being compared, and "N", "L" and "V" each represent different hypothetical nucleotides. Unless specifically stated otherwise, all % nucleic acid sequence identity values used herein are obtained as described in the immediately preceding paragraph using the ALIGN-2 computer program.

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In other embodiments, TAT variant polynucleotides are nucleic acid molecules that encode a TAT polypeptide and which are capable of hybridizing, preferably under stringent hybridization and wash conditions, to nucleotide sequences encoding a full-length TAT polypeptide as disclosed herein. TAT variant polypeptides may be those that are encoded by a TAT variant polynucleotide.

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The term "full-length coding region" when used in reference to a nucleic acid encoding a TAT polypeptide refers to the sequence of nucleotides which encode the full-length TAT polypeptide of the invention (which is often shown between start and stop codons, inclusive thereof, in the accompanying figures). The term "full-length coding region" when used in reference to an ATCC deposited nucleic acid refers to the TAT polypeptide-encoding portion of the cDNA that is inserted into the vector deposited with the ATCC (which is often shown between start and stop codons, inclusive thereof, in the accompanying figures).

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"Isolated," when used to describe the various TAT polypeptides disclosed herein, means polypeptide that has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials that would typically interfere with diagnostic or therapeutic uses for the polypeptide, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous solutes. In preferred embodiments, the polypeptide will be purified (1) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (2) to homogeneity by SDS-PAGE under non-reducing or reducing conditions using Coomassie blue or,

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preferably, silver stain. Isolated polypeptide includes polypeptide in situ within recombinant cells, since at least one component of the TAT polypeptide natural environment will not be present. Ordinarily, however, isolated polypeptide will be prepared by at least one purification step.

An "isolated" TAT polypeptide-encoding nucleic acid or other polypeptide-encoding nucleic acid is a nucleic acid molecule that is identified and separated from at least one contaminant nucleic acid molecule with which it is ordinarily associated in the natural source of the polypeptide-encoding nucleic acid. An isolated polypeptide-encoding nucleic acid molecule is other than in the form or setting in which it is found in nature. Isolated polypeptide-encoding nucleic acid molecules therefore are distinguished from the specific polypeptide-encoding nucleic acid molecule as it exists in natural cells. However, an isolated polypeptide-encoding nucleic acid molecules contained in cells that ordinarily express the polypeptide where, for example, the nucleic acid molecule is in a chromosomal location different from that of natural cells.

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The term "control sequences" refers to DNA sequences necessary for the expression of an operably linked coding sequence in a particular host organism. The control sequences that are suitable for prokaryotes, for example, include a promoter, optionally an operator sequence, and a ribosome binding site. Eukaryotic cells are known to utilize promoters, polyadenylation signals, and enhancers.

Nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading phase. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while shorter probes need lower temperatures. Hybridization generally depends on the ability of denatured DNA to reanneal when complementary strands are present in an environment below their melting temperature. The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature which can be used. As a result, it follows that higher relative temperatures would tend to make the reaction conditions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel et al., Current Protocols in Molecular Biology, Wiley Interscience Publishers, (1995).

"Stringent conditions" or "high stringency conditions", as defined herein, may be identified by those that: (1) employ low ionic strength and high temperature for washing, for example 0.015 M sodium

chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42°C; or (3) overnight hybridization in a solution that employs 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with a 10 minute wash at 42°C in 0.2 x SSC (sodium chloride/sodium citrate) followed by a 10 minute high-stringency wash consisting of 0.1 x SSC containing EDTA at 55°C.

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"Moderately stringent conditions" may be identified as described by Sambrook et al., Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Press, 1989, and include the use of washing solution and hybridization conditions (e.g., temperature, ionic strength and %SDS) less stringent that those described above. An example of moderately stringent conditions is overnight incubation at 37°C in a solution comprising: 20% formamide, 5 x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5 x Denhardt's solution, 10% dextran sulfate, and 20 mg/ml denatured sheared salmon sperm DNA, followed by washing the filters in 1 x SSC at about 37-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc. as necessary to accommodate factors such as probe length and the like.

The term "epitope tagged" when used herein refers to a chimeric polypeptide comprising a TAT polypeptide or anti-TAT antibody fused to a "tag polypeptide". The tag polypeptide has enough residues to provide an epitope against which an antibody can be made, yet is short enough such that it does not interfere with activity of the polypeptide to which it is fused. The tag polypeptide preferably also is fairly unique so that the antibody does not substantially cross-react with other epitopes. Suitable tag polypeptides generally have at least six amino acid residues and usually between about 8 and 50 amino acid residues (preferably, between about 10 and 20 amino acid residues).

"Active" or "activity" for the purposes herein refers to form(s) of a TAT polypeptide which retain a biological and/or an immunological activity of native or naturally-occurring TAT, wherein "biological" activity refers to a biological function (either inhibitory or stimulatory) caused by a native or naturally-occurring TAT other than the ability to induce the production of an antibody against an antigenic epitope possessed by a native or naturally-occurring TAT and an "immunological" activity refers to the ability to induce the production of an antibody against an antigenic epitope possessed by a native or naturally-occurring TAT.

The term "antagonist" is used in the broadest sense, and includes any molecule that partially or fully blocks, inhibits, or neutralizes a biological activity of a native TAT polypeptide disclosed herein. In a similar manner, the term "agonist" is used in the broadest sense and includes any molecule that mimics a biological activity of a native TAT polypeptide disclosed herein. Suitable agonist or antagonist molecules specifically include agonist or antagonist antibodies or antibody fragments, fragments or amino acid sequence variants of native TAT polypeptides, peptides, antisense oligonucleotides, small organic molecules, etc. Methods for identifying agonists or antagonists of a TAT polypeptide may comprise contacting a TAT polypeptide with a candidate agonist or antagonist molecule and measuring a detectable change in one or more biological activities

normally associated with the TAT polypeptide.

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"Treating" or "treatment" or "alleviation" refers to both therapeutic treatment and prophylactic or preventative measures, wherein the object is to prevent or slow down (lessen) the targeted pathologic condition or disorder. Those in need of treatment include those already with the disorder as well as those prone to have the disorder or those in whom the disorder is to be prevented. A subject or mammal is successfully "treated" for a TAT polypeptide-expressing cancer if, after receiving a therapeutic amount of an anti-TAT antibody, TAT binding oligopeptide or TAT binding organic molecule according to the methods of the present invention, the patient shows observable and/or measurable reduction in or absence of one or more of the following: reduction in the number of cancer cells or absence of the cancer cells; reduction in the tumor size; inhibition (i.e., slow to some extent and preferably stop) of cancer cell infiltration into peripheral organs including the spread of cancer into soft tissue and bone; inhibition (i.e., slow to some extent and preferably stop) of tumor metastasis; inhibition, to some extent, of tumor growth; and/or relief to some extent, one or more of the symptoms associated with the specific cancer; reduced morbidity and mortality, and improvement in quality of life issues. To the extent the anti-TAT antibody or TAT binding oligopeptide may prevent growth and/or kill existing cancer cells, it may be cytostatic and/or cytotoxic. Reduction of these signs or symptoms may also be felt by the patient.

The above parameters for assessing successful treatment and improvement in the disease are readily measurable by routine procedures familiar to a physician. For cancer therapy, efficacy can be measured, for example, by assessing the time to disease progression (TTP) and/or determining the response rate (RR). Metastasis can be determined by staging tests and by bone scan and tests for calcium level and other enzymes to determine spread to the bone. CT scans can also be done to look for spread to the pelvis and lymph nodes in the area. Chest X-rays and measurement of liver enzyme levels by known methods are used to look for metastasis to the lungs and liver, respectively. Other routine methods for monitoring the disease include transrectal ultrasonography (TRUS) and transrectal needle biopsy (TRNB).

For bladder cancer, which is a more localized cancer, methods to determine progress of disease include urinary cytologic evaluation by cystoscopy, monitoring for presence of blood in the urine, visualization of the urothelial tract by sonography or an intravenous pyelogram, computed tomography (CT) and magnetic resonance imaging (MRI). The presence of distant metastases can be assessed by CT of the abdomen, chest x-rays, or radionuclide imaging of the skeleton.

"Chronic" administration refers to administration of the agent(s) in a continuous mode as opposed to an acute mode, so as to maintain the initial therapeutic effect (activity) for an extended period of time. "Intermittent" administration is treatment that is not consecutively done without interruption, but rather is cyclic in nature.

"Mammal" for purposes of the treatment of, alleviating the symptoms of or diagnosis of a cancer refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, sports, or pet animals, such as dogs, cats, cattle, horses, sheep, pigs, goats, rabbits, etc. Preferably, the mammal is human.

Administration "in combination with" one or more further therapeutic agents includes simultaneous

(concurrent) and consecutive administration in any order.

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"Carriers" as used herein include pharmaceutically acceptable carriers, excipients, or stabilizers which are nontoxic to the cell or mammal being exposed thereto at the dosages and concentrations employed. Often the physiologically acceptable carrier is an aqueous pH buffered solution. Examples of physiologically acceptable carriers include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptide; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEEN®, polyethylene glycol (PEG), and PLURONICS®.

By "solid phase" or "solid support" is meant a non-aqueous matrix to which an antibody, TAT binding oligopeptide or TAT binding organic molecule of the present invention can adhere or attach. Examples of solid phases encompassed herein include those formed partially or entirely of glass (e.g., controlled pore glass), polysaccharides (e.g., agarose), polyacrylamides, polystyrene, polyvinyl alcohol and silicones. In certain embodiments, depending on the context, the solid phase can comprise the well of an assay plate; in others it is a purification column (e.g., an affinity chromatography column). This term also includes a discontinuous solid phase of discrete particles, such as those described in U.S. Patent No. 4,275,149.

A "liposome" is a small vesicle composed of various types of lipids, phospholipids and/or surfactant which is useful for delivery of a drug (such as a TAT polypeptide, an antibody thereto or a TAT binding oligopeptide) to a mammal. The components of the liposome are commonly arranged in a bilayer formation, similar to the lipid arrangement of biological membranes.

A "small" molecule or "small" organic molecule is defined herein to have a molecular weight below about 500 Daltons.

An "effective amount" of a polypeptide, antibody, TAT binding oligopeptide, TAT binding organic molecule or an agonist or antagonist thereof as disclosed herein is an amount sufficient to carry out a specifically stated purpose. An "effective amount" may be determined empirically and in a routine manner, in relation to the stated purpose.

The term "therapeutically effective amount" refers to an amount of an antibody, polypeptide, TAT binding oligopeptide, TAT binding organic molecule or other drug effective to "treat" a disease or disorder in a subject or mammal. In the case of cancer, the therapeutically effective amount of the drug may reduce the number of cancer cells; reduce the tumor size; inhibit (i.e., slow to some extent and preferably stop) cancer cell infiltration into peripheral organs; inhibit (i.e., slow to some extent and preferably stop) tumor metastasis; inhibit, to some extent, tumor growth; and/or relieve to some extent one or more of the symptoms associated with the cancer. See the definition herein of "treating". To the extent the drug may prevent growth and/or kill existing cancer cells, it may be cytostatic and/or cytotoxic.

A "growth inhibitory amount" of an anti-TAT antibody, TAT polypeptide, TAT binding oligopeptide

or TAT binding organic molecule is an amount capable of inhibiting the growth of a cell, especially tumor, e.g., cancer cell, either *in vitro* or *in vivo*. A "growth inhibitory amount" of an anti-TAT antibody, TAT polypeptide, TAT binding oligopeptide or TAT binding organic molecule for purposes of inhibiting neoplastic cell growth may be determined empirically and in a routine manner.

A "cytotoxic amount" of an anti-TAT antibody, TAT polypeptide, TAT binding oligopeptide or TAT binding organic molecule is an amount capable of causing the destruction of a cell, especially tumor, e.g., cancer cell, either *in vitro* or *in vivo*. A "cytotoxic amount" of an anti-TAT antibody, TAT polypeptide, TAT binding oligopeptide or TAT binding organic molecule for purposes of inhibiting neoplastic cell growth may be determined empirically and in a routine manner.

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The term "antibody" is used in the broadest sense and specifically covers, for example, single anti-TAT monoclonal antibodies (including agonist, antagonist, and neutralizing antibodies), anti-TAT antibody compositions with polyepitopic specificity, polyclonal antibodies, single chain anti-TAT antibodies, and fragments of anti-TAT antibodies (see below) as long as they exhibit the desired biological or immunological activity. The term "immunoglobulin" (Ig) is used interchangeable with antibody herein.

An "isolated antibody" is one which has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or nonproteinaceous solutes. In preferred embodiments, the antibody will be purified (1) to greater than 95% by weight of antibody as determined by the Lowry method, and most preferably more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (3) to homogeneity by SDS-PAGE under reducing or nonreducing conditions using Coomassie blue or, preferably, silver stain. Isolated antibody includes the antibody in situ within recombinant cells since at least one component of the antibody's natural environment will not be present. Ordinarily, however, isolated antibody will be prepared by at least one purification step.

The basic 4-chain antibody unit is a heterotetrameric glycoprotein composed of two identical light (L) chains and two identical heavy (H) chains (an IgM antibody consists of 5 of the basic heterotetramer unit along with an additional polypeptide called J chain, and therefore contain 10 antigen binding sites, while secreted IgA antibodies can polymerize to form polyvalent assemblages comprising 2-5 of the basic 4-chain units along with J chain). In the case of IgGs, the 4-chain unit is generally about 150,000 daltons. Each L chain is linked to a H chain by one covalent disulfide bond, while the two H chains are linked to each other by one or more disulfide bonds depending on the H chain isotype. Each H and L chain also has regularly spaced intrachain disulfide bridges. Each H chain has at the N-terminus, a variable domain (V_H) followed by three constant domains (V_H) for each of the v_H and v_H chains and four v_H domains for v_H and v_H is aligned with the v_H and the v_H and the v_H is aligned with the first constant domain of the heavy chain (v_H). Particular amino acid residues are believed to form an interface between the light chain and heavy chain variable domains. The pairing of a v_H and v_H together forms a single antigen-binding site. For the structure and properties of the

different classes of antibodies, see, e.g., <u>Basic and Clinical Immunology</u>, 8th edition, Daniel P. Stites, Abba I. Terr and Tristram G. Parslow (eds.), Appleton & Lange, Norwalk, CT, 1994, page 71 and Chapter 6.

The L chain from any vertebrate species can be assigned to one of two clearly distinct types, called kappa and lambda, based on the amino acid sequences of their constant domains. Depending on the amino acid sequence of the constant domain of their heavy chains (C_H) , immunoglobulins can be assigned to different classes or isotypes. There are five classes of immunoglobulins: IgA, IgD, IgE, IgG, and IgM, having heavy chains designated α , δ , ϵ , γ , and μ , respectively. The γ and α classes are further divided into subclasses on the basis of relatively minor differences in C_H sequence and function, e.g., humans express the following subclasses: IgG1, IgG2, IgG3, IgG4, IgA1, and IgA2.

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The term "variable" refers to the fact that certain segments of the variable domains differ extensively in sequence among antibodies. The V domain mediates antigen binding and define specificity of a particular antibody for its particular antigen. However, the variability is not evenly distributed across the 110-amino acid span of the variable domains. Instead, the V regions consist of relatively invariant stretches called framework regions (FRs) of 15-30 amino acids separated by shorter regions of extreme variability called "hypervariable regions" that are each 9-12 amino acids long. The variable domains of native heavy and light chains each comprise four FRs, largely adopting a β-sheet configuration, connected by three hypervariable regions, which form loops connecting, and in some cases forming part of, the β-sheet structure. The hypervariable regions in each chain are held together in close proximity by the FRs and, with the hypervariable regions from the other chain, contribute to the formation of the antigen-binding site of antibodies (see Kabat et al., Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD. (1991)). The constant domains are not involved directly in binding an antibody to an antigen, but exhibit various effector functions, such as participation of the antibody in antibody dependent cellular cytotoxicity (ADCC).

The term "hypervariable region" when used herein refers to the amino acid residues of an antibody which are responsible for antigen-binding. The hypervariable region generally comprises amino acid residues from a "complementarity determining region" or "CDR" (e.g. around about residues 24-34 (L1), 50-56 (L2) and 89-97 (L3) in the V_L, and around about 1-35 (H1), 50-65 (H2) and 95-102 (H3) in the V_H; Kabat et al., Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD. (1991)) and/or those residues from a "hypervariable loop" (e.g. residues 26-32 (L1), 50-52 (L2) and 91-96 (L3) in the V_L, and 26-32 (H1), 53-55 (H2) and 96-101 (H3) in the V_H; Chothia and Lesk J. Mol. Biol. 196:901-917 (1987)).

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to polyclonal antibody preparations which include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they may be synthesized uncontaminated by other antibodies.

The modifier "monoclonal" is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies useful in the present invention may be prepared by the hybridoma methodology first described by Kohler et al., Nature, 256:495 (1975), or may be made using recombinant DNA methods in bacterial, eukaryotic animal or plant cells (see, e.g., U.S. Patent No. 4,816,567). The "monoclonal antibodies" may also be isolated from phage antibody libraries using the techniques described in Clackson et al., Nature, 352:624-628 (1991) and Marks et al., J. Mol. Biol., 222:581-597 (1991), for example.

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The monoclonal antibodies herein include "chimeric" antibodies in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (see U.S. Patent No. 4,816,567; and Morrison et al., <u>Proc. Natl. Acad. Sci. USA.</u> 81:6851-6855 (1984)). Chimeric antibodies of interest herein include "primatized" antibodies comprising variable domain antigen-binding sequences derived from a non-human primate (e.g. Old World Monkey, Ape etc), and human constant region sequences.

An "intact" antibody is one which comprises an antigen-binding site as well as a C_L and at least heavy chain constant domains, C_H1 , C_H2 and C_H3 . The constant domains may be native sequence constant domains (e.g. human native sequence constant domains) or amino acid sequence variant thereof. Preferably, the intact antibody has one or more effector functions.

"Antibody fragments" comprise a portion of an intact antibody, preferably the antigen binding or variable region of the intact antibody. Examples of antibody fragments include Fab, Fab', F(ab') 2, and Fv fragments; diabodies; linear antibodies (see U.S. Patent No. 5,641,870, Example 2; Zapata et al., <u>Protein Eng.</u> 8(10): 1057-1062 [1995]); single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, and a residual "Fc" fragment, a designation reflecting the ability to crystallize readily. The Fab fragment consists of an entire L chain along with the variable region domain of the H chain (V_H) , and the first constant domain of one heavy chain (C_H1) . Each Fab fragment is monovalent with respect to antigen binding, i.e., it has a single antigen-binding site. Pepsin treatment of an antibody yields a single large F(ab') fragment which roughly corresponds to two disulfide linked Fab fragments having divalent antigen-binding activity and is still capable of cross-linking antigen. Fab' fragments differ from Fab fragments by having additional few residues at the carboxy terminus of the C_H1 domain including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. $F(ab')_2$ antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

The Fc fragment comprises the carboxy-terminal portions of both H chains held together by disulfides.

The effector functions of antibodies are determined by sequences in the Fc region, which region is also the part recognized by Fc receptors (FcR) found on certain types of cells.

"Fv" is the minimum antibody fragment which contains a complete antigen-recognition and -binding site. This fragment consists of a dimer of one heavy- and one light-chain variable region domain in tight, non-covalent association. From the folding of these two domains emanate six hypervariable loops (3 loops each from the H and L chain) that contribute the amino acid residues for antigen binding and confer antigen binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

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"Single-chain Fv" also abbreviated as "sFv" or "scFv" are antibody fragments that comprise the V_H and V_L antibody domains connected into a single polypeptide chain. Preferably, the sFv polypeptide further comprises a polypeptide linker between the V_H and V_L domains which enables the sFv to form the desired structure for antigen binding. For a review of sFv, see Pluckthun in The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenburg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994); Borrebaeck 1995, infra.

The term "diabodies" refers to small antibody fragments prepared by constructing sFv fragments (see preceding paragraph) with short linkers (about 5-10 residues) between the V_H and V_L domains such that interchain but not intra-chain pairing of the V domains is achieved, resulting in a bivalent fragment, i.e., fragment having two antigen-binding sites. Bispecific diabodies are heterodimers of two "crossover" sFv fragments in which the V_H and V_L domains of the two antibodies are present on different polypeptide chains. Diabodies are described more fully in, for example, EP 404,097; WO 93/11161; and Hollinger et al., Proc. Natl. Acad. Sci. USA, 90:6444-6448 (1993).

"Humanized" forms of non-human (e.g., rodent) antibodies are chimeric antibodies that contain minimal sequence derived from the non-human antibody. For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which residues from a hypervariable region of the recipient are replaced by residues from a hypervariable region of a non-human species (donor antibody) such as mouse, rat, rabbit or non-human primate having the desired antibody specificity, affinity, and capability. In some instances, framework region (FR) residues of the human immunoglobulin are replaced by corresponding non-human residues. Furthermore, humanized antibodies may comprise residues that are not found in the recipient antibody or in the donor antibody. These modifications are made to further refine antibody performance. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the hypervariable loops correspond to those of a non-human immunoglobulin and all or substantially all of the FRs are those of a human immunoglobulin sequence. The humanized antibody optionally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. For further details, see Jones et al., Nature 321:522-525 (1986); Riechmann et al., Nature 332:323-329 (1988); and Presta, Curr. Op. Struct. Biol. 2:593-596 (1992).

A "species-dependent antibody," e.g., a mammalian anti-human IgE antibody, is an antibody which

has a stronger binding affinity for an antigen from a first mammalian species than it has for a homologue of that antigen from a second mammalian species. Normally, the species-dependent antibody "bind specifically" to a human antigen (i.e., has a binding affinity (Kd) value of no more than about 1×10^{-7} M, preferably no more than about 1×10^{-8} and most preferably no more than about 1×10^{-9} M) but has a binding affinity for a homologue of the antigen from a second non-human mammalian species which is at least about 50 fold, or at least about 500 fold, or at least about 1000 fold, weaker than its binding affinity for the human antigen. The species-dependent antibody can be of any of the various types of antibodies as defined above, but preferably is a humanized or human antibody.

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A "TAT binding oligopeptide" is an oligopeptide that binds, preferably specifically, to a TAT polypeptide as described herein. TAT binding oligopeptides may be chemically synthesized using known oligopeptide synthesis methodology or may be prepared and purified using recombinant technology. TAT binding oligopeptides are usually at least about 5 amino acids in length, alternatively at least about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, or 100 amino acids in length or more, wherein such oligopeptides that are capable of binding, preferably specifically, to a TAT polypeptide as described herein. TAT binding oligopeptides may be identified without undue experimentation using well known techniques. In this regard, it is noted that techniques for screening oligopeptide libraries for oligopeptides that are capable of specifically binding to a polypeptide target are well known in the art (see, e.g., U.S. Patent Nos. 5,556,762, 5,750,373, 4,708,871, 4,833,092, 5,223,409, 5,403,484, 5,571,689, 5,663,143; PCT Publication Nos. WO 84/03506 and WO84/03564; Geysen et al., Proc. Natl. Acad. Sci. U.S.A., 81:3998-4002 (1984); Geysen et al., Proc. Natl. Acad. Sci. U.S.A., 82:178-182 (1985); Geysen et al., in Synthetic Peptides as Antigens, 130-149 (1986); Geysen et al., J. Immunol. Meth., 102:259-274 (1987); Schoofs et al., J. Immunol., 140:611-616 (1988), Cwirla, S. E. et al. (1990) Proc. Natl. Acad. Sci. USA, 87:6378; Lowman, H.B. et al. (1991) Biochemistry, 30:10832; Clackson, T. et al. (1991) Nature, 352: 624; Marks, J. D. et al. (1991), J. Mol. Biol., 222:581; Kang, A.S. et al. (1991) Proc. Natl. Acad. Sci. USA, 88:8363, and Smith, G. P. (1991) Current Opin. Biotechnol., 2:668).

A "TAT binding organic molecule" is an organic molecule other than an oligopeptide or antibody as defined herein that binds, preferably specifically, to a TAT polypeptide as described herein. TAT binding organic molecules may be identified and chemically synthesized using known methodology (see, e.g., PCT Publication Nos. WO00/00823 and WO00/39585). TAT binding organic molecules are usually less than about 2000 daltons in size, alternatively less than about 1500, 750, 500, 250 or 200 daltons in size, wherein such organic molecules that are capable of binding, preferably specifically, to a TAT polypeptide as described herein may be identified without undue experimentation using well known techniques. In this regard, it is noted that techniques for screening organic molecule libraries for molecules that are capable of binding to a polypeptide target are well known in the art (see, e.g., PCT Publication Nos. WO00/00823 and WO00/39585).

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An antibody, oligopeptide or other organic molecule "which binds" an antigen of interest, e.g. a tumorassociated polypeptide antigen target, is one that binds the antigen with sufficient affinity such that the antibody. oligopeptide or other organic molecule is useful as a diagnostic and/or therapeutic agent in targeting a cell or tissue expressing the antigen, and does not significantly cross-react with other proteins. In such embodiments, the extent of binding of the antibody, oligopeptide or other organic molecule to a "non-target" protein will be less than about 10% of the binding of the antibody, oligopeptide or other organic molecule to its particular target protein as determined by fluorescence activated cell sorting (FACS) analysis or radioimmunoprecipitation (RIA). With regard to the binding of an antibody, oligopeptide or other organic molecule to a target molecule, the term "specific binding" or "specifically binds to" or is "specific for" a particular polypeptide or an epitope on a particular polypeptide target means binding that is measurably different from a non-specific interaction. Specific binding can be measured, for example, by determining binding of a molecule compared to binding of a control molecule, which generally is a molecule of similar structure that does not have binding activity. For example, specific binding can be determined by competition with a control molecule that is similar to the target, for example, an excess of non-labeled target. In this case, specific binding is indicated if the binding of the labeled target to a probe is competitively inhibited by excess unlabeled target. The term "specific binding" or "specifically binds to" or is "specific for" a particular polypeptide or an epitope on a particular polypeptide target as used herein can be exhibited, for example, by a molecule having a Kd for the target of at least about 10⁴ M, alternatively at least about 10⁵ M, alternatively at least about 10⁶ M, alternatively at least about 10⁷ M, alternatively at least about 10⁻⁸ M, alternatively at least about 10⁻¹⁰ M, alternatively at least about 10⁻¹⁰ M, alternatively at least about 10⁻¹¹ M, alternatively at least about 10⁻¹² M, or greater. In one embodiment, the term "specific binding" refers to binding where a molecule binds to a particular polypeptide or epitope on a particular polypeptide without substantially binding to any other polypeptide or polypeptide epitope.

An antibody, oligopeptide or other organic molecule that "inhibits the growth of tumor cells expressing a TAT polypeptide" or a "growth inhibitory" antibody, oligopeptide or other organic molecule is one which results in measurable growth inhibition of cancer cells expressing or overexpressing the appropriate TAT polypeptide. The TAT polypeptide may be a transmembrane polypeptide expressed on the surface of a cancer cell or may be a polypeptide that is produced and secreted by a cancer cell. Preferred growth inhibitory anti-TAT antibodies, oligopeptides or organic molecules inhibit growth of TAT-expressing tumor cells by greater

than 20%, preferably from about 20% to about 50%, and even more preferably, by greater than 50% (e.g., from about 50% to about 100%) as compared to the appropriate control, the control typically being tumor cells not treated with the antibody, oligopeptide or other organic molecule being tested. In one embodiment, growth inhibition can be measured at an antibody concentration of about 0.1 to 30 μ g/ml or about 0.5 nM to 200 nM in cell culture, where the growth inhibition is determined 1-10 days after exposure of the tumor cells to the antibody. Growth inhibition of tumor cells in vivo can be determined in various ways such as is described in the Experimental Examples section below. The antibody is growth inhibitory in vivo if administration of the anti-TAT antibody at about 1 μ g/kg to about 100 mg/kg body weight results in reduction in tumor size or tumor cell proliferation within about 5 days to 3 months from the first administration of the antibody, preferably within about 5 to 30 days.

An antibody, oligopeptide or other organic molecule which "induces apoptosis" is one which induces programmed cell death as determined by binding of annexin V, fragmentation of DNA, cell shrinkage, dilation of endoplasmic reticulum, cell fragmentation, and/or formation of membrane vesicles (called apoptotic bodies). The cell is usually one which overexpresses a TAT polypeptide. Preferably the cell is a tumor cell, e.g., a prostate, breast, ovarian, stomach, endometrial, lung, kidney, colon, bladder cell. Various methods are available for evaluating the cellular events associated with apoptosis. For example, phosphatidyl serine (PS) translocation can be measured by annexin binding; DNA fragmentation can be evaluated through DNA laddering; and nuclear/chromatin condensation along with DNA fragmentation can be evaluated by any increase in hypodiploid cells. Preferably, the antibody, oligopeptide or other organic molecule which induces apoptosis is one which results in about 2 to 50 fold, preferably about 5 to 50 fold, and most preferably about 10 to 50 fold, induction of annexin binding relative to untreated cell in an annexin binding assay.

Antibody "effector functions" refer to those biological activities attributable to the Fc region (a native sequence Fc region or amino acid sequence variant Fc region) of an antibody, and vary with the antibody isotype. Examples of antibody effector functions include: C1q binding and complement dependent cytotoxicity; Fc receptor binding; antibody-dependent cell-mediated cytotoxicity (ADCC); phagocytosis; down regulation of cell surface receptors (e.g., B cell receptor); and B cell activation.

"Antibody-dependent cell-mediated cytotoxicity" or "ADCC" refers to a form of cytotoxicity in which secreted Ig bound onto Fc receptors (FcRs) present on certain cytotoxic cells (e.g., Natural Killer (NK) cells, neutrophils, and macrophages) enable these cytotoxic effector cells to bind specifically to an antigen-bearing target cell and subsequently kill the target cell with cytotoxins. The antibodies "arm" the cytotoxic cells and are absolutely required for such killing. The primary cells for mediating ADCC, NK cells, express Fc γRIII only, whereas monocytes express FcγRI, FcγRII and FcγRIII. FcR expression on hematopoietic cells is summarized in Table 3 on page 464 of Ravetch and Kinet, Annu. Rev. Immunol. 9:457-92 (1991). To assess ADCC activity of a molecule of interest, an in vitro ADCC assay, such as that described in US Patent No. 5,500,362 or 5,821,337 may be performed. Useful effector cells for such assays include peripheral blood mononuclear cells (PBMC) and Natural Killer (NK) cells. Alternatively, or additionally, ADCC activity of the molecule of interest may be assessed in vivo, e.g., in a animal model such as that disclosed in Clynes et al.

(USA) 95:652-656 (1998).

"Fc receptor" or "FcR" describes a receptor that binds to the Fc region of an antibody. The preferred FcR is a native sequence human FcR. Moreover, a preferred FcR is one which binds an IgG antibody (a gamma receptor) and includes receptors of the FcγRI, FcγRII and FcγRIII subclasses, including allelic variants and alternatively spliced forms of these receptors. FcγRII receptors include FcγRIIA (an "activating receptor") and FcγRIIB (an "inhibiting receptor"), which have similar amino acid sequences that differ primarily in the cytoplasmic domains thereof. Activating receptor Fc γRIIA contains an immunoreceptor tyrosine-based activation motif (ITAM) in its cytoplasmic domain. Inhibiting receptor FcγRIIB contains an immunoreceptor tyrosine-based inhibition motif (ITIM) in its cytoplasmic domain. (see review M. in Daëron, Annu. Rev. Immunol. 15:203-234 (1997)). FcRs are reviewed in Ravetch and Kinet, Annu. Rev. Immunol. 9:457-492 (1991); Capel et al., Immunomethods 4:25-34 (1994); and de Haas et al., I. Lab. Clin. Med. 126:330-41 (1995). Other FcRs, including those to be identified in the future, are encompassed by the term "FcR" herein. The term also includes the neonatal receptor, FcRn, which is responsible for the transfer of maternal IgGs to the fetus (Guyer et al., J. Immunol. 117:587 (1976) and Kim et al., J. Immunol. 24:249 (1994)).

"Human effector cells" are leukocytes which express one or more FcRs and perform effector functions. Preferably, the cells express at least Fc γ RIII and perform ADCC effector function. Examples of human leukocytes which mediate ADCC include peripheral blood mononuclear cells (PBMC), natural killer (NK) cells, monocytes, cytotoxic T cells and neutrophils; with PBMCs and NK cells being preferred. The effector cells may be isolated from a native source, e.g., from blood.

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"Complement dependent cytotoxicity" or "CDC" refers to the lysis of a target cell in the presence of complement. Activation of the classical complement pathway is initiated by the binding of the first component of the complement system (C1q) to antibodies (of the appropriate subclass) which are bound to their cognate antigen. To assess complement activation, a CDC assay, e.g., as described in Gazzano-Santoro et al., <u>J. Immunol. Methods</u> 202:163 (1996), may be performed.

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The terms "cancer" and "cancerous" refer to or describe the physiological condition in mammals that is typically characterized by unregulated cell growth. Examples of cancer include, but are not limited to, carcinoma, lymphoma, blastoma, sarcoma, and leukemia or lymphoid malignancies. More particular examples of such cancers include squamous cell cancer (e.g., epithelial squamous cell cancer), lung cancer including small-cell lung cancer, non-small cell lung cancer, adenocarcinoma of the lung and squamous carcinoma of the lung, cancer of the peritoneum, hepatocellular cancer, gastric or stomach cancer including gastrointestinal cancer, pancreatic cancer, glioblastoma, cervical cancer, ovarian cancer, liver cancer, bladder cancer, cancer of the urinary tract, hepatoma, breast cancer, colon cancer, rectal cancer, colorectal cancer, endometrial or uterine carcinoma, salivary gland carcinoma, kidney or renal cancer, prostate cancer, vulval cancer, thyroid cancer, hepatic carcinoma, anal carcinoma, penile carcinoma, melanoma, multiple myeloma and B-cell lymphoma, brain, as well as head and neck cancer, and associated metastases.

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The terms "cell proliferative disorder" and "proliferative disorder" refer to disorders that are

associated with some degree of abnormal cell proliferation. In one embodiment, the cell proliferative disorder is cancer.

"Tumor", as used herein, refers to all neoplastic cell growth and proliferation, whether malignant or benign, and all pre-cancerous and cancerous cells and tissues.

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An antibody, oligopeptide or other organic molecule which "induces cell death" is one which causes a viable cell to become nonviable. The cell is one which expresses a TAT polypeptide, preferably a cell that overexpresses a TAT polypeptide as compared to a normal cell of the same tissue type. The TAT polypeptide may be a transmembrane polypeptide expressed on the surface of a cancer cell or may be a polypeptide that is produced and secreted by a cancer cell. Preferably, the cell is a cancer cell, e.g., a breast, ovarian, stomach, endometrial, salivary gland, lung, kidney, colon, thyroid, pancreatic or bladder cell. Cell death *in vitro* may be determined in the absence of complement and immune effector cells to distinguish cell death induced by antibody-dependent cell-mediated cytotoxicity (ADCC) or complement dependent cytotoxicity (CDC). Thus, the assay for cell death may be performed using heat inactivated serum (i.e., in the absence of complement) and in the absence of immune effector cells. To determine whether the antibody, oligopeptide or other organic molecule is able to induce cell death, loss of membrane integrity as evaluated by uptake of propidium iodide (PI), trypan blue (see Moore et al. Cytotechnology 17:1-11 (1995)) or 7AAD can be assessed relative to untreated cells. Preferred cell death-inducing antibodies, oligopeptides or other organic molecules are those which induce PI uptake in the PI uptake assay in BT474 cells.

A "TAT-expressing cell" is a cell which expresses an endogenous or transfected TAT polypeptide either on the cell surface or in a secreted form. A "TAT-expressing cancer" is a cancer comprising cells that have a TAT polypeptide present on the cell surface or that produce and secrete a TAT polypeptide. A "TATexpressing cancer" optionally produces sufficient levels of TAT polypeptide on the surface of cells thereof, such that an anti-TAT antibody, oligopeptide ot other organic molecule can bind thereto and have a therapeutic effect with respect to the cancer. In another embodiment, a "TAT-expressing cancer" optionally produces and secretes sufficient levels of TAT polypeptide, such that an anti-TAT antibody, oligopeptide ot other organic molecule antagonist can bind thereto and have a therapeutic effect with respect to the cancer. With regard to the latter, the antagonist may be an antisense oligonucleotide which reduces, inhibits or prevents production and secretion of the secreted TAT polypeptide by tumor cells. A cancer which "overexpresses" a TAT polypeptide is one which has significantly higher levels of TAT polypeptide at the cell surface thereof, or produces and secretes, compared to a noncancerous cell of the same tissue type. Such overexpression may be caused by gene amplification or by increased transcription or translation. TAT polypeptide overexpression may be determined in a diagnostic or prognostic assay by evaluating increased levels of the TAT protein present on the surface of a cell, or secreted by the cell (e.g., via an immunohistochemistry assay using anti-TAT antibodies prepared against an isolated TAT polypeptide which may be prepared using recombinant DNA technology from an isolated nucleic acid encoding the TAT polypeptide; FACS analysis, etc.). Alternatively, or additionally, one may measure levels of TAT polypeptide-encoding nucleic acid or mRNA in the cell, e.g., via fluorescent in situ hybridization using a nucleic acid based probe corresponding to a TAT-encoding nucleic acid or the complement

thereof; (FISH; see WO98/45479 published October, 1998), Southern blotting, Northern blotting, or polymerase chain reaction (PCR) techniques, such as real time quantitative PCR (RT-PCR). One may also study TAT polypeptide overexpression by measuring shed antigen in a biological fluid such as serum, e.g, using antibody-based assays (see also, e.g., U.S. Patent No. 4,933,294 issued June 12, 1990; WO91/05264 published April 18, 1991; U.S. Patent 5,401,638 issued March 28, 1995; and Sias et al., J. Immunol. Methods 132:73-80 (1990)). Aside from the above assays, various in vivo assays are available to the skilled practitioner. For example, one may expose cells within the body of the patient to an antibody which is optionally labeled with a detectable label, e.g., a radioactive isotope, and binding of the antibody to cells in the patient can be evaluated, e.g., by external scanning for radioactivity or by analyzing a biopsy taken from a patient previously exposed to the antibody.

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As used herein, the term "immunoadhesin" designates antibody-like molecules which combine the binding specificity of a heterologous protein (an "adhesin") with the effector functions of immunoglobulin constant domains. Structurally, the immunoadhesins comprise a fusion of an amino acid sequence with the desired binding specificity which is other than the antigen recognition and binding site of an antibody (i.e., is "heterologous"), and an immunoglobulin constant domain sequence. The adhesin part of an immunoadhesin molecule typically is a contiguous amino acid sequence comprising at least the binding site of a receptor or a ligand. The immunoglobulin constant domain sequence in the immunoadhesin may be obtained from any immunoglobulin, such as IgG-1, IgG-2, IgG-3, or IgG-4 subtypes, IgA (including IgA-1 and IgA-2), IgE, IgD or IgM.

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The word "label" when used herein refers to a detectable compound or composition which is conjugated directly or indirectly to the antibody, oligopeptide or other organic molecule so as to generate a "labeled" antibody, oligopeptide or other organic molecule. The label may be detectable by itself (e.g. radioisotope labels or fluorescent labels) or, in the case of an enzymatic label, may catalyze chemical alteration of a substrate compound or composition which is detectable.

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The term "cytotoxic agent" as used herein refers to a substance that inhibits or prevents the function of cells and/or causes destruction of cells. The term is intended to include radioactive isotopes (e.g., At, I¹³¹, I¹²⁵, Y⁹⁰, Re¹⁸⁶, Re¹⁸⁸, Sm¹⁵³, Bi²¹², P³² and radioactive isotopes of Lu), chemotherapeutic agents e.g. methotrexate, adriamicin, vinca alkaloids (vincristine, vinblastine, etoposide), doxorubicin, melphalan, mitomycin C, chlorambucil, daunorubicin or other intercalating agents, enzymes and fragments thereof such as nucleolytic enzymes, antibiotics, and toxins such as small molecule toxins or enzymatically active toxins of bacterial, fungal, plant or animal origin, including fragments and/or variants thereof, and the various antitumor or anticancer agents disclosed below. Other cytotoxic agents are described below. A tumoricidal agent causes destruction of tumor cells.

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A "growth inhibitory agent" when used herein refers to a compound or composition which inhibits growth of a cell, especially a TAT-expressing cancer cell, either *in vitro* or *in vivo*. Thus, the growth inhibitory agent may be one which significantly reduces the percentage of TAT-expressing cells in S phase. Examples of growth inhibitory agents include agents that block cell cycle progression (at a place other than S phase), such

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as agents that induce G1 arrest and M-phase arrest. Classical M-phase blockers include the vincas (vincristine and vinblastine), taxanes, and topoisomerase II inhibitors such as doxorubicin, epirubicin, daunorubicin, etoposide, and bleomycin. Those agents that arrest G1 also spill over into S-phase arrest, for example, DNA alkylating agents such as tamoxifen, prednisone, dacarbazine, mechlorethamine, cisplatin, methotrexate, 5-fluorouracil, and ara-C. Further information can be found in Molecular Basis of Cancer, Mendelsohn and Israel, eds., Chapter 1, entitled "Cell cycle regulation, oncogenes, and antineoplastic drugs" by Murakami et al. (WB Saunders: Philadelphia, 1995), especially p. 13. The taxanes (paclitaxel and docetaxel) are anticancer drugs both derived from the yew tree. Docetaxel (TAXOTERE®, Rhone-Poulenc Rorer), derived from the European yew, is a semisynthetic analogue of paclitaxel (TAXOL®, Bristol-Myers Squibb). Paclitaxel and docetaxel promote the assembly of microtubules from tubulin dimers and stabilize microtubules by preventing depolymerization, which results in the inhibition of mitosis in cells.

"Doxorubicin" is an anthracycline antibiotic. The full chemical name of doxorubicin is (8S-cis)-10-[(3-amińo-2,3,6-trideoxy α -L-lyxo-hexapyranosyl)oxy]-7,8,9,10-tetrahydro-6,8,11-trihydroxy-8-(hydroxyacetyl)-1-methoxy-5,12-naphthacenedione.

The term "cytokine" is a generic term for proteins released by one cell population which act on another cell as intercellular mediators. Examples of such cytokines are lymphokines, monokines, and traditional polypeptide hormones. Included among the cytokines are growth hormone such as human growth hormone, Nmethionyl human growth hormone, and bovine growth hormone; parathyroid hormone; thyroxine; insulin; proinsulin; relaxin; prorelaxin; glycoprotein hormones such as follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH), and luteinizing hormone (LH); hepatic growth factor; fibroblast growth factor; prolactin; placental lactogen; tumor necrosis factor-α and -β; mullerian-inhibiting substance; mouse gonadotropin-associated peptide; inhibin; activin; vascular endothelial growth factor; integrin; thrombopoietin (TPO); nerve growth factors such as NGF-β; platelet-growth factor; transforming growth factors (TGFs) such as TGF-α and TGF-β; insulin-like growth factor-I and -II; erythropoietin (EPO); osteoinductive factors; interferons such as interferon -α, -β, and -γ; colony stimulating factors (CSFs) such as macrophage-CSF (M-CSF); granulocyte-macrophage-CSF (GM-CSF); and granulocyte-CSF (G-CSF); interleukins (ILs) such as IL-1, IL- 1a, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-11, IL-12; a tumor necrosis factor such as TNF-α or TNF-B; and other polypeptide factors including LIF and kit ligand (KL). As used herein, the term cytokine includes proteins from natural sources or from recombinant cell culture and biologically active equivalents of the native sequence cytokines.

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The term "package insert" is used to refer to instructions customarily included in commercial packages of therapeutic products, that contain information about the indications, usage, dosage, administration, contraindications and/or warnings concerning the use of such therapeutic products.

Table 1

```
/*
           * C-C increased from 12 to 15
           * Z is average of EQ
 5
           * B is average of ND
           * match with stop is _M; stop-stop = 0; J (joker) match = 0
           */
                                        /* value of a match with a stop */
          #define M
                     day[26][26] = {
10
           int
                  ABCDEFGHIJKLMNOPQRSTUVWXYZ*/
           /*
                      \{2, 0, -2, 0, 0, -4, 1, -1, -1, 0, -1, -2, -1, 0, M, 1, 0, -2, 1, 1, 0, 0, -6, 0, -3, 0\},\
           /* A */
                      { 0, 3,-4, 3, 2,-5, 0, 1,-2, 0, 0,-3,-2, 2, M,-1, 1, 0, 0, 0, 0, -2,-5, 0,-3, 1},
           /* B */
                       \{-2,-4,15,-5,-5,-4,-3,-3,-2,0,-5,-6,-5,-4,\_M,-3,-5,-4,0,-2,0,-2,-8,0,0,-5\},
           /* C */
                       {0, 3,-5, 4, 3,-6, 1, 1,-2, 0, 0,-4,-3, 2, M,-1, 2,-1, 0, 0, 0,-2,-7, 0,-4, 2}, {0, 2,-5, 3, 4,-5, 0, 1,-2, 0, 0,-3,-2, 1, M,-1, 2,-1, 0, 0, 0,-2,-7, 0,-4, 3},
           /* D */
15
           /* E */
                       {-4,-5,-4,-6,-5, 9,-5,-2, 1, 0,-5, 2, 0,-4, M,-5,-5,-4,-3,-3, 0,-1, 0, 0, 7,-5},
           /* F */
                       { 1, 0, -3, 1, 0, -5, 5, -2, -3, 0, -2, -4, -3, 0, M, -1, -1, -3, 1, 0, 0, -1, -7, 0, -5, 0},
           /* G */
                       {-1, 1,-3, 1, 1,-2,-2, 6,-2, 0, 0,-2,-2, 2, M, 0, 3, 2,-1,-1, 0,-2,-3, 0, 0, 2},
           /* H */
                       \{-1, -2, -2, -2, -2, 1, -3, -2, 5, 0, -2, 2, 2, -2, M, -2, -2, -1, 0, 0, 4, -5, 0, -1, -2\},
20
           /* I */
                       /* J */
                       {-1, 0,-5, 0, 0,-5,-2, 0,-2, 0, 5,-3, 0, 1, M,-1, 1, 3, 0, 0, 0,-2,-3, 0,-4, 0},
           /* K */
                       {-2,-3,-6,-4,-3, 2,-4,-2, 2, 0,-3, 6, 4,-3,_M,-3,-2,-3,-3,-1, 0, 2,-2, 0,-1,-2},
           /* L */
                       {-1,-2,-5,-3,-2, 0,-3,-2, 2, 0, 0, 4, 6,-2, M,-2,-1, 0,-2,-1, 0, 2,-4, 0,-2,-1},
           /* M */
                       { 0, 2, 4, 2, 1, 4, 0, 2, -2, 0, 1, -3, -2, 2, M, -1, 1, 0, 1, 0, 0, -2, -4, 0, -2, 1},
25
           /* N */
                       /*O*/
                        \begin{array}{l} \{1,\overline{1},-3,-1,\overline{1},-5,-1,\overline{0},-2,0,\overline{1},-3,-2,\overline{1},\overline{M},6,0,0,1,0,0,-1,-6,0,-5,0\},\\ \{0,1,-5,2,2,-5,-1,3,-2,0,1,-2,-1,1,\overline{M},0,4,1,-1,-1,0,-2,-5,0,-4,3\}. \end{array} 
            /* P */
            /* Q */
                       {-2, 0, 4, -1, -1, -4, -3, 2, -2, 0, 3, -3, 0, 0, M, 0, 1, 6, 0, -1, 0, -2, 2, 0, -4, 0},
            /* R */
                       { 1, 0, 0, 0, 0, -3, 1, -1, -1, 0, 0, -3, -2, 1, M, 1, -1, 0, 2, 1, 0, -1, -2, 0, -3, 0},
30
            /* S */
                        { 1, 0,-2, 0, 0,-3, 0,-1, 0, 0, 0,-1,-1, 0, M, 0,-1,-1, 1, 3, 0, 0,-5, 0,-3, 0},
            /* T */
                        /* U */
            /* V */
                        {-6,-5,-8,-7,-7, 0,-7,-3,-5, 0,-3,-2,-4,-4,_M,-6,-5, 2,-2,-5, 0,-6,17, 0, 0,-6},
            /* W */
                        {0,0,0,0,0,0,0,0,0,0,0,0,0,0,0,0,<u>M</u>,0,0,0,0,0,0,0,0,0,0,0,0,0},
35
            /* X */
                        {-3,-3, 0,-4,-4, 7,-5, 0,-1, 0,-4,-1,-2,-2,_M,-5,-4,-4,-3,-3, 0,-2, 0, 0,10,-4},
            /* Y */
                        { 0, 1,-5, 2, 3,-5, 0, 2,-2, 0, 0,-2,-1, 1, M, 0, 3, 0, 0, 0, 0, -2,-6, 0,-4, 4}
            /* Z */
            };
 40
```

45

50

Table 1 (cont')

```
#include <stdio.h>
           #include <ctype.h>
 5
           #define MAXJMP
                                        16
                                                 /* max jumps in a diag */
           #define MAXGAP
                                                 /* don't continue to penalize gaps larger than this */
                                        24
                                       1024
           #define JMPS
                                                 /* max jmps in an path */
           #define MX
                                        4
                                                 /* save if there's at least MX-1 bases since last jmp */
10
           #define DMAT
                                        3
                                                 /* value of matching bases */
                                       0
           #define DMIS
                                                 /* penalty for mismatched bases */
           #define DINSO
                                       8
                                                 /* penalty for a gap */
           #define DINS1
                                       1
                                                 /* penalty per base */
15
           #define PINSO
                                       8
                                                 /* penalty for a gap */
           #define PINS1
                                                 /* penalty per residue */
           struct jmp {
                                       n[MAXJMP];
                    short
                                                           /* size of jmp (neg for dely) */
20
                    unsigned short
                                       x[MAXJMP];
                                                           /* base no. of jmp in seq x */
          };
                                                           /* limits seq to 2^16 -1 */
          struct diag {
                                                           /* score at last jmp */
                    int
                                       score;
25
                    long
                                       offset;
                                                           /* offset of prev block */
                    short
                                       ijmp;
                                                           /* current jmp index */
                    struct jmp
                                       jp;
                                                           /* list of jmps */
          };
30
          struct path {
                                                 /* number of leading spaces */
                    short
                             n[JMPS]; /* size of jmp (gap) */
                    int
                             x[JMPS]; /* loc of jmp (last elem before gap) */
          };
35
          char
                             *ofile:
                                                           /* output file name */
          char
                             *namex[2];
                                                           /* seq names: getseqs() */
          char
                             *prog;
                                                           /* prog name for err msgs */
          char
                              *seqx[2];
                                                          /* seqs: getseqs() */
40
          int
                             dmax;
                                                          /* best diag: nw() */
                                                          /* final diag */
          int
                             dmax0;
          int
                             dna:
                                                          /* set if dna: main() */
                                                          /* set if penalizing end gaps */
          int
                             endgaps;
          int
                             gapx, gapy;
                                                          /* total gaps in seqs */
45
                                                          /* seq lens */
          int
                             len0, len1;
          int
                             ngapx, ngapy;
                                                          /* total size of gaps */
          int
                                                          /* max score: nw() */
                             smax;
                                                          /* bitmap for matching */
          int
                             *xbm;
          long
                             offset;
                                                          /* current offset in jmp file */
50
          struct
                   diag
                                                          /* holds diagonals */
                             *dx;
          struct
                   path
                             pp[2];
                                                          /* holds path for seqs */
          char
                             *calloc(), *malloc(), *index(), *strcpy();
                             *getseq(), *g_calloc();
          char
```

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```
/* Needleman-Wunsch alignment program
           * usage: progs file1 file2
              where file1 and file2 are two dna or two protein sequences.
 5
              The sequences can be in upper- or lower-case an may contain ambiguity
              Any lines beginning with ';', '>' or '<' are ignored
              Max file length is 65535 (limited by unsigned short x in the jmp struct)
              A sequence with 1/3 or more of its elements ACGTU is assumed to be DNA
               Output is in the file "align.out"
10
           * The program may create a tmp file in /tmp to hold info about traceback.
           * Original version developed under BSD 4.3 on a vax 8650
           #include "nw.h"
15
           #include "day.h"
                     _{dbval[26]} = {
           static
                     1.14.2.13.0.0.4.11,0,0,12,0,3,15,0,0,0,5,6,8,8,7,9,0,10,0
           };
20
                     _pbval[26] = {
           static
                     1, 2|(1 < < ('D'-'A'))|(1 < < ('N'-'A')), 4, 8, 16, 32, 64,
                     128, 256, 0xFFFFFFF, 1 < < 10, 1 < < 11, 1 < < 12, 1 < < 13, 1 < < 14,
                     1<<15, 1<<16, 1<<17, 1<<18, 1<<19, 1<<20, 1<<21, 1<<22,
                     1<<23, 1<<24, 1<<25|(1<<('E'-'A'))|(1<<('Q'-'A'))
25
           };
                                                                                                                                main
           main(ac, av)
                     int
                               ac;
30
                               *av[];
                     char
           {
                     prog = av[0];
                     if (ac! = 3) {
                               fprintf(stderr, "usage: %s file1 file2\n", prog);
fprintf(stderr, "where file1 and file2 are two dna or two protein sequences.\n");
fprintf(stderr, "The sequences can be in upper- or lower-case\n");
35
                               fprintf(stderr, "Any lines beginning with ';' or '<' are ignored\n");
                               fprintf(stderr, "Output is in the file \"align.out\"\n");
                               exit(1);
40
                     namex[0] = av[1];
                     namex[1] = av[2];
                     seqx[0] = getseq(namex[0], \&len0);
                     seqx[1] = getseq(namex[1], &len1);
                     xbm = (dna)? dbval: pbval;
45
                                                    /* 1 to penalize endgaps */
                     endgaps = 0;
                                                              /* output file */
                     ofile = "align.out";
                                         /* fill in the matrix, get the possible jmps */
50
                     nw();
                                          /* get the actual jmps */
                     readjmps();
                                          /* print stats, alignment */
                     print();
                                         /* unlink any tmp files */}
                     cleanup(0);
```

```
/* do the alignment, return best score: main()
            * dna: values in Fitch and Smith, PNAS, 80, 1382-1386, 1983
            * pro: PAM 250 values
            * When scores are equal, we prefer mismatches to any gap, prefer
            * a new gap to extending an ongoing gap, and prefer a gap in seqx
 5
            * to a gap in seq y.
            */
                                                                                                                                      nw
           nw()
           {
                                                               /* seqs and ptrs */
10
                                          *px, *py;
                     char
                                                               /* keep track of dely */
                                          *ndely, *dely;
                     int
                                                               /* keep track of delx */
                                          ndelx, delx;
                     int
                                                               /* for swapping row0, row1 */
                                          *tmp;
                     int
                                                               /* score for each type */
                     int
                                          mis;
                                                               /* insertion penalties */
15
                                          ins0, ins1;
                     int
                                          id;
                                                               /* diagonal index */
                     register
                                                               /* jmp index */
                     register
                                          ij;
                                           *col0, *col1;
                                                               /* score for curr, last row */
                     register
                      register
                                                               /* index into seqs */
                                          xx, yy;
20
                      dx = (struct diag *)g_calloc("to get diags", len0+len1+1, sizeof(struct diag));
                     ndely = (int *)g_calloc("to get ndely", len1+1, sizeof(int));
                     dely = (int *)g_calloc("to get dely", len1 + 1, sizeof(int));

col0 = (int *)g_calloc("to get col0", len1 + 1, sizeof(int));

col1 = (int *)g_calloc("to get col1", len1 + 1, sizeof(int));
25
                      ins0 = (dna)? DINS0 : PINS0;
                      ins1 = (dna)? DINS1: PINS1;
                      smax = -10000;
                      if (endgaps) {
                                for (col0[0] = dely[0] = -ins0, yy = 1; yy <= len1; yy++) {
30
                                          col0[yy] = dely[yy] = col0[yy-1] - ins1;
                                          ndely[yy] = yy;
                                                     /* Waterman Bull Math Biol 84 */
                                col0[0] = 0;
35
                      }
                      else
                                for (yy = 1; yy \le len1; yy++)
                                           dely[yy] = -ins0;
                      /* fill in match matrix
40
                      for (px = seqx[0], xx = 1; xx <= len0; px++, xx++) {
                                /* initialize first entry in col
                                if (endgaps) {
45
                                           if (xx == 1)
                                                     col1[0] = delx = -(ins0+ins1);
                                           else
                                                     col1[0] = delx = col0[0] - ins1;
                                           ndelx = xx;
 50
                                 else {
                                           coi1[0] = 0;
                                           delx = -ins0;
                                           ndelx = 0;
 55
                                 }
```

```
...nw
                            for (py = seqx[1], yy = 1; yy \le len1; py++, yy++) {
                                     mis = col0[yy-1];
                                     if (dna)
                                              mis += (xbm[*px-'A']\&xbm[*py-'A'])? DMAT : DMIS;
 5
                                     else
                                              mis += _day[*px-'A'][*py-'A'];
                                     /* update penalty for del in x seq;
                                      * favor new del over ongong del
10
                                      * ignore MAXGAP if weighting endgaps
                                      if (endgaps | | ndely[yy] < MAXGAP) {
                                               if (col0[yy] - ins0 > = dely[yy]) {
                                                        dely[yy] = col0[yy] - (ins0+ins1);
15
                                                        ndely[yy] = 1;
                                               } else {
                                                        dely[yy] -= ins1;
                                                        ndely[yy]++;
20
                                      } else {
                                               if (col0[yy] - (ins0 + ins1) > = dely[yy]) {
                                                        dely[yy] = col0[yy] - (ins0+ins1);
                                                        ndely[yy] = 1;
25
                                               } else
                                                        ndely[yy]++;
                                      }
                                      /* update penalty for del in y seq;
                                       * favor new del over ongong del
30
                                      if (endgaps | | ndelx < MAXGAP) {
                                               if (col1[yy-1] - ins0 > = delx) {
                                                         delx = col1[yy-1] - (ins0+ins1);
                                                        ndelx = 1;
35
                                               } else {
                                                         delx -= ins1;
                                                         ndelx++;
40
                                      } else {
                                               if (col1[yy-1] - (ins0+ins1) > = delx) {
                                                         delx = coll[yy-1] - (ins0 + ins1);
                                                         ndelx = 1;
                                               } else
                                                         ndelx++;
 45
                                      }
                                      /* pick the maximum score; we're favoring
                                       * mis over any del and delx over dely
 50
                                                                                                                      ...nw
                                      id = xx - yy + len1 - 1;
                                      if (mis > = delx && mis > = dely[yy])
                                                coll[yy] = mis;
 55
```

Table 1 (cont') else if (delx > = dely[yy]) { coll[yy] = delx;ij = dx[id].ijmp;if (dx[id].jp.n[0] && (!dna | | (ndelx > = MAXJMP))&& xx > dx[id].jp.x[ij]+MX) || mis > dx[id].score+DINS0)) { 5 dx[id].ijmp++;if (++ij > = MAXJMP) { writejmps(id); ij = dx[id].ijmp = 0;dx[id].offset = offset;10 offset += sizeof(struct jmp) + sizeof(offset); } dx[id].jp.n[ij] = ndelx;15 dx[id].jp.x[ij] = xx;dx[id].score = delx;else { coll[yy] = dely[yy];20 ij = dx(id).ijmp;if (dx[id].jp.n[0] && (!dna | | (ndely[yy]) > = MAXJMP&& xx > dx[id].jp.x[ij]+MX) || mis > dx[id].score+DINSO)) { dx[id].ijmp++; if (++ij >= MAXJMP) { 25 writejmps(id); ij = dx[id].ijmp = 0;dx[id].offset = offset; offset += sizeof(struct jmp) + sizeof(offset); } 30 dx[id].jp.n[ij] = -ndely[yy];dx[id].jp.x[ij] = xx;dx[id].score = dely[yy]; 35 if (xx == len0 && yy < len1) { /* last col */ if (endgaps) coll[yy] -= ins0 + ins1*(len1-yy);if (coll[yy] > smax) { 40 smax = coll[yy];dmax = id;} } 45 if (endgaps && xx < len0) col1[yy-1] -= ins0 + ins1*(len0-xx);if (coll[yy-1] > smax) { smax = coll[yy-1];dmax = id;50 tmp = col0; col0 = col1; col1 = tmp; . } (void) free((char *)ndely); (void) free((char *)dely); (void) free((char *)col0); (void) free((char *)col1); 55 }

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```
* print() -- only routine visible outside this module
 5
            * getmat() - trace back best path, count matches: print()
            * pr align() -- print alignment of described in array p[]: print()
            * dumpblock() - dump a block of lines with numbers, stars: pr_align()
            * nums() -- put out a number line: dumpblock()
            * putline() -- put out a line (name, [num], seq, [num]): dumpblock()
10
            * stars() - -put a line of stars: dumpblock()
            * stripname() -- strip any path and prefix from a seqname
           #include "nw.h"
15
           #define SPC
                                         /* maximum output line */
           #define P_LINE 256
                                          /* space between name or num and seq */
           #define P_SPC
20
            extern
                      _day[26][26];
                                          /* set output line length */
                     olen;
           int
                                          /* output file */
           FILE
                      *fx;
                                                                                                                                 print
25
           print()
                                                              /* overlap */
                      int
                                lx, ly, firstgap, lastgap;
                      if ((fx = fopen(ofile, "w")) == 0) {
                                fprintf(stderr, "%s: can't write %s\n", prog, ofile);
30
                                cleanup(1);
                      fprintf(fx, "<first sequence: %s (length = %d)\n", namex[0], len0); fprintf(fx, "<second sequence: %s (length = %d)\n", namex[1], len1);
35
                      olen = 60;
                      1x = 1en0;
                      ly = len1;
                      firstgap = lastgap = 0;
                      if (dmax < len1 - 1) {
                                                    /* leading gap in x */
                                pp[0].spc = firstgap = len1 - dmax - 1;
40
                                ly -= pp[0].spc;
                      else if (dmax > len1 - 1) { /* leading gap in y */
                                pp[1].spc = firstgap = dmax - (len1 - 1);
 45
                                lx -= pp[1].spc;
                      if (dmax0 < len0 - 1) {
                                                    /* trailing gap in x */
                                lastgap = len0 - dmax0 - 1;
                                lx -= lastgap;
 50
                      else if (dmax0 > len0 - 1) { /* trailing gap in y */
                                 lastgap = dmax0 - (len0 - 1);
                                ly -= lastgap;
                       getmat(lx, ly, firstgap, lastgap);
 55
                      pr_align();
```

```
* trace back the best path, count matches
            */
           static
  5
           getmat(lx, ly, firstgap, lastgap)
                                                                                                                       getmat
                    int
                                                          /* "core" (minus endgaps) */
                              lx, ly;
                                                          /* leading trailing overlap */
                    int
                              firstgap, lastgap;
           {
                    int
                                       nm, i0, i1, siz0, siz1;
10
                                       outx[32];
                    char
                     double
                                       pct;
                    register
                                       n0, n1;
                    register char
                                        *p0, *p1;
                    /* get total matches, score
15
                     */
                    i0 = i1 = siz0 = siz1 = 0;
                    p0 = seqx[0] + pp[1].spc;
p1 = seqx[1] + pp[0].spc;
                    n0 = pp[1].spc + 1;
20
                    n1 = pp[0].spc + 1;
                    nm = 0;
                    while ( *p0 && *p1 ) {
                              if (siz0) {
                                       p1++;
25
                                       n1++;
                                       siz0--;
                              else if (siz1) {
                                       p0++;
30
                                       n0++;
                                       siz1-;
                              élse {
                                       if (xbm[*p0-'A']&xbm[*p1-'A'])
35
                                                 nm++;
                                       if (n0++==pp[0].x[i0])
                                                 siz0 = pp[0].n[i0++];
                                       if (n1++==pp[1].x[i1])
                                                siz1 = pp[1].n[i1++];
40
                                       p0++;
                                       p1++;
                             }
                    }
45
                    /* pct homology:
                     * if penalizing endgaps, base is the shorter seq
                     * else, knock off overhangs and take shorter core
                    if (endgaps)
50
                             lx = (len0 < len1)? len0 : len1;
                    else
                             lx = (lx < ly)? lx : ly;
                    pct = 100.*(double)nm/(double)lx;
                    fprintf(fx, "\n");
55
                    fprintf(fx, "<%d match%s in an overlap of %d: %.2f percent similarity\n",
                             nm, (nm = = 1)? "": "es", lx, pct);
```

```
...getmat
                     fprintf(fx, "<gaps in first sequence: %d", gapx);
                     if (gapx) {
                               (void) sprintf(outx, " (%d %s%s)",
                                          ngapx, (dna)? "base": "residue", (ngapx == 1)? "": "s");
 5
                                fprintf(fx, "%s", outx);
                     fprintf(fx, ", gaps in second sequence: %d", gapy);
if (gapy) {
                               (void) sprintf(outx, " (%d %s%s)",
                                          ngapy, (dna)? "base": "residue", (ngapy == 1)? "": "s");
10
                                fprintf(fx, "%s", outx);
                     }
if (dna)
                                fprintf(fx,
                                "n < score: %d (match = %d, mismatch = %d, gap penalty = %d + %d per base)n",
15
                                smax, DMAT, DMIS, DINSO, DINS1);
                      else
                                fprintf(fx,
                                "\n < score: %d (Dayhoff PAM 250 matrix, gap penalty = %d + %d per residue)\n",
                                smax, PINSO, PINS1);
20
                      if (endgaps)
                                fprintf(fx,
                                "<endgaps penalized. left endgap: %d %s%s, right endgap: %d %s%s\n", firstgap, (dna)? "base": "residue", (firstgap == 1)? "": "s", lastgap, (dna)? "base": "residue", (lastgap == 1)? "": "s");
25
                      else
                                fprintf(fx, "<endgaps not penalized\n");
            }
                                                     /* matches in core -- for checking */
            static
                                nm;
            static
                                                     /* lengths of stripped file names */
                                lmax;
                                                     /* jmp index for a path */
30
            static
                                ij[2];
                                                     /* number at start of current line */
            static
                                nc[2];
                                                     /* current elem number -- for gapping */
            static
                                ni[2];
            static
                                siz[2];
                                                     /* ptr to current element */
            static char
                                 *ps[2];
                                                     /* ptr to next output char slot */
35
            static char
                                *po[2];
                                out[2][P_LINE];
                                                     /* output line */
            static char
                                star[P_LINE];
                                                     /* set by stars() */
            static char
             * print alignment of described in struct path pp[]
40
            static
                                                                                                                               pr_align
           pr_align()
            {
                                                     /* char count */
                      int
                                           nn;
45
                      int
                                           more;
                      register
                                           i;
                      for (i = 0, lmax = 0; i < 2; i++)
                                nn = stripname(namex[i]);
50
                                if (nn > lmax)
                                           lmax = nn;
                                 nc[i] = 1;
                                ni[i] = 1;
                                 siz[i] = ij[i] = 0;
55
                                 ps[i] = seqx[i];
                                                                          }
                                po[i] = out[i];
```

```
...pr align
                    for (nn = nm = 0, more = 1; more;)
                             for (i = more = 0; i < 2; i++) {
 5
                                        * do we have more of this sequence?
                                        */
                                       if (!*ps[i])
                                                continue;
                                       more++;
10
                                       if (pp[i].spc) { /* leading space */
                                                 *po[i]++ = ' ';
                                                pp[i].spc--;
                                       else if (siz[i]) { /* in a gap */
15
                                                 *po[i]++ = '-';
                                                siz[i]--;
                                       }
                                       else {
                                                          /* we're putting a seq element
                                                          */
20
                                                 *po[i] = *ps[i];
                                                if (islower(*ps[i]))
                                                          *ps[i] = toupper(*ps[i]);
                                                po[i]++;
                                                ps[i]++;
25
                                                 * are we at next gap for this seq?
                                                if (ni[i] == pp[i].x[ij[i]]) {
/*
30
                                                           * we need to merge all gaps
                                                           * at this location
                                                          siz[i] = pp[i].n[ij[i]++];
                                                          while (ni[i] == pp[i].x[ij[i]])

siz[i] += pp[i].n[ij[i]++];
35
                                                ni[i]++;
                                       }
40
                             if (++nn == olen \mid | !more && nn) {
                                       dumpblock();
                                       for (i = 0; i < 2; i++)
                                                po[i] = out[i];
                                       nn = 0;
45
                             }
                    }
          }
/*
           * dump a block of lines, including numbers, stars: pr_align()
50
          static
                                                                                                                dumpblock
          dumpblock()
          {
                    register i;
55
                    for (i = 0; i < 2; i++)
                             po[i] = '0';
```

Table 1 (cont')

...dumpblock

```
(void) putc('\n', fx);
                    for (i = 0; i < 2; i++)
                              if (*out[i] && (*out[i] != ' ' | | *(po[i]) != ' ')) {
 5
                                        if (i == 0)
                                                  nums(i);
                                        if (i == 0 && *out[1])
                                                  stars();
                                        putline(i);
                                        if (i == 0 && *out[1])
10
                                                   fprintf(fx, star);
                                        if (i == 1)
                                                   nums(i);
                              }
15
                    }
           * put out a number line: dumpblock()
20
           static
                                                                                                                              nums
           nums(ix)
                                         /* index in out[] holding seq line */
                     int
                              ix;
           {
                                         nline[P_LINE];
                     char
25
                     register
                                         i, j;
                                         *pn, *px, *py;
                     register char
                     for (pn = nline, i = 0; i < lmax+P_SPC; i++, pn++)
                               *pn = ' ';
                     for (i = nc[ix], py = out[ix]; *py; py++, pn++) {
    if (*py == ' ' | | *py == '-')
        *pn = ' ';
30
                               else {
                                         if (i\%10 == 0 \mid \mid (i == 1 \&\& nc[ix] != 1)) {
                                                   j = (i < 0)? -i : i;
                                                   for (px = pn; j; j /= 10, px-)
35
                                                              px = j\%10 + '0';
                                                             *px = '-';
40
                                         else
                                                    *pn = ' ';
                                         i++;
                               }
                      pn = '0';
45
                     nc[ix] = i;
                     for (pn = nline; *pn; pn++)
                               (void) putc(*pn, fx);
                     (void) putc('\n', fx);
 50
             * put out a line (name, [num], seq, [num]): dumpblock()
            static
                                                                                                                            putline
 55
            putline(ix)
                                                              {
                      int
                               ix;
```

```
...putline
                 int
                 register char
                                   *px;
5
                 for (px = namex[ix], i = 0; *px && *px != ':'; px++, i++)
                          (void) putc(*px, fx);
                 for (; i < lmax+P_SPC; i++)
                          (void) putc(' ', fx);
10
                  /* these count from 1:
                  * ni[] is current element (from 1)
                  * nc[] is number at start of current line
                  for (px = out[ix]; *px; px++)
15
                          (void) putc(*px&0x7F, fx);
                  (void) putc('\n', fx);
         }
20
          * put a line of stars (seqs always in out[0], out[1]): dumpblock()
          static
                                                                                                             stars
25
          stars()
          {
                  int
                                    *p0, *p1, cx, *px;
                  register char
                  30
                           return;
                  px = star;
                  for (i = lmax + P_SPC; i; i-)
                           *px++='';
35
                   for (p0 = out[0], p1 = out[1]; *p0 && *p1; p0++, p1++) {
                           if (isalpha(*p0) && isalpha(*p1)) {
                                    if (xbm[*p0-'A']&xbm[*p1-'A']) {
 40
                                             cx = '*';
                                             nm++;
                                    }
                                    else if (!dna && _day[*p0-'A'][*p1-'A'] > 0)
                                             cx = '.';
 45
                                    else
                                             cx = ' ';
                            else
                                    cx = ' ';
 50
                            *px++=cx;
                   *px++ = '\n';
                   *px = '0';
 55
           }
```

Table 1 (cont')

stripname

```
* cleanup() -- cleanup any tmp file
            * getseq() -- read in seq, set dna, len, maxlen
            * g_calloc() - calloc() with error checkin
           * readjmps() -- get the good jmps, from tmp file if necessary
 5
            * writejmps() -- write a filled array of jmps to a tmp file: nw()
           #include "nw.h"
           #include < sys/file.h>
10
                                                                        /* tmp file for jmps */
                     *jname = "/tmp/homgXXXXXX";
           char
           FILE
                      *fj;
                                                                        /* cleanup tmp file */
           int
                     cleanup();
                     lseek();
           long
15
            * remove any tmp file if we blow
            */
                                                                                                                              cleanup
           cleanup(i)
                                i;
                      int
20
           {
                      if (fj)
                                (void) unlink(jname);
                      exit(i);
25
            * read, return ptr to seq, set dna, len, maxlen
* skip lines starting with ';', '<', or '>'
            * seq in upper or lower case
30
            char
                                                                                                                                getseq
            getseq(file, len)
                                          /* file name */
                                *file;
                      char
                                          /* seq len */
                                *len:
                      int
35
                                          line[1024], *pseq;
                      char
                      register char
                                           *px, *py;
                                          natge, tlen;
                      int
                      FILE
                      if ((fp = fopen(file, "r")) == 0) {
                                fprintf(stderr, "%s: can't read %s\n", prog, file);
 40
                      tlen = natgc = 0;
                      while (fgets(line, 1024, fp)) {
    if (*line == ';' | | *line == '<' | | *line == '>')
 45
                                           continue;
                                 for (px = line; *px != '\n'; px++)
                                           if (isupper(*px) | | islower(*px))
                                                     tlen++;
 50
                       if ((pseq = malloc((unsigned)(tlen+6))) == 0) {
                                 fprintf(stderr, "%s: malloc() failed to get %d bytes for %s\n", prog, tlen+6, file);
                                 exit(1);
                       pseq[0] = pseq[1] = pseq[2] = pseq[3] = '\0';
 55
```

```
...getseq
                    py = pseq + 4;
                    *len = tlen;
                    rewind(fp);
                    while (fgets(line, 1024, fp)) {
    if (*line == ';' || *line == '<' || *line == '>')
 5
                                       continue;
                             for (px = line; *px != '\n'; px++) {
                                       if (isupper(*px))
10
                                                *py++ = *px;
                                       else if (islower(*px))
                                       *py++ = toupper(*px);
if (index("ATGCU",*(py-1)))
                                                natgc++;
15
                             }
                     *py++ = '\0';
                    *py = '\0';
                    (void) fclose(fp);
20
                    dna = natgc > (tlen/3);
                    return(pseq+4);
          char
                                                                                                                      g_calloc
          g_calloc(msg, nx, sz)
25
                                                 /* program, calling routine */
                    char
                             *msg;
                                                 /* number and size of elements */
                    int
                             nx, sz;
           {
                                       *px, *calloc();
                    char
                    if ((px = calloc((unsigned)nx, (unsigned)sz)) == 0) {
30
                              if (*msg) {
                                       fprintf(stderr, "%s: g_calloc() failed %s (n=%d, sz=%d)\n", prog, msg, nx, sz);
                              }
35
                    return(px);
          }
           * get final jmps from dx[] or tmp file, set pp[], reset dmax: main()
40
                                                                                                                   readjmps
          readjmps()
           {
                                       fd = -1;
                    int
                                       siz, i0, i1;
                    int
45
                    register i, j, xx;
                    if (fj) {
                              (void) fclose(fj);
                              if ((fd = open(jname, O_RDONLY, 0)) < 0) {
                                       fprintf(stderr, "%s: can't open() %s\n", prog, jname);
50
                                       cleanup(1);
                              }
                    for (i = i0 = i1 = 0, dmax0 = dmax, xx = len0; ; i++) {
                              while (1) {
55
                                       for (j = dx[dmax].ijmp; j >= 0 && dx[dmax].jp.x[j] >= xx; j--)
```

Table 1 (cont')

...readjmps

```
if (j < 0 && dx[dmax].offset && fj) {
                                                   (void) lseek(fd, dx[dmax].offset, 0);
                                                   (void) read(fd, (char *)&dx[dmax].jp, sizeof(struct jmp));
                                                   (void) read(fd, (char *)&dx[dmax].offset, sizeof(dx[dmax].offset));
 5
                                                   dx[dmax].ijmp = MAXJMP-1;
                                         else
                               if (i > = JMPS) {
                                         fprintf(stderr, "%s: too many gaps in alignment\n", prog);
10
                                         cleanup(1);
                               if (j > = 0) {
                                         siz = dx[dmax].jp.n[j];
15
                                         xx = dx[dmax].jp.x[j];
                                         dmax += siz;
                                         if (siz < 0) {
                                                                       /* gap in second seq */
                                                   pp[1].n[i1] = -siz;
                                                   xx += siz;
                                                                                                                */
20
                                                   /* id = xx - yy + len1 - 1
                                                   pp[1].x[i1] = xx - dmax + len1 - 1;
                                                   gapy++;
                                                   ngapy -= siz;
           /* ignore MAXGAP when doing endgaps */
                                                   siz = (-siz < MAXGAP | | endgaps)? -siz : MAXGAP;
25
                                         else if (siz > 0) { /* gap in first seq */
                                                   pp[0].n[i0] = siz;
                                                   pp[0].x[i0] = xx;
30
                                                   gapx++;
                                                   ngapx += siz;
           /* ignore MAXGAP when doing endgaps */
                                                   siz = (siz < MAXGAP | | endgaps)? siz : MAXGAP;
35
                                         }
                               }
                                else
                                         break;
40
                     /* reverse the order of jmps */
                     for (j = 0, i0 -; j < i0; j++, i0-) \{

i = pp[0].n[j]; pp[0].n[j] = pp[0].n[i0]; pp[0].n[i0] = i;
                                i = pp[0].x[i]; pp[0].x[i] = pp[0].x[i0]; pp[0].x[i0] = i;
45
                     for (j = 0, i1-; j < i1; j++, i1-)
                               i = pp[1].n[j]; pp[1].n[j] = pp[1].n[i1]; pp[1].n[i1] = i;

i = pp[1].x[j]; pp[1].x[j] = pp[1].x[i1]; pp[1].x[i1] = i;
50
                     if (fd >= 0)
                               (void) close(fd);
                     if (fj) {
                                (void) unlink(jname);
                                f_j = 0;
55
                                offset = 0;
                                                             }
                     }
```

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```
* write a filled jmp struct offset of the prev one (if any): nw()
                                                                                                                                                                      writejmps
 5
               writejmps(ix)
                                           ix;
               {
                             char
                                           *mktemp();
                             if (!fj) {
                                           if (mktemp(jname) < 0) {
          fprintf(stderr, "%s: can't mktemp() %s\n", prog, jname);
          cleanup(1);</pre>
10
                                          }
if ((fj = fopen(jname, "w")) == 0) {
    fprintf(stderr, "%s: can't write %s\n", prog, jname);
15
                                           }
                             }
(void) fwrite((char *)&dx[ix].jp, sizeof(struct jmp), 1, fj);
(void) fwrite((char *)&dx[ix].offset, sizeof(dx[ix].offset), 1, fj);
20
                }
```

Table 2

TAT XXXXXXXXXXXXXXX (Length = 15 amino acids)

5 % amino acid sequence identity =

(the number of identically matching amino acid residues between the two polypeptide sequences as determined by ALIGN-2) divided by (the total number of amino acid residues of the TAT polypeptide) =

10 5 divided by 15 = 33.3%

Table 3

TAT XXXXXXXXXX (Length = 10 amino acids)

15 Comparison Protein XXXXXYYYYYYZZYZ (Length = 15 amino acids)

% amino acid sequence identity =

(the number of identically matching amino acid residues between the two polypeptide sequences as determined by ALIGN-2) divided by (the total number of amino acid residues of the TAT polypeptide) =

5 divided by 10 = 50%

Table 4

25

30

TAT-DNA NNNNNNNNNNN (Length = 14 nucleotides)

Comparison DNA NNNNNLLLLLLLLL (Length = 16 nucleotides)

% nucleic acid sequence identity =

(the number of identically matching nucleotides between the two nucleic acid sequences as determined by ALIGN-2) divided by (the total number of nucleotides of the TAT-DNA nucleic acid sequence) = 6 divided by 14 = 42.9%

Table 5

TAT-DNA

 (Length = 12 nucleotides)
(Length = 9 nucleotides)

Comparison DNA

5 % nucleic acid sequence identity =

(the number of identically matching nucleotides between the two nucleic acid sequences as determined by ALIGN-2) divided by (the total number of nucleotides of the TAT-DNA nucleic acid sequence) =

10 4 divided by 12 = 33.3%

15

20

25

30

35

II. Compositions and Methods of the Invention

A. Anti-TAT Antibodies

In one embodiment, the present invention provides anti-TAT antibodies which may find use herein as therapeutic and/or diagnostic agents. Exemplary antibodies include polyclonal, monoclonal, humanized, bispecific, and heteroconjugate antibodies.

1. Polyclonal Antibodies

Polyclonal antibodies are preferably raised in animals by multiple subcutaneous (sc) or intraperitoneal (ip) injections of the relevant antigen and an adjuvant. It may be useful to conjugate the relevant antigen (especially when synthetic peptides are used) to a protein that is immunogenic in the species to be immunized. For example, the antigen can be conjugated to keyhole limpet hemocyanin (KLH), serum albumin, bovine thyroglobulin, or soybean trypsin inhibitor, using a bifunctional or derivatizing agent, e.g., maleimidobenzoyl sulfosuccinimide ester (conjugation through cysteine residues), N-hydroxysuccinimide (through lysine residues), glutaraldehyde, succinic anhydride, SOCl₂, or R¹N=C=NR, where R and R¹ are different alkyl groups.

Animals are immunized against the antigen, immunogenic conjugates, or derivatives by combining, e.g., 100 µg or 5 µg of the protein or conjugate (for rabbits or mice, respectively) with 3 volumes of Freund's complete adjuvant and injecting the solution intradermally at multiple sites. One month later, the animals are boosted with 1/5 to 1/10 the original amount of peptide or conjugate in Freund's complete adjuvant by subcutaneous injection at multiple sites. Seven to 14 days later, the animals are bled and the serum is assayed for antibody titer. Animals are boosted until the titer plateaus. Conjugates also can be made in recombinant cell culture as protein fusions. Also, aggregating agents such as alum are suitably used to enhance the immune

response.

2. Monoclonal Antibodies

Monoclonal antibodies may be made using the hybridoma method first described by Kohler et al., Nature, 256:495 (1975), or may be made by recombinant DNA methods (U.S. Patent No. 4,816,567).

In the hybridoma method, a mouse or other appropriate host animal, such as a hamster, is immunized

as described above to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the protein used for immunization. Alternatively, lymphocytes may be immunized *in vitro*. After immunization, lymphocytes are isolated and then fused with a myeloma cell line using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell (Goding, Monoclonal Antibodies: Principles and Practice, pp.59-103 (Academic Press, 1986)).

5

The hybridoma cells thus prepared are seeded and grown in a suitable culture medium which medium preferably contains one or more substances that inhibit the growth or survival of the unfused, parental myeloma cells (also referred to as fusion partner). For example, if the parental myeloma cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the selective culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine (HAT medium), which substances prevent the growth of HGPRT-deficient cells.

10

Preferred fusion partner myeloma cells are those that fuse efficiently, support stable high-level production of antibody by the selected antibody-producing cells, and are sensitive to a selective medium that selects against the unfused parental cells. Preferred myeloma cell lines are murine myeloma lines, such as those derived from MOPC-21 and MPC-11 mouse tumors available from the Salk Institute Cell Distribution Center, San Diego, California USA, and SP-2 and derivatives e.g., X63-Ag8-653 cells available from the American Type Culture Collection, Manassas, Virginia, USA. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies (Kozbor, J. Immunol., 133:3001 (1984); and Brodeur et al., Monoclonal Antibody Production Techniques and Applications, pp. 51-63 (Marcel Dekker, Inc., New York, 1987)).

20

15

Culture medium in which hybridoma cells are growing is assayed for production of monoclonal antibodies directed against the antigen. Preferably, the binding specificity of monoclonal antibodies produced by hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunosorbent assay (ELISA).

25

The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis described in Munson et al., <u>Anal. Biochem.</u>, 107:220 (1980).

30

Once hybridoma cells that produce antibodies of the desired specificity, affinity, and/or activity are identified, the clones may be subcloned by limiting dilution procedures and grown by standard methods (Goding, Monoclonal Antibodies: Principles and Practice, pp.59-103 (Academic Press, 1986)). Suitable culture media for this purpose include, for example, D-MEM or RPMI-1640 medium. In addition, the hybridoma cells may be grown *in vivo* as ascites tumors in an animal e.g., by i.p. injection of the cells into mice.

The monoclonal antibodies secreted by the subclones are suitably separated from the culture medium, ascites fluid, or serum by conventional antibody purification procedures such as, for example, affinity chromatography (e.g., using protein A or protein G-Sepharose) or ion-exchange chromatography, hydroxylapatite chromatography, gel electrophoresis, dialysis, etc.

35

DNA encoding the monoclonal antibodies is readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the

heavy and light chains of murine antibodies). The hybridoma cells serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as *E. coli* cells, simian COS cells, Chinese Hamster Ovary (CHO) cells, or myeloma cells that do not otherwise produce antibody protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. Review articles on recombinant expression in bacteria of DNA encoding the antibody include Skerra et al., <u>Curr. Opinion in Immunol.</u>, 5:256-262 (1993) and Plückthun, <u>Immunol. Revs.</u> 130:151-188 (1992).

In a further embodiment, monoclonal antibodies or antibody fragments can be isolated from antibody phage libraries generated using the techniques described in McCafferty et al., Nature, 348:552-554 (1990). Clackson et al., Nature, 352:624-628 (1991) and Marks et al., J. Mol. Biol., 222:581-597 (1991) describe the isolation of murine and human antibodies, respectively, using phage libraries. Subsequent publications describe the production of high affinity (nM range) human antibodies by chain shuffling (Marks et al., Bio/Technology, 10:779-783 (1992)), as well as combinatorial infection and in vivo recombination as a strategy for constructing very large phage libraries (Waterhouse et al., Nuc. Acids. Res. 21:2265-2266 (1993)). Thus, these techniques are viable alternatives to traditional monoclonal antibody hybridoma techniques for isolation of monoclonal antibodies.

15

20

10

5

The DNA that encodes the antibody may be modified to produce chimeric or fusion antibody polypeptides, for example, by substituting human heavy chain and light chain constant domain (C _H and C_L) sequences for the homologous murine sequences (U.S. Patent No. 4,816,567; and Morrison, et al., <u>Proc. Natl Acad. Sci. USA</u>, 81:6851 (1984)), or by fusing the immunoglobulin coding sequence with all or part of the coding sequence for a non-immunoglobulin polypeptide (heterologous polypeptide). The non-immunoglobulin polypeptide sequences can substitute for the constant domains of an antibody, or they are substituted for the variable domains of one antigen-combining site of an antibody to create a chimeric bivalent antibody comprising one antigen-combining site having specificity for an antigen and another antigen-combining site having specificity for a different antigen.

3. Human and Humanized Antibodies

25

30

35

The anti-TAT antibodies of the invention may further comprise humanized antibodies or human antibodies. Humanized forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', F(ab') 2 or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. Humanized antibodies include human immunoglobulins (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, Fv framework residues of the human immunoglobulin are replaced by corresponding non-human residues. Humanized antibodies may also comprise residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are

those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin [Jones et al., Nature, 321:522-525 (1986); Riechmann et al., Nature, 332:323-329 (1988); and Presta, Curr. Op. Struct. Biol., 2:593-596 (1992)].

Methods for humanizing non-human antibodies are well known in the art. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source which is non-human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers [Jones et al., Nature, 321:522-525 (1986); Riechmann et al., Nature, 332:323-327 (1988); Verhoeyen et al., Science, 239:1534-1536 (1988)], by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Patent No. 4,816,567), wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

The choice of human variable domains, both light and heavy, to be used in making the humanized antibodies is very important to reduce antigenicity and HAMA response (human anti-mouse antibody) when the antibody is intended for human therapeutic use. According to the so-called "best-fit" method, the sequence of the variable domain of a rodent antibody is screened against the entire library of known human variable domain sequences. The human V domain sequence which is closest to that of the rodent is identified and the human framework region (FR) within it accepted for the humanized antibody (Sims et al., J. Immunol. 151:2296 (1993); Chothia et al., J. Mol. Biol., 196:901 (1987)). Another method uses a particular framework region derived from the consensus sequence of all human antibodies of a particular subgroup of light or heavy chains. The same framework may be used for several different humanized antibodies (Carter et al., Proc. Natl. Acad. Sci. USA, 89:4285 (1992); Presta et al., J. Immunol. 151:2623 (1993)).

It is further important that antibodies be humanized with retention of high binding affinity for the antigen and other favorable biological properties. To achieve this goal, according to a preferred method, humanized antibodies are prepared by a process of analysis of the parental sequences and various conceptual humanized products using three-dimensional models of the parental and humanized sequences. Three-dimensional immunoglobulin models are commonly available and are familiar to those skilled in the art. Computer programs are available which illustrate and display probable three-dimensional conformational structures of selected candidate immunoglobulin sequences. Inspection of these displays permits analysis of the likely role of the residues in the functioning of the candidate immunoglobulin sequence, i.e., the analysis of residues that influence the ability of the candidate immunoglobulin to bind its antigen. In this way, FR residues can be selected and combined from the recipient and import sequences so that the desired antibody characteristic, such as increased affinity for the target antigen(s), is achieved. In general, the hypervariable region residues are directly and most substantially involved in influencing antigen binding.

Various forms of a humanized anti-TAT antibody are contemplated. For example, the humanized antibody may be an antibody fragment, such as a Fab, which is optionally conjugated with one or more cytotoxic agent(s) in order to generate an immunoconjugate. Alternatively, the humanized antibody may be an intact antibody, such as an intact IgG1 antibody.

As an alternative to humanization, human antibodies can be generated. For example, it is now possible to produce transgenic animals (e.g., mice) that are capable, upon immunization, of producing a full repertoire of human antibodies in the absence of endogenous immunoglobulin production. For example, it has been described that the homozygous deletion of the antibody heavy-chain joining region (J_H) gene in chimeric and germ-line mutant mice results in complete inhibition of endogenous antibody production. Transfer of the human germ-line immunoglobulin gene array into such germ-line mutant mice will result in the production of human antibodies upon antigen challenge. See, e.g., Jakobovits et al., <u>Proc. Natl. Acad. Sci. USA</u>, 90:2551 (1993); Jakobovits et al., <u>Nature</u>, 362:255-258 (1993); Bruggemann et al., <u>Year in Immuno.</u> 7:33 (1993); U.S. Patent Nos. 5,545,806, 5,569,825, 5,591,669 (all of GenPharm); 5,545,807; and WO 97/17852.

Alternatively, phage display technology (McCafferty et al., Nature 348:552-553 [1990]) can be used to produce human antibodies and antibody fragments in vitro, from immunoglobulin variable (V) domain gene repertoires from unimmunized donors. According to this technique, antibody V domain genes are cloned inframe into either a major or minor coat protein gene of a filamentous bacteriophage, such as M13 or fd, and displayed as functional antibody fragments on the surface of the phage particle. Because the filamentous particle contains a single-stranded DNA copy of the phage genome, selections based on the functional properties of the antibody also result in selection of the gene encoding the antibody exhibiting those properties. Thus, the phage mimics some of the properties of the B-cell. Phage display can be performed in a variety of formats, reviewed in, e.g., Johnson, Kevin S. and Chiswell, David J., Current Opinion in Structural Biology 3:564-571 (1993). Several sources of V-gene segments can be used for phage display. Clackson et al. Nature, 352:624-628 (1991) isolated a diverse array of anti-oxazolone antibodies from a small random combinatorial library of V genes derived from the spleens of immunized mice. A repertoire of V genes from unimmunized human donors can be constructed and antibodies to a diverse array of antigens (including self-antigens) can be isolated essentially following the techniques described by Marks et al., J. Mol. Biol. 222:581-597 (1991), or Griffith et al., EMBO J. 12:725-734 (1993). See, also, U.S. Patent Nos. 5,565,332 and 5,573,905.

As discussed above, human antibodies may also be generated by *in vitro* activated B cells (see U.S. Patents 5,567,610 and 5,229,275).

4. Antibody fragments

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In certain circumstances there are advantages of using antibody fragments, rather than whole antibodies. The smaller size of the fragments allows for rapid clearance, and may lead to improved access to solid tumors.

Various techniques have been developed for the production of antibody fragments. Traditionally, these fragments were derived via proteolytic digestion of intact antibodies (see, e.g., Morimoto et al., <u>Journal of</u> Biochemical and <u>Biophysical Methods</u> 24:107-117 (1992); and Brennan et al., <u>Science</u>, 229:81 (1985)).

However, these fragments can now be produced directly by recombinant host cells. Fab, Fv and ScFv antibody fragments can all be expressed in and secreted from *E. coli*, thus allowing the facile production of large amounts of these fragments. Antibody fragments can be isolated from the antibody phage libraries discussed above. Alternatively, Fab'-SH fragments can be directly recovered from *E. coli* and chemically coupled to form F(ab')₂ fragments (Carter et al., Bio/Technology 10:163-167 (1992)). According to another approach, F(ab') 2 fragments can be isolated directly from recombinant host cell culture. Fab and F(ab') fragment with increased in vivo half-life comprising a salvage receptor binding epitope residues are described in U.S. Patent No. 5,869,046. Other techniques for the production of antibody fragments will be apparent to the skilled practitioner. In other embodiments, the antibody of choice is a single chain Fv fragment (scFv). See WO 93/16185; U.S. Patent No. 5,571,894; and U.S. Patent No. 5,587,458. Fv and sFv are the only species with intact combining sites that are devoid of constant regions; thus, they are suitable for reduced nonspecific binding during in vivo use. sFv fusion proteins may be constructed to yield fusion of an effector protein at either the amino or the carboxy terminus of an sFv. See Antibody Engineering, ed. Borrebaeck, supra. The antibody fragment may also be a "linear antibody", e.g., as described in U.S. Patent 5,641,870 for example. Such linear antibody fragments may be monospecific or bispecific.

5. Bispecific Antibodies

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Bispecific antibodies are antibodies that have binding specificities for at least two different epitopes. Exemplary bispecific antibodies may bind to two different epitopes of a TAT protein as described herein. Other such antibodies may combine a TAT binding site with a binding site for another protein. Alternatively, an anti-TAT arm may be combined with an arm which binds to a triggering molecule on a leukocyte such as a T-cell receptor molecule (e.g. CD3), or Fc receptors for IgG (FcγR), such as FcγRI (CD64), FcγRII (CD32) and FcγRII (CD16), so as to focus and localize cellular defense mechanisms to the TAT-expressing cell. Bispecific antibodies may also be used to localize cytotoxic agents to cells which express TAT. These antibodies possess a TAT-binding arm and an arm which binds the cytotoxic agent (e.g., saporin, anti-interferon-α, vinca alkaloid, ricin A chain, methotrexate or radioactive isotope hapten). Bispecific antibodies can be prepared as full length antibodies or antibody fragments (e.g., F(ab')₂ bispecific antibodies).

WO 96/16673 describes a bispecific anti-ErbB2/anti-Fc γ RIII antibody and U.S. Patent No. 5,837,234 discloses a bispecific anti-ErbB2/anti-Fc γ RI antibody. A bispecific anti-ErbB2/Fc α antibody is shown in WO98/02463. U.S. Patent No. 5,821,337 teaches a bispecific anti-ErbB2/anti-CD3 antibody.

Methods for making bispecific antibodies are known in the art. Traditional production of full length bispecific antibodies is based on the co-expression of two immunoglobulin heavy chain-light chain pairs, where the two chains have different specificities (Millstein et al., Nature 305:537-539 (1983)). Because of the random assortment of immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of 10 different antibody molecules, of which only one has the correct bispecific structure. Purification of the correct molecule, which is usually done by affinity chromatography steps, is rather cumbersome, and the product yields are low. Similar procedures are disclosed in WO 93/08829, and in Traunecker et al., EMBO J. 10:3655-3659 (1991).

According to a different approach, antibody variable domains with the desired binding specificities (antibody-antigen combining sites) are fused to immunoglobulin constant domain sequences. Preferably, the fusion is with an Ig heavy chain constant domain, comprising at least part of the hinge, C_H2 , and C_H3 regions. It is preferred to have the first heavy-chain constant region (C_H1) containing the site necessary for light chain bonding, present in at least one of the fusions. DNAs encoding the immunoglobulin heavy chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host cell. This provides for greater flexibility in adjusting the mutual proportions of the three polypeptide fragments in embodiments when unequal ratios of the three polypeptide chains used in the construction provide the optimum yield of the desired bispecific antibody. It is, however, possible to insert the coding sequences for two or all three polypeptide chains into a single expression vector when the expression of at least two polypeptide chains in equal ratios results in high yields or when the ratios have no significant affect on the yield of the desired chain combination.

In a preferred embodiment of this approach, the bispecific antibodies are composed of a hybrid immunoglobulin heavy chain with a first binding specificity in one arm, and a hybrid immunoglobulin heavy chain-light chain pair (providing a second binding specificity) in the other arm. It was found that this asymmetric structure facilitates the separation of the desired bispecific compound from unwanted immunoglobulin chain combinations, as the presence of an immunoglobulin light chain in only one half of the bispecific molecule provides for a facile way of separation. This approach is disclosed in WO 94/04690. For further details of generating bispecific antibodies see, for example, Suresh et al., Methods in Enzymology 121:210 (1986).

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According to another approach described in U.S. Patent No. 5,731,168, the interface between a pair of antibody molecules can be engineered to maximize the percentage of heterodimers which are recovered from recombinant cell culture. The preferred interface comprises at least a part of the C_H3 domain. In this method, one or more small amino acid side chains from the interface of the first antibody molecule are replaced with larger side chains (e.g., tyrosine or tryptophan). Compensatory "cavities" of identical or similar size to the large side chain(s) are created on the interface of the second antibody molecule by replacing large amino acid side chains with smaller ones (e.g., alanine or threonine). This provides a mechanism for increasing the yield of the heterodimer over other unwanted end-products such as homodimers.

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Bispecific antibodies include cross-linked or "heteroconjugate" antibodies. For example, one of the antibodies in the heteroconjugate can be coupled to avidin, the other to biotin. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells (U.S. Patent No. 4,676,980), and for treatment of HIV infection (WO 91/00360, WO 92/200373, and EP 03089). Heteroconjugate antibodies may be made using any convenient cross-linking methods. Suitable cross-linking agents are well known in the art, and are disclosed in U.S. Patent No. 4,676,980, along with a number of cross-linking techniques.

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Techniques for generating bispecific antibodies from antibody fragments have also been described in the literature. For example, bispecific antibodies can be prepared using chemical linkage. Brennan et al., Science 229:81 (1985) describe a procedure wherein intact antibodies are proteolytically cleaved to generate

F(ab')₂ fragments. These fragments are reduced in the presence of the dithiol complexing agent, sodium arsenite, to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The Fab' fragments generated are then converted to thionitrobenzoate (TNB) derivatives. One of the Fab'-TNB derivatives is then reconverted to the Fab'-thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other Fab'-TNB derivative to form the bispecific antibody. The bispecific antibodies produced can be used as agents for the selective immobilization of enzymes.

Recent progress has facilitated the direct recovery of Fab'-SH fragments from E. coli, which can be chemically coupled to form bispecific antibodies. Shalaby et al., J. Exp. Med. 175: 217-225 (1992) describe the production of a fully humanized bispecific antibody F(ab')₂ molecule. Each Fab' fragment was separately secreted from E. coli and subjected to directed chemical coupling in vitro to form the bispecific antibody. The bispecific antibody thus formed was able to bind to cells overexpressing the ErbB2 receptor and normal human T cells, as well as trigger the lytic activity of human cytotoxic lymphocytes against human breast tumor targets.

Various techniques for making and isolating bispecific antibody fragments directly from recombinant cell culture have also been described. For example, bispecific antibodies have been produced using leucine zippers. Kostelny et al., J. Immunol., 148(5):1547-1553 (1992). The leucine zipper peptides from the Fos and Jun proteins were linked to the Fab' portions of two different antibodies by gene fusion. The antibody homodimers were reduced at the hinge region to form monomers and then re-oxidized to form the antibody heterodimers. This method can also be utilized for the production of antibody homodimers. The "diabody" technology described by Hollinger et al., Proc. Natl. Acad. Sci. USA 90:6444-6448 (1993) has provided an alternative mechanism for making bispecific antibody fragments. The fragments comprise a V_H connected to a V_L by a linker which is too short to allow pairing between the two domains on the same chain. Accordingly, the V_H and V_L domains of one fragment are forced to pair with the complementary V_L and V_H domains of another fragment, thereby forming two antigen-binding sites. Another strategy for making bispecific antibody fragments by the use of single-chain Fv (sFv) dimers has also been reported. See Gruber et al., J. Immunol., 152:5368 (1994).

Antibodies with more than two valencies are contemplated. For example, trispecific antibodies can be prepared. Tutt et al., <u>J. Immunol.</u> 147:60 (1991).

6. Heteroconjugate Antibodies

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Heteroconjugate antibodies are also within the scope of the present invention. Heteroconjugate antibodies are composed of two covalently joined antibodies. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells [U.S. Patent No. 4,676,980], and for treatment of HIV infection [WO 91/00360; WO 92/200373; EP 03089]. It is contemplated that the antibodies may be prepared in vitro using known methods in synthetic protein chemistry, including those involving crosslinking agents. For example, immunotoxins may be constructed using a disulfide exchange reaction or by forming a thioether bond. Examples of suitable reagents for this purpose include iminothiolate and methyl-4-mercaptobutyrimidate and those disclosed, for example, in U.S. Patent No. 4,676,980.

7. Multivalent Antibodies

A multivalent antibody may be internalized (and/or catabolized) faster than a bivalent antibody by a cell expressing an antigen to which the antibodies bind. The antibodies of the present invention can be multivalent antibodies (which are other than of the IgM class) with three or more antigen binding sites (e.g. tetravalent antibodies), which can be readily produced by recombinant expression of nucleic acid encoding the polypeptide chains of the antibody. The multivalent antibody can comprise a dimerization domain and three or more antigen binding sites. The preferred dimerization domain comprises (or consists of) an Fc region or a hinge region. In this scenario, the antibody will comprise an Fc region and three or more antigen binding sites amino-terminal to the Fc region. The preferred multivalent antibody herein comprises (or consists of) three to about eight, but preferably four, antigen binding sites. The multivalent antibody comprises at least one polypeptide chain (and preferably two polypeptide chains), wherein the polypeptide chain(s) comprise two or more variable domains. For instance, the polypeptide chain(s) may comprise VD1-(X1) _n-VD2-(X2)_n-Fc, wherein VD1 is a first variable domain, VD2 is a second variable domain, Fc is one polypeptide chain of an Fc region, X1 and X2 represent an amino acid or polypeptide, and n is 0 or 1. For instance, the polypeptide chain(s) may comprise: VH-CH1-flexible linker-VH-CH1-Fc region chain; or VH-CH1-VH-CH1-Fc region chain. The multivalent antibody herein preferably further comprises at least two (and preferably four) light chain variable domain polypeptides. The multivalent antibody herein may, for instance, comprise from about two to about eight light chain variable domain polypeptides. The light chain variable domain polypeptides contemplated here comprise a light chain variable domain and, optionally, further comprise a CL domain.

8. Effector Function Engineering

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as to enhance antigen-dependent cell-mediated cyotoxicity (ADCC) and/or complement dependent cytotoxicity (CDC) of the antibody. This may be achieved by introducing one or more amino acid substitutions in an Fc region of the antibody. Alternatively or additionally, cysteine residue(s) may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). See Caron et al., J. Exp Med. 176:1191-1195 (1992) and Shopes, B. J. Immunol. 148:2918-2922 (1992). Homodimeric antibodies with enhanced anti-tumor activity may also be prepared using heterobifunctional cross-linkers as described in Wolff et al., Cancer Research 53:2560-2565 (1993). Alternatively, an antibody can be engineered which has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. See Stevenson et al., Anti-Cancer Drug Design 3:219-230 (1989).

To increase the serum half life of the antibody, one may incorporate a salvage receptor binding epitope into the antibody (especially an antibody fragment) as described in U.S. Patent 5,739,277, for example. As used herein, the term "salvage receptor binding epitope" refers to an epitope of the Fc region of an IgG molecule (e.g., IgG₁, IgG₂, IgG₃, or IgG₄) that is responsible for increasing the *in vivo* serum half-life of the IgG molecule.

9. <u>Immunoconjugates</u>

The invention also pertains to immunoconjugates comprising an antibody conjugated to a cytotoxic

agent such as a chemotherapeutic agent, a growth inhibitory agent, a toxin (e.g., an enzymatically active toxin of bacterial, fungal, plant, or animal origin, or fragments thereof), or a radioactive isotope (i.e., a radioconiugate).

Chemotherapeutic agents useful in the generation of such immunoconjugates have been described above. Enzymatically active toxins and fragments thereof that can be used include diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from Pseudomonas aeruginosa), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, Aleurites fordii proteins, dianthin proteins, Phytolaca americana proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcin, crotin, sapaonaria officinalis inhibitor, gelonin, mitogellin, restrictocin, phenomycin, enomycin, and the tricothecenes. A variety of radionuclides are available for the production of radioconjugated antibodies. Examples include 212Bi, 131I. ¹³¹In, ⁹⁰Y, and ¹⁸⁶Re. Conjugates of the antibody and cytotoxic agent are made using a variety of bifunctional protein-coupling agents such as N-succinimidyl-3-(2-pyridyldithiol) propionate (SPDP), iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutareldehyde), bis-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), bis-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as tolyene 2,6-diisocyanate), and bis-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). For example, a ricin immunotoxin can be prepared as described in Vitetta et al., Science, 238: 1098 (1987). Carbon-14-labeled 1-isothiocyanatobenzyl-3-methyldiethylene triaminepentaacetic acid (MX-DTPA) is an exemplary chelating agent for conjugation of radionucleotide to the antibody. See WO94/11026.

Conjugates of an antibody and one or more small molecule toxins, such as a calicheamicin, maytansinoids, a trichothene, and CC1065, and the derivatives of these toxins that have toxin activity, are also contemplated herein.

Maytansine and maytansinoids

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In one preferred embodiment, an anti-TAT antibody (full length or fragments) of the invention is conjugated to one or more maytansinoid molecules.

Maytansinoids are mitototic inhibitors which act by inhibiting tubulin polymerization. Maytansine was first isolated from the east African shrub *Maytenus serrata* (U.S. Patent No. 3,896,111). Subsequently, it was discovered that certain microbes also produce maytansinoids, such as maytansinol and C-3 maytansinol esters (U.S. Patent No. 4,151,042). Synthetic maytansinol and derivatives and analogues thereof are disclosed, for example, in U.S. Patent Nos. 4,137,230; 4,248,870; 4,256,746; 4,260,608; 4,265,814; 4,294,757; 4,307,016; 4,308,268; 4,308,269; 4,309,428; 4,313,946; 4,315,929; 4,317,821; 4,322,348; 4,331,598; 4,361,650; 4,364,866; 4,424,219; 4,450,254; 4,362,663; and 4,371,533, the disclosures of which are hereby expressly incorporated by reference.

Maytansinoid-antibody conjugates

In an attempt to improve their therapeutic index, maytansine and maytansinoids have been conjugated to antibodies specifically binding to tumor cell antigens. Immunoconjugates containing maytansinoids and their therapeutic use are disclosed, for example, in U.S. Patent Nos. 5,208,020, 5,416,064 and European Patent EP

0 425 235 B1, the disclosures of which are hereby expressly incorporated by reference. Liu et al., <u>Proc. Natl. Acad. Sci. USA</u> 93:8618-8623 (1996) described immunoconjugates comprising a maytansinoid designated DM1 linked to the monoclonal antibody C242 directed against human colorectal cancer. The conjugate was found to be highly cytotoxic towards cultured colon cancer cells, and showed antitumor activity in an *in vivo* tumor growth assay. Chari et al., <u>Cancer Research</u> 52:127-131 (1992) describe immunoconjugates in which a maytansinoid was conjugated via a disulfide linker to the murine antibody A7 binding to an antigen on human colon cancer cell lines, or to another murine monoclonal antibody TA.1 that binds the HER-2/neu oncogene. The cytotoxicity of the TA.1-maytansonoid conjugate was tested *in vitro* on the human breast cancer cell line SK-BR-3, which expresses 3 x 10⁵ HER-2 surface antigens per cell. The drug conjugate achieved a degree of cytotoxicity similar to the free maytansonid drug, which could be increased by increasing the number of maytansinoid molecules per antibody molecule. The A7-maytansinoid conjugate showed low systemic cytotoxicity in mice.

Anti-TAT polypeptide antibody-maytansinoid conjugates (immunoconjugates)

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Anti-TAT antibody-maytansinoid conjugates are prepared by chemically linking an anti-TAT antibody to a maytansinoid molecule without significantly diminishing the biological activity of either the antibody or the maytansinoid molecule. An average of 3-4 maytansinoid molecules conjugated per antibody molecule has shown efficacy in enhancing cytotoxicity of target cells without negatively affecting the function or solubility of the antibody, although even one molecule of toxin/antibody would be expected to enhance cytotoxicity over the use of naked antibody. Maytansinoids are well known in the art and can be synthesized by known techniques or isolated from natural sources. Suitable maytansinoids are disclosed, for example, in U.S. Patent No. 5,208,020 and in the other patents and nonpatent publications referred to hereinabove. Preferred maytansinoids are maytansinol and maytansinol analogues modified in the aromatic ring or at other positions of the maytansinol molecule, such as various maytansinol esters.

There are many linking groups known in the art for making antibody-maytansinoid conjugates, including, for example, those disclosed in U.S. Patent No. 5,208,020 or EP Patent 0 425 235 B1, and Chari et al., <u>Cancer Research</u> 52:127-131 (1992). The linking groups include disufide groups, thioether groups, acid labile groups, photolabile groups, peptidase labile groups, or esterase labile groups, as disclosed in the above-identified patents, disulfide and thioether groups being preferred.

Conjugates of the antibody and maytansinoid may be made using a variety of bifunctional protein coupling agents such as N-succinimidyl-3-(2-pyridyldithio) propionate (SPDP), succinimidyl-4-(N-maleimidomethyl) cyclohexane-1-carboxylate, iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutareldehyde), bis-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), bis-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as toluene 2,6-diisocyanate), and bis-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). Particularly preferred coupling agents include N-succinimidyl-3-(2-pyridyldithio) propionate (SPDP) (Carlsson et al. <u>Biochem. J.</u> 173:723-737 [1978]) and N-succinimidyl-4-(2-pyridylthio)pentanoate (SPP) to provide for a disulfide linkage.

The linker may be attached to the maytansinoid molecule at various positions, depending on the type of the link. For example, an ester linkage may be formed by reaction with a hydroxyl group using conventional coupling techniques. The reaction may occur at the C-3 position having a hydroxyl group, the C-14 position modified with hydroxymethyl, the C-15 position modified with a hydroxyl group, and the C-20 position having a hydroxyl group. In a preferred embodiment, the linkage is formed at the C-3 position of maytansinol or a maytansinol analogue.

Calicheamicin

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Another immunoconjugate of interest comprises an anti-TAT antibody conjugated to one or more calicheamicin molecules. The calicheamicin family of antibiotics are capable of producing double-stranded DNA breaks at sub-picomolar concentrations. For the preparation of conjugates of the calicheamicin family, see U.S. patents 5,712,374, 5,714,586, 5,739,116, 5,767,285, 5,770,701, 5,770,710, 5,773,001, 5,877,296 (all to American Cyanamid Company). Structural analogues of calicheamicin which may be used include, but are not limited to, γ_1^I , α_2^I , α_3^I , N-acetyl- γ_1^I , PSAG and θ_1^I (Hinman et al., Cancer Research 53:3336-3342 (1993), Lode et al., Cancer Research 58:2925-2928 (1998) and the aforementioned U.S. patents to American Cyanamid). Another anti-tumor drug that the antibody can be conjugated is QFA which is an antifolate. Both calicheamicin and QFA have intracellular sites of action and do not readily cross the plasma membrane. Therefore, cellular uptake of these agents through antibody mediated internalization greatly enhances their cytotoxic effects.

Other cytotoxic agents

Other antitumor agents that can be conjugated to the anti-TAT antibodies of the invention include BCNU, streptozoicin, vincristine and 5-fluorouracil, the family of agents known collectively LL-E33288 complex described in U.S. patents 5,053,394, 5,770,710, as well as esperamicins (U.S. patent 5,877,296).

Enzymatically active toxins and fragments thereof which can be used include diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, *Aleurites fordii* proteins, dianthin proteins, *Phytolaca americana* proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcin, crotin, sapaonaria officinalis inhibitor, gelonin, mitogellin, restrictocin, phenomycin, enomycin and the tricothecenes. See, for example, WO 93/21232 published October 28, 1993.

The present invention further contemplates an immunoconjugate formed between an antibody and a compound with nucleolytic activity (e.g., a ribonuclease or a DNA endonuclease such as a deoxyribonuclease; DNase).

For selective destruction of the tumor, the antibody may comprise a highly radioactive atom. A variety of radioactive isotopes are available for the production of radioconjugated anti-TAT antibodies. Examples include At²¹¹, I¹³¹, I¹²⁵, Y⁹⁰, Re¹⁸⁶, Re¹⁸⁸, Sm¹⁵³, Bi²¹², P³², Pb²¹² and radioactive isotopes of Lu. When the conjugate is used for diagnosis, it may comprise a radioactive atom for scintigraphic studies, for example tc^{99m} or I¹²³, or a spin label for nuclear magnetic resonance (NMR) imaging (also known as magnetic resonance

imaging, mri), such as iodine-123 again, iodine-131, indium-111, fluorine-19, carbon-13, nitrogen-15, oxygen-17, gadolinium, manganese or iron.

The radio- or other labels may be incorporated in the conjugate in known ways. For example, the peptide may be biosynthesized or may be synthesized by chemical amino acid synthesis using suitable amino acid precursors involving, for example, fluorine-19 in place of hydrogen. Labels such as tc^{99m} or I¹²³, .Re¹⁸⁶, Re¹⁸⁸ and In¹¹¹ can be attached via a cysteine residue in the peptide. Yttrium-90 can be attached via a lysine residue. The IODOGEN method (Fraker et al (1978) Biochem. Biophys. Res. Commun. 80: 49-57 can be used to incorporate iodine-123. "Monoclonal Antibodies in Immunoscintigraphy" (Chatal, CRC Press 1989) describes other methods in detail.

Conjugates of the antibody and cytotoxic agent may be made using a variety of bifunctional protein coupling agents such as N-succinimidyl-3-(2-pyridyldithio) propionate (SPDP), succinimidyl-4-(N-maleimidomethyl) cyclohexane-1-carboxylate, iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutareldehyde), bis-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), bis-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as tolyene 2,6-diisocyanate), and bis-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). For example, a ricin immunotoxin can be prepared as described in Vitetta et al., Science 238:1098 (1987). Carbon-14-labeled 1-isothiocyanatobenzyl-3-methyldiethylene triaminepentaacetic acid (MX-DTPA) is an exemplary chelating agent for conjugation of radionucleotide to the antibody. See WO94/11026. The linker may be a "cleavable linker" facilitating release of the cytotoxic drug in the cell. For example, an acid-labile linker, peptidase-sensitive linker, photolabile linker, dimethyl linker or disulfide-containing linker (Chari et al., Cancer Research 52:127-131 (1992); U.S. Patent No. 5,208,020) may be used.

Alternatively, a fusion protein comprising the anti-TAT antibody and cytotoxic agent may be made, e.g., by recombinant techniques or peptide synthesis. The length of DNA may comprise respective regions encoding the two portions of the conjugate either adjacent one another or separated by a region encoding a linker peptide which does not destroy the desired properties of the conjugate.

In yet another embodiment, the antibody may be conjugated to a "receptor" (such streptavidin) for utilization in tumor pre-targeting wherein the antibody-receptor conjugate is administered to the patient, followed by removal of unbound conjugate from the circulation using a clearing agent and then administration of a "ligand" (e.g., avidin) which is conjugated to a cytotoxic agent (e.g., a radionucleotide).

10. <u>Immunoliposomes</u>

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The anti-TAT antibodies disclosed herein may also be formulated as immunoliposomes. A "liposome" is a small vesicle composed of various types of lipids, phospholipids and/or surfactant which is useful for delivery of a drug to a mammal. The components of the liposome are commonly arranged in a bilayer formation, similar to the lipid arrangement of biological membranes. Liposomes containing the antibody are prepared by methods known in the art, such as described in Epstein et al., Proc. Natl. Acad. Sci. USA 82:3688 (1985); Hwang et al., Proc. Natl. Acad. Sci. USA 77:4030 (1980); U.S. Pat. Nos. 4,485,045 and 4,544,545;

and WO97/38731 published October 23, 1997. Liposomes with enhanced circulation time are disclosed in U.S. Patent No. 5,013,556.

Particularly useful liposomes can be generated by the reverse phase evaporation method with a lipid composition comprising phosphatidylcholine, cholesterol and PEG-derivatized phosphatidylethanolamine (PEG-PE). Liposomes are extruded through filters of defined pore size to yield liposomes with the desired diameter. Fab' fragments of the antibody of the present invention can be conjugated to the liposomes as described in Martin et al., J. Biol. Chem. 257:286-288 (1982) via a disulfide interchange reaction. A chemotherapeutic agent is optionally contained within the liposome. See Gabizon et al., J. National Cancer Inst. 81(19):1484 (1989).

B. TAT Binding Oligopeptides

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TAT binding oligopeptides of the present invention are oligopeptides that bind, preferably specifically, to a TAT polypeptide as described herein. TAT binding oligopeptides may be chemically synthesized using known oligopeptide synthesis methodology or may be prepared and purified using recombinant technology. TAT binding oligopeptides are usually at least about 5 amino acids in length, alternatively at least about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, or 100 amino acids in length or more, wherein such oligopeptides that are capable of binding, preferably specifically, to a TAT polypeptide as described herein. TAT binding oligopeptides may be identified without undue experimentation using well known techniques. In this regard, it is noted that techniques for screening oligopeptide libraries for oligopeptides that are capable of specifically binding to a polypeptide target are well known in the art (see, e.g., U.S. Patent Nos. 5,556,762, 5,750,373, 4,708,871, 4,833,092, 5,223,409, 5,403,484, 5,571,689, 5,663,143; PCT Publication Nos. WO 84/03506 and WO84/03564; Geysen et al., Proc. Natl. Acad. Sci. U.S.A., 81:3998-4002 (1984); Geysen et al., Proc. Natl. Acad. Sci. U.S.A., 82:178-182 (1985); Geysen et al., in Synthetic Peptides as Antigens, 130-149 (1986); Geysen et al., J. Immunol. Meth., 102:259-274 (1987); Schoofs et al., J. Immunol., 140:611-616 (1988), Cwirla, S. E. et al. (1990) Proc. Natl. Acad. Sci. USA, 87:6378; Lowman, H.B. et al. (1991) Biochemistry, 30:10832; Clackson, T. et al. (1991) Nature, 352: 624; Marks, J. D. et al. (1991), J. Mol. Biol., 222:581; Kang, A.S. et al. (1991) Proc. Natl. Acad. Sci. USA, 88:8363, and Smith, G. P. (1991) Current Opin. Biotechnol., 2:668).

In this regard, bacteriophage (phage) display is one well known technique which allows one to screen large oligopeptide libraries to identify member(s) of those libraries which are capable of specifically binding to a polypeptide target. Phage display is a technique by which variant polypeptides are displayed as fusion proteins to the coat protein on the surface of bacteriophage particles (Scott, J.K. and Smith, G. P. (1990) Science 249: 386). The utility of phage display lies in the fact that large libraries of selectively randomized protein variants (or randomly cloned cDNAs) can be rapidly and efficiently sorted for those sequences that bind to a target molecule with high affinity. Display of peptide (Cwirla, S. E. et al. (1990) Proc. Natl. Acad. Sci.

USA, 87:6378) or protein (Lowman, H.B. et al. (1991) Biochemistry, 30:10832; Clackson, T. et al. (1991) Nature, 352: 624; Marks, J. D. et al. (1991), J. Mol. Biol., 222:581; Kang, A.S. et al. (1991) Proc. Natl. Acad. Sci. USA, 88:8363) libraries on phage have been used for screening millions of polypeptides or oligopeptides for ones with specific binding properties (Smith, G. P. (1991) Current Opin. Biotechnol., 2:668). Sorting phage libraries of random mutants requires a strategy for constructing and propagating a large number of variants, a procedure for affinity purification using the target receptor, and a means of evaluating the results of binding enrichments. U.S. Patent Nos. 5,223,409, 5,403,484, 5,571,689, and 5,663,143.

Although most phage display methods have used filamentous phage, lambdoid phage display systems (WO 95/34683; U.S. 5,627,024), T4 phage display systems (Ren, Z-J. et al. (1998) Gene 215:439; Zhu, Z. (1997) CAN 33:534; Jiang, J. et al. (1997) can 128:44380; Ren, Z-J. et al. (1997) CAN 127:215644; Ren, Z-J. (1996) Protein Sci. 5:1833; Efimov, V. P. et al. (1995) Virus Genes 10:173) and T7 phage display systems (Smith, G. P. and Scott, J.K. (1993) Methods in Enzymology, 217, 228-257; U.S. 5,766,905) are also known.

Many other improvements and variations of the basic phage display concept have now been developed. These improvements enhance the ability of display systems to screen peptide libraries for binding to selected target molecules and to display functional proteins with the potential of screening these proteins for desired properties. Combinatorial reaction devices for phage display reactions have been developed (WO 98/14277) and phage display libraries have been used to analyze and control bimolecular interactions (WO 98/20169; WO 98/20159) and properties of constrained helical peptides (WO 98/20036). WO 97/35196 describes a method of isolating an affinity ligand in which a phage display library is contacted with one solution in which the ligand will bind to a target molecule and a second solution in which the affinity ligand will not bind to the target molecule, to selectively isolate binding ligands. WO 97/46251 describes a method of biopanning a random phage display library with an affinity purified antibody and then isolating binding phage, followed by a micropanning process using microplate wells to isolate high affinity binding phage. The use of Staphlylococcus aureus protein A as an affinity tag has also been reported (Li et al. (1998) Mol Biotech., 9:187). WO 97/47314 describes the use of substrate subtraction libraries to distinguish enzyme specificities using a combinatorial library which may be a phage display library. A method for selecting enzymes suitable for use in detergents using phage display is described in WO 97/09446. Additional methods of selecting specific binding proteins are described in U.S. Patent Nos. 5,498,538, 5,432,018, and WO 98/15833.

Methods of generating peptide libraries and screening these libraries are also disclosed in U.S. Patent Nos. 5,723,286, 5,432,018, 5,580,717, 5,427,908, 5,498,530, 5,770,434, 5,734,018, 5,698,426, 5,763,192, and 5,723,323.

C. TAT Binding Organic Molecules

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TAT binding organic molecules are organic molecules other than oligopeptides or antibodies as defined herein that bind, preferably specifically, to a TAT polypeptide as described herein. TAT binding organic molecules may be identified and chemically synthesized using known methodology (see, e.g., PCT Publication Nos. WO00/00823 and WO00/39585). TAT binding organic molecules are usually less than about 2000 daltons in size, alternatively less than about 1500, 750, 500, 250 or 200 daltons in size, wherein such organic molecules

that are capable of binding, preferably specifically, to a TAT polypeptide as described herein may be identified without undue experimentation using well known techniques. In this regard, it is noted that techniques for screening organic molecule libraries for molecules that are capable of binding to a polypeptide target are well known in the art (see, e.g., PCT Publication Nos. WO00/00823 and WO00/39585). TAT binding organic molecules may be, for example, aldehydes, ketones, oximes, hydrazones, semicarbazones, carbazides, primary amines, secondary amines, tertiary amines, N-substituted hydrazines, hydrazides, alcohols, ethers, thiols, thioethers, disulfides, carboxylic acids, esters, amides, ureas, carbamates, carbonates, ketals, thioketals, acetals, thioacetals, aryl halides, aryl sulfonates, alkyl halides, alkyl sulfonates, aromatic compounds, heterocyclic compounds, anilines, alkenes, alkynes, diols, amino alcohols, oxazolidines, oxazolines, thiazolidines, thiazolines, enamines, sulfonamides, epoxides, aziridines, isocyanates, sulfonyl chlorides, diazo compounds, acid chlorides, or the like.

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D. <u>Screening for Anti-TAT Antibodies, TAT Binding Oligopeptides and TAT Binding Organic</u> <u>Molecules With the Desired Properties</u>

Techniques for generating antibodies, oligopeptides and organic molecules that bind to TAT polypeptides have been described above. One may further select antibodies, oligopeptides or other organic molecules with certain biological characteristics, as desired.

The growth inhibitory effects of an anti-TAT antibody, oligopeptide or other organic molecule of the invention may be assessed by methods known in the art, e.g., using cells which express a TAT polypeptide either endogenously or following transfection with the TAT gene. For example, appropriate tumor cell lines and TAT-transfected cells may treated with an anti-TAT monoclonal antibody, oligopeptide or other organic molecule of the invention at various concentrations for a few days (e.g., 2-7) days and stained with crystal violet or MTT or analyzed by some other colorimetric assay. Another method of measuring proliferation would be by comparing 3H-thymidine uptake by the cells treated in the presence or absence an anti-TAT antibody, TAT binding oligopeptide or TAT binding organic molecule of the invention. After treatment, the cells are harvested and the amount of radioactivity incorporated into the DNA quantitated in a scintillation counter. Appropriate positive controls include treatment of a selected cell line with a growth inhibitory antibody known to inhibit growth of that cell line. Growth inhibition of tumor cells in vivo can be determined in various ways known in the art. Preferably, the tumor cell is one that overexpresses a TAT polypeptide. Preferably, the anti-TAT antibody, TAT binding oligopeptide or TAT binding organic molecule will inhibit cell proliferation of a TATexpressing tumor cell in vitro or in vivo by about 25-100% compared to the untreated tumor cell, more preferably, by about 30-100%, and even more preferably by about 50-100% or 70-100%, in one embodiment, at an antibody concentration of about 0.5 to 30 µg/ml. Growth inhibition can be measured at an antibody concentration of about 0.5 to 30 µg/ml or about 0.5 nM to 200 nM in cell culture, where the growth inhibition is determined 1-10 days after exposure of the tumor cells to the antibody. The antibody is growth inhibitory in vivo if administration of the anti-TAT antibody at about 1 µg/kg to about 100 mg/kg body weight results in reduction in tumor size or reduction of tumor cell proliferation within about 5 days to 3 months from the first administration of the antibody, preferably within about 5 to 30 days.

To select for an anti-TAT antibody, TAT binding oligopeptide or TAT binding organic molecule which induces cell death, loss of membrane integrity as indicated by, e.g., propidium iodide (PI), trypan blue or 7AAD uptake may be assessed relative to control. A PI uptake assay can be performed in the absence of complement and immune effector cells. TAT polypeptide-expressing tumor cells are incubated with medium alone or medium containing the appropriate anti-TAT antibody (e.g., at about 10 µg/ml), TAT binding oligopeptide or TAT binding organic molecule. The cells are incubated for a 3 day time period. Following each treatment, cells are washed and aliquoted into 35 mm strainer-capped 12 x 75 tubes (1ml per tube, 3 tubes per treatment group) for removal of cell clumps. Tubes then receive PI (10 µg/ml). Samples may be analyzed using a FACSCAN® flow cytometer and FACSCONVERT® CellQuest software (Becton Dickinson). Those anti-TAT antibodies, TAT binding oligopeptides or TAT binding organic molecules that induce statistically significant levels of cell death as determined by PI uptake may be selected as cell death-inducing anti-TAT antibodies, TAT binding organic molecules.

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To screen for antibodies, oligopeptides or other organic molecules which bind to an epitope on a TAT polypeptide bound by an antibody of interest, a routine cross-blocking assay such as that described in Antibodies, A Laboratory Manual, Cold Spring Harbor Laboratory, Ed Harlow and David Lane (1988), can be performed. This assay can be used to determine if a test antibody, oligopeptide or other organic molecule binds the same site or epitope as a known anti-TAT antibody. Alternatively, or additionally, epitope mapping can be performed by methods known in the art. For example, the antibody sequence can be mutagenized such as by alanine scanning, to identify contact residues. The mutant antibody is initially tested for binding with polyclonal antibody to ensure proper folding. In a different method, peptides corresponding to different regions of a TAT polypeptide can be used in competition assays with the test antibodies or with a test antibody and an antibody with a characterized or known epitope.

E. Antibody Dependent Enzyme Mediated Prodrug Therapy (ADEPT)

The antibodies of the present invention may also be used in ADEPT by conjugating the antibody to a prodrug-activating enzyme which converts a prodrug (e.g., a peptidyl chemotherapeutic agent, see WO81/01145) to an active anti-cancer drug. See, for example, WO 88/07378 and U.S. Patent No. 4,975,278.

The enzyme component of the immunoconjugate useful for ADEPT includes any enzyme capable of acting on a prodrug in such a way so as to covert it into its more active, cytotoxic form.

Enzymes that are useful in the method of this invention include, but are not limited to, alkaline phosphatase useful for converting phosphate-containing prodrugs into free drugs; arylsulfatase useful for converting sulfate-containing prodrugs into free drugs; cytosine deaminase useful for converting non-toxic 5-fluorocytosine into the anti-cancer drug, 5-fluorouracil; proteases, such as serratia protease, thermolysin, subtilisin, carboxypeptidases and cathepsins (such as cathepsins B and L), that are useful for converting peptide-containing prodrugs into free drugs; D-alanylcarboxypeptidases, useful for converting prodrugs that contain D-amino acid substituents; carbohydrate-cleaving enzymes such as β-galactosidase and neuraminidase useful for converting glycosylated prodrugs into free drugs; β-lactamase useful for converting drugs derivatized with β-lactams into free drugs; and penicillin amidases, such as penicillin V amidase or penicillin G amidase, useful

for converting drugs derivatized at their amine nitrogens with phenoxyacetyl or phenylacetyl groups, respectively, into free drugs. Alternatively, antibodies with enzymatic activity, also known in the art as "abzymes", can be used to convert the prodrugs of the invention into free active drugs (see, e.g., Massey, Nature 328:457-458 (1987)). Antibody-abzyme conjugates can be prepared as described herein for delivery of the abzyme to a tumor cell population.

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The enzymes of this invention can be covalently bound to the anti-TAT antibodies by techniques well known in the art such as the use of the heterobifunctional crosslinking reagents discussed above. Alternatively, fusion proteins comprising at least the antigen binding region of an antibody of the invention linked to at least a functionally active portion of an enzyme of the invention can be constructed using recombinant DNA techniques well known in the art (see, e.g., Neuberger et al., Nature 312:604-608 (1984).

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F. Full-Length TAT Polypeptides

The present invention also provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as TAT polypeptides. In particular, cDNAs (partial and full-length) encoding various TAT polypeptides have been identified and isolated, as disclosed in further detail in the Examples below.

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As disclosed in the Examples below, various cDNA clones have been deposited with the ATCC. The actual nucleotide sequences of those clones can readily be determined by the skilled artisan by sequencing of the deposited clone using routine methods in the art. The predicted amino acid sequence can be determined from the nucleotide sequence using routine skill. For the TAT polypeptides and encoding nucleic acids described herein, in some cases, Applicants have identified what is believed to be the reading frame best identifiable with the sequence information available at the time.

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G. Anti-TAT Antibody and TAT Polypeptide Variants

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In addition to the anti-TAT antibodies and full-length native sequence TAT polypeptides described herein, it is contemplated that anti-TAT antibody and TAT polypeptide variants can be prepared. Anti-TAT antibody and TAT polypeptide variants can be prepared by introducing appropriate nucleotide changes into the encoding DNA, and/or by synthesis of the desired antibody or polypeptide. Those skilled in the art will appreciate that amino acid changes may alter post-translational processes of the anti-TAT antibody or TAT polypeptide, such as changing the number or position of glycosylation sites or altering the membrane anchoring characteristics.

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Variations in the anti-TAT antibodies and TAT polypeptides described herein, can be made, for example, using any of the techniques and guidelines for conservative and non-conservative mutations set forth, for instance, in U.S. Patent No. 5,364,934. Variations may be a substitution, deletion or insertion of one or more codons encoding the antibody or polypeptide that results in a change in the amino acid sequence as compared with the native sequence antibody or polypeptide. Optionally the variation is by substitution of at least one amino acid with any other amino acid in one or more of the domains of the anti-TAT antibody or TAT polypeptide. Guidance in determining which amino acid residue may be inserted, substituted or deleted without adversely affecting the desired activity may be found by comparing the sequence of the anti-TAT antibody or

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TAT polypeptide with that of homologous known protein molecules and minimizing the number of amino acid sequence changes made in regions of high homology. Amino acid substitutions can be the result of replacing one amino acid with another amino acid having similar structural and/or chemical properties, such as the replacement of a leucine with a serine, i.e., conservative amino acid replacements. Insertions or deletions may optionally be in the range of about 1 to 5 amino acids. The variation allowed may be determined by systematically making insertions, deletions or substitutions of amino acids in the sequence and testing the resulting variants for activity exhibited by the full-length or mature native sequence.

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Anti-TAT antibody and TAT polypeptide fragments are provided herein. Such fragments may be truncated at the N-terminus or C-terminus, or may lack internal residues, for example, when compared with a full length native antibody or protein. Certain fragments lack amino acid residues that are not essential for a desired biological activity of the anti-TAT antibody or TAT polypeptide.

Anti-TAT antibody and TAT polypeptide fragments may be prepared by any of a number of conventional techniques. Desired peptide fragments may be chemically synthesized. An alternative approach involves generating antibody or polypeptide fragments by enzymatic digestion, e.g., by treating the protein with an enzyme known to cleave proteins at sites defined by particular amino acid residues, or by digesting the DNA with suitable restriction enzymes and isolating the desired fragment. Yet another suitable technique involves isolating and amplifying a DNA fragment encoding a desired antibody or polypeptide fragment, by polymerase chain reaction (PCR). Oligonucleotides that define the desired termini of the DNA fragment are employed at the 5' and 3' primers in the PCR. Preferably, anti-TAT antibody and TAT polypeptide fragments share at least one biological and/or immunological activity with the native anti-TAT antibody or TAT polypeptide disclosed herein.

In particular embodiments, conservative substitutions of interest are shown in Table 6 under the heading of preferred substitutions. If such substitutions result in a change in biological activity, then more substantial changes, denominated exemplary substitutions in Table 6, or as further described below in reference to amino acid classes, are introduced and the products screened.

Table 6

	Original	Exemplary	Preferred
	Residue	Substitutions	Substitutions
	Ala (A)	val; leu; ile	val
5	Arg (R)	lys; gln; asn	lys
	Asn (N)	gln; his; lys; arg	gln
	Asp (D)	glu	glu
	Cys (C)	ser	ser
	Gln (Q)	asn	asn
10	Glu (E)	asp	asp
	Gly (G)	pro; ala	ala
	His (H)	asn; gln; lys; arg	arg
	Ile (I)	leu; val; met; ala; phe;	
		norleucine	leu
15	Leu (L)	norleucine; ile; val;	
		met; ala; phe	ile
	Lys (K)	arg; gln; asn	arg
	Met (M)	leu; phe; ile	leu
	Phe (F)	leu; val; ile; ala; tyr	leu
20	Pro (P)	ala	ala
	Ser (S)	thr	thr
	Thr (T)	ser	ser
	Trp (W)	tyr; phe	tyr
	Tyr (Y)	trp; phe; thr; ser	phe
25	Val (V)	ile; leu; met; phe;	•
		ala; norleucine	leu

Substantial modifications in function or immunological identity of the anti-TAT antibody or TAT polypeptide are accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues are divided into groups based on common side-chain properties:

- (1) hydrophobic: norleucine, met, ala, val, leu, ile;
- (2) neutral hydrophilic: cys, ser, thr;
- 35 (3) acidic: asp, glu;

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- (4) basic: asn, gln, his, lys, arg;
- (5) residues that influence chain orientation: gly, pro; and
- (6) aromatic: trp, tyr, phe.

Non-conservative substitutions will entail exchanging a member of one of these classes for another class. Such substituted residues also may be introduced into the conservative substitution sites or, more preferably, into the remaining (non-conserved) sites.

The variations can be made using methods known in the art such as oligonucleotide-mediated (site-directed) mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis [Carter et al. <u>Nucl. Acids Res.</u>, 13:4331 (1986); Zoller et al., <u>Nucl. Acids Res.</u>, 10:6487 (1987)], cassette mutagenesis [Wells et al., <u>Gene</u>, 34:315 (1985)], restriction selection mutagenesis [Wells et al., <u>Philos. Trans. R. Soc. London SerA</u>, 317:415 (1986)] or other known techniques can be performed on the cloned DNA to produce the anti-TAT antibody or TAT polypeptide variant DNA.

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Scanning amino acid analysis can also be employed to identify one or more amino acids along a contiguous sequence. Among the preferred scanning amino acids are relatively small, neutral amino acids. Such amino acids include alanine, glycine, serine, and cysteine. Alanine is typically a preferred scanning amino acid among this group because it eliminates the side-chain beyond the beta-carbon and is less likely to alter the main-chain conformation of the variant [Cunningham and Wells, Science, 244:1081-1085 (1989)]. Alanine is also typically preferred because it is the most common amino acid. Further, it is frequently found in both buried and exposed positions [Creighton, The Proteins, (W.H. Freeman & Co., N.Y.); Chothia, J. Mol. Biol., 150:1 (1976)]. If alanine substitution does not yield adequate amounts of variant, an isoteric amino acid can be used.

Any cysteine residue not involved in maintaining the proper conformation of the anti-TAT antibody or TAT polypeptide also may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant crosslinking. Conversely, cysteine bond(s) may be added to the anti-TAT antibody or TAT polypeptide to improve its stability (particularly where the antibody is an antibody fragment such as an Fv fragment).

A particularly preferred type of substitutional variant involves substituting one or more hypervariable region residues of a parent antibody (e.g., a humanized or human antibody). Generally, the resulting variant(s) selected for further development will have improved biological properties relative to the parent antibody from which they are generated. A convenient way for generating such substitutional variants involves affinity maturation using phage display. Briefly, several hypervariable region sites (e.g., 6-7 sites) are mutated to generate all possible amino substitutions at each site. The antibody variants thus generated are displayed in a monovalent fashion from filamentous phage particles as fusions to the gene III product of M13 packaged within each particle. The phage-displayed variants are then screened for their biological activity (e.g., binding affinity) as herein disclosed. In order to identify candidate hypervariable region sites for modification, alanine scanning mutagenesis can be performed to identify hypervariable region residues contributing significantly to antigen binding. Alternatively, or additionally, it may be beneficial to analyze a crystal structure of the antigen-antibody complex to identify contact points between the antibody and human TAT polypeptide. Such contact residues and neighboring residues are candidates for substitution according to the techniques elaborated herein. Once such variants are generated, the panel of variants is subjected to screening as described herein and antibodies with superior properties in one or more relevant assays may be selected for further development.

Nucleic acid molecules encoding amino acid sequence variants of the anti-TAT antibody are prepared by a variety of methods known in the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally occurring amino acid sequence variants) or preparation by oligonucleotide-

mediated (or site-directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of the anti-TAT antibody.

H. Modifications of Anti-TAT Antibodies and TAT Polypeptides

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Covalent modifications of anti-TAT antibodies and TAT polypeptides are included within the scope of this invention. One type of covalent modification includes reacting targeted amino acid residues of an anti-TAT antibody or TAT polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C- terminal residues of the anti-TAT antibody or TAT polypeptide. Derivatization with bifunctional agents is useful, for instance, for crosslinking anti-TAT antibody or TAT polypeptide to a water-insoluble support matrix or surface for use in the method for purifying anti-TAT antibodies, and vice-versa. Commonly used crosslinking agents include, e.g., 1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde, N-hydroxysuccinimide esters, for example, esters with 4-azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis(succinimidylpropionate), bifunctional maleimides such as bis-N-maleimido-1,8-octane and agents such as methyl-3-[(p-azidophenyl)dithio]propioimidate.

Other modifications include deamidation of glutaminyl and asparaginyl residues to the corresponding glutamyl and aspartyl residues, respectively, hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α -amino groups of lysine, arginine, and histidine side chains [T.E. Creighton, <u>Proteins: Structure and Molecular Properties</u>, W.H. Freeman & Co., San Francisco, pp. 79-86 (1983)], acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

Another type of covalent modification of the anti-TAT antibody or TAT polypeptide included within the scope of this invention comprises altering the native glycosylation pattern of the antibody or polypeptide. "Altering the native glycosylation pattern" is intended for purposes herein to mean deleting one or more carbohydrate moieties found in native sequence anti-TAT antibody or TAT polypeptide (either by removing the underlying glycosylation site or by deleting the glycosylation by chemical and/or enzymatic means), and/or adding one or more glycosylation sites that are not present in the native sequence anti-TAT antibody or TAT polypeptide. In addition, the phrase includes qualitative changes in the glycosylation of the native proteins, involving a change in the nature and proportions of the various carbohydrate moieties present.

Glycosylation of antibodies and other polypeptides is typically either N-linked or O-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tripeptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one of the sugars N-aceylgalactosamine, galactose, or xylose to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

Addition of glycosylation sites to the anti-TAT antibody or TAT polypeptide is conveniently accomplished by altering the amino acid sequence such that it contains one or more of the above-described tripeptide sequences (for N-linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to the sequence of the original anti-TAT antibody or TAT polypeptide (for O-linked glycosylation sites). The anti-TAT antibody or TAT polypeptide amino acid sequence may optionally be altered through changes at the DNA level, particularly by mutating the DNA encoding the anti-TAT antibody or TAT polypeptide at preselected bases such that codons are generated that will translate into the desired amino acids.

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Another means of increasing the number of carbohydrate moieties on the anti-TAT antibody or TAT polypeptide is by chemical or enzymatic coupling of glycosides to the polypeptide. Such methods are described in the art, e.g., in WO 87/05330 published 11 September 1987, and in Aplin and Wriston, <u>CRC Crit. Rev. Biochem.</u>, pp. 259-306 (1981).

Removal of carbohydrate moieties present on the anti-TAT antibody or TAT polypeptide may be accomplished chemically or enzymatically or by mutational substitution of codons encoding for amino acid residues that serve as targets for glycosylation. Chemical deglycosylation techniques are known in the art and described, for instance, by Hakimuddin, et al., Arch. Biochem. Biophys., 259:52 (1987) and by Edge et al., Anal. Biochem., 118:131 (1981). Enzymatic cleavage of carbohydrate moieties on polypeptides can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura et al., Meth. Enzymol., 138:350 (1987).

Another type of covalent modification of anti-TAT antibody or TAT polypeptide comprises linking the antibody or polypeptide to one of a variety of nonproteinaceous polymers, e.g., polyethylene glycol (PEG), polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337. The antibody or polypeptide also may be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization (for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacylate) microcapsules, respectively), in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nanoparticles and nanocapsules), or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences, 16th edition, Oslo, A., Ed., (1980).

The anti-TAT antibody or TAT polypeptide of the present invention may also be modified in a way to form chimeric molecules comprising an anti-TAT antibody or TAT polypeptide fused to another, heterologous polypeptide or amino acid sequence.

In one embodiment, such a chimeric molecule comprises a fusion of the anti-TAT antibody or TAT polypeptide with a tag polypeptide which provides an epitope to which an anti-tag antibody can selectively bind. The epitope tag is generally placed at the amino- or carboxyl- terminus of the anti-TAT antibody or TAT polypeptide. The presence of such epitope-tagged forms of the anti-TAT antibody or TAT polypeptide can be detected using an antibody against the tag polypeptide. Also, provision of the epitope tag enables the anti-TAT

antibody or TAT polypeptide to be readily purified by affinity purification using an anti-tag antibody or another type of affinity matrix that binds to the epitope tag. Various tag polypeptides and their respective antibodies are well known in the art. Examples include poly-histidine (poly-his) or poly-histidine-glycine (poly-his-gly) tags; the flu HA tag polypeptide and its antibody 12CA5 [Field et al., Mol. Cell. Biol., 8:2159-2165 (1988)]; the c-myc tag and the 8F9, 3C7, 6E10, G4, B7 and 9E10 antibodies thereto [Evan et al., Molecular and Cellular Biology, 5:3610-3616 (1985)]; and the Herpes Simplex virus glycoprotein D (gD) tag and its antibody [Paborsky et al., Protein Engineering, 3(6):547-553 (1990)]. Other tag polypeptides include the Flag-peptide [Hopp et al., BioTechnology, 6:1204-1210 (1988)]; the KT3 epitope peptide [Martin et al., Science, 255:192-194 (1992)]; an α-tubulin epitope peptide [Skinner et al., J. Biol. Chem., 266:15163-15166 (1991)]; and the T7 gene 10 protein peptide tag [Lutz-Freyermuth et al., Proc. Natl. Acad. Sci. USA, 87:6393-6397 (1990)].

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In an alternative embodiment, the chimeric molecule may comprise a fusion of the anti-TAT antibody or TAT polypeptide with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the chimeric molecule (also referred to as an "immunoadhesin"), such a fusion could be to the Fc region of an IgG molecule. The Ig fusions preferably include the substitution of a soluble (transmembrane domain deleted or inactivated) form of an anti-TAT antibody or TAT polypeptide in place of at least one variable region within an Ig molecule. In a particularly preferred embodiment, the immunoglobulin fusion includes the hinge, CH₂ and CH₃, or the hinge, CH₁, CH₂ and CH₃ regions of an IgG1 molecule. For the production of immunoglobulin fusions see also US Patent No. 5,428,130 issued June 27, 1995.

I. Preparation of Anti-TAT Antibodies and TAT Polypeptides

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The description below relates primarily to production of anti-TAT antibodies and TAT polypeptides by culturing cells transformed or transfected with a vector containing anti-TAT antibody- and TAT polypeptide-encoding nucleic acid. It is, of course, contemplated that alternative methods, which are well known in the art, may be employed to prepare anti-TAT antibodies and TAT polypeptides. For instance, the appropriate amino acid sequence, or portions thereof, may be produced by direct peptide synthesis using solid-phase techniques [see, e.g., Stewart et al., Solid-Phase Peptide Synthesis, W.H. Freeman Co., San Francisco, CA (1969); Merrifield, J. Am. Chem. Soc., 85:2149-2154 (1963)]. In vitro protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be accomplished, for instance, using an Applied Biosystems Peptide Synthesizer (Foster City, CA) using manufacturer's instructions. Various portions of the anti-TAT antibody or TAT polypeptide may be chemically synthesized separately and combined using chemical or enzymatic methods to produce the desired anti-TAT antibody or TAT polypeptide.

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1. Isolation of DNA Encoding Anti-TAT Antibody or TAT Polypeptide

DNA encoding anti-TAT antibody or TAT polypeptide may be obtained from a cDNA library prepared from tissue believed to possess the anti-TAT antibody or TAT polypeptide mRNA and to express it at a detectable level. Accordingly, human anti-TAT antibody or TAT polypeptide DNA can be conveniently obtained from a cDNA library prepared from human tissue. The anti-TAT antibody- or TAT polypeptide-encoding gene may also be obtained from a genomic library or by known synthetic procedures (e.g., automated

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nucleic acid synthesis).

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Libraries can be screened with probes (such as oligonucleotides of at least about 20-80 bases) designed to identify the gene of interest or the protein encoded by it. Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the gene encoding anti-TAT antibody or TAT polypeptide is to use PCR methodology [Sambrook et al., supra; Dieffenbach et al., PCR Primer: A Laboratory Manual (Cold Spring Harbor Laboratory Press, 1995)].

Techniques for screening a cDNA library are well known in the art. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized. The oligonucleotide is preferably labeled such that it can be detected upon hybridization to DNA in the library being screened. Methods of labeling are well known in the art, and include the use of radiolabels like ³²P-labeled ATP, biotinylation or enzyme labeling. Hybridization conditions, including moderate stringency and high stringency, are provided in Sambrook et al., supra.

Sequences identified in such library screening methods can be compared and aligned to other known sequences deposited and available in public databases such as GenBank or other private sequence databases. Sequence identity (at either the amino acid or nucleotide level) within defined regions of the molecule or across the full-length sequence can be determined using methods known in the art and as described herein.

Nucleic acid having protein coding sequence may be obtained by screening selected cDNA or genomic libraries using the deduced amino acid sequence disclosed herein for the first time, and, if necessary, using conventional primer extension procedures as described in Sambrook et al., <u>supra</u>, to detect precursors and processing intermediates of mRNA that may not have been reverse-transcribed into cDNA.

2. Selection and Transformation of Host Cells

Host cells are transfected or transformed with expression or cloning vectors described herein for anti-TAT antibody or TAT polypeptide production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in <u>Mammalian Cell Biotechnology</u>: a Practical Approach, M. Butler, ed. (IRL Press, 1991) and Sambrook et al., suppra.

Methods of eukaryotic cell transfection and prokaryotic cell transformation are known to the ordinarily skilled artisan, for example, CaCl₂, CaPO₄, liposome-mediated and electroporation. Depending on the host cell used, transformation is performed using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described in Sambrook et al., <u>supra</u>, or electroporation is generally used for prokaryotes. Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., <u>Gene</u>, <u>23</u>:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, <u>Virology</u>,

52:456-457 (1978) can be employed. General aspects of mammalian cell host system transfections have been described in U.S. Patent No. 4,399,216. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, e.g., polybrene, polyornithine, may also be used. For various techniques for transforming mammalian cells, see Keown et al., Methods in Enzymology, 185:527-537 (1990) and Mansour et al., Nature, 336:348-352 (1988).

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Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as E. coli. Various E. coli strains are publicly available, such as E. coli K12 strain MM294 (ATCC 31,446); E. coli X1776 (ATCC 31,537); E. coli strain W3110 (ATCC 27,325) and K5 772 (ATCC 53,635). Other suitable prokaryotic host cells include Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g., B. licheniformis 41P disclosed in DD 266,710 published 12 April 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. These examples are illustrative rather than limiting. Strain W3110 is one particularly preferred host or parent host because it is a common host strain for recombinant DNA product fermentations. Preferably, the host cell secretes minimal amounts of proteolytic enzymes. For example, strain W3110 may be modified to effect a genetic mutation in the genes encoding proteins endogenous to the host, with examples of such hosts including E. coli W3110 strain 1A2, which has the complete genotype tonA; E. coli W3110 strain 9E4, which has the complete genotype tonA ptr3; E. coli W3110 strain 27C7 (ATCC 55,244), which has the complete genotype tonA ptr3 phoA E15 (argF-lac)169 degP ompT kan'; E. coli W3110 strain 37D6, which has the complete genotype tonA ptr3 phoA E15 (argF-lac)169 degP ompT rbs7 ilvG karl; E. coli W3110 strain 40B4, which is strain 37D6 with a non-kanamycin resistant degP deletion mutation; and an E. coli strain having mutant periplasmic protease disclosed in U.S. Patent No. 4,946,783 issued 7 August 1990. Alternatively, in vitro methods of cloning, e.g., PCR or other nucleic acid polymerase reactions, are suitable.

Full length antibody, antibody fragments, and antibody fusion proteins can be produced in bacteria, in particular when glycosylation and Fc effector function are not needed, such as when the therapeutic antibody is conjugated to a cytotoxic agent (e.g., a toxin) and the immunoconjugate by itself shows effectiveness in tumor cell destruction. Full length antibodies have greater half life in circulation. Production in E. coli is faster and more cost efficient. For expression of antibody fragments and polypeptides in bacteria, see, e.g., U.S. 5,648,237 (Carter et. al.), U.S. 5,789,199 (Joly et al.), and U.S. 5,840,523 (Simmons et al.) which describes translation initiation regio (TIR) and signal sequences for optimizing expression and secretion, these patents incorporated herein by reference. After expression, the antibody is isolated from the E. coli cell paste in a soluble fraction and can be purified through, e.g., a protein A or G column depending on the isotype. Final purification can be carried out similar to the process for purifying antibody expressed e.g., in CHO cells.

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for anti-TAT antibody- or TAT polypeptide-encoding vectors. Saccharomyces cerevisiae is a commonly used lower eukaryotic host microorganism. Others include Schizosaccharomyces pombe (Beach and Nurse, Nature, 290: 140 [1981]; EP 139,383 published 2 May 1985); Kluyveromyces hosts (U.S. Patent No. 4,943,529; Fleer et al., Bio/Technology, 9:968-975 (1991)) such as, e.g., K. lactis (MW98-8C, CBS683, CBS4574; Louvencourt et al., J. Bacteriol., 154(2):737-742 [1983]), K. fragilis (ATCC 12,424), K. bulgaricus (ATCC 16,045), K. wickeramii (ATCC 24,178), K. waltii (ATCC 56,500), K. drosophilarum (ATCC 36,906; Van den Berg et al., Bio/Technology, 8:135 (1990)), K. thermotolerans, and K. marxianus; yarrowia (EP 402,226); Pichia pastoris (EP 183,070; Sreekrishna et al., J. Basic Microbiol., 28:265-278 [1988]); Candida; Trichoderma reesia (EP 244,234); Neurospora crassa (Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 [1979]); Schwanniomyces such as Schwanniomyces occidentalis (EP 394,538 published 31 October 1990); and filamentous fungi such as, e.g., Neurospora, Penicillium, Tolypocladium (WO 91/00357 published 10 January 1991), and Aspergillus hosts such as A. nidulans (Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 [1983]; Tilburn et al., Gene, 26:205-221 [1983]; Yelton et al., Proc. Natl. Acad. Sci. USA, 81: 1470-1474 [1984]) and A. niger (Kelly and Hynes, EMBO J., 4:475-479 [1985]). Methylotropic yeasts are suitable herein and include, but are not limited to, yeast capable of growth on methanol selected from the genera consisting of Hansenula, Candida, Kloeckera, Pichia, Saccharomyces, Torulopsis, and Rhodotorula. A list of specific species that are exemplary of this class of yeasts may be found in C. Anthony, The Biochemistry of Methylotrophs, 269 (1982).

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Suitable host cells for the expression of glycosylated anti-TAT antibody or TAT polypeptide are derived from multicellular organisms. Examples of invertebrate cells include insect cells such as Drosophila S2 and Spodoptera Sf9, as well as plant cells, such as cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as Spodoptera frugiperda (caterpillar), Aedes aegypti (mosquito), Aedes albopictus (mosquito), Drosophila melanogaster (fruitfly), and Bombyx mori have been identified. A variety of viral strains for transfection are publicly available, e.g., the L-1 variant of Autographa californica NPV and the Bm-5 strain of Bombyx mori NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of Spodoptera frugiperda cells.

However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., <u>J. Gen Virol.</u> 36:59 (1977)); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub et al., <u>Proc. Natl. Acad. Sci. USA</u>77:4216 (1980)); mouse sertoli cells (TM4, Mather, <u>Biol. Reprod.</u> 23:243-251 (1980)); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2,

HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci. 383:44-68 (1982)); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors for anti-TAT antibody or TAT polypeptide production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

3. Selection and Use of a Replicable Vector

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The nucleic acid (e.g., cDNA or genomic DNA) encoding anti-TAT antibody or TAT polypeptide may be inserted into a replicable vector for cloning (amplification of the DNA) or for expression. Various vectors are publicly available. The vector may, for example, be in the form of a plasmid, cosmid, viral particle, or phage. The appropriate nucleic acid sequence may be inserted into the vector by a variety of procedures. In general, DNA is inserted into an appropriate restriction endonuclease site(s) using techniques known in the art. Vector components generally include, but are not limited to, one or more of a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence. Construction of suitable vectors containing one or more of these components employs standard ligation techniques which are known to the skilled artisan.

The TAT may be produced recombinantly not only directly, but also as a fusion polypeptide with a heterologous polypeptide, which may be a signal sequence or other polypeptide having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the vector, or it may be a part of the anti-TAT antibody- or TAT polypeptide-encoding DNA that is inserted into the vector. The signal sequence may be a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, lpp, or heat-stable enterotoxin II leaders. For yeast secretion the signal sequence may be, e.g., the yeast invertase leader, alpha factor leader (including Saccharomyces and Kluyveromyces α -factor leaders, the latter described in U.S. Patent No. 5,010,182), or acid phosphatase leader, the C. albicans glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression, mammalian signal sequences may be used to direct secretion of the protein, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders.

Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells.

Expression and cloning vectors will typically contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for *Bacilli*.

An example of suitable selectable markers for mammalian cells are those that enable the identification

of cells competent to take up the anti-TAT antibody- or TAT polypeptide-encoding nucleic acid, such as DHFR or thymidine kinase. An appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and propagated as described by Urlaub et al., <u>Proc. Natl. Acad. Sci. USA</u>, 77:4216 (1980). A suitable selection gene for use in yeast is the *trp1* gene present in the yeast plasmid YRp7 [Stinchcomb et al., <u>Nature</u>, 282:39 (1979); Kingsman et al., <u>Gene</u>, 7:141 (1979); Tschemper et al., <u>Gene</u>, 10:157 (1980)]. The *trp1* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076 or PEP4-1 [Jones, <u>Genetics</u>, 85:12 (1977)].

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Expression and cloning vectors usually contain a promoter operably linked to the anti-TAT antibodyor TAT polypeptide-encoding nucleic acid sequence to direct mRNA synthesis. Promoters recognized by a
variety of potential host cells are well known. Promoters suitable for use with prokaryotic hosts include theslactamase and lactose promoter systems [Chang et al., Nature, 275:615 (1978); Goeddel et al., Nature, 281:544
(1979)], alkaline phosphatase, a tryptophan (trp) promoter system [Goeddel, Nucleic Acids Res., 8:4057 (1980);
EP 36,776], and hybrid promoters such as the tac promoter [deBoer et al., Proc. Natl. Acad. Sci. USA, 80:2125 (1983)]. Promoters for use in bacterial systems also will contain a Shine-Dalgarno (S.D.) sequence operably
linked to the DNA encoding anti-TAT antibody or TAT polypeptide.

Examples of suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase [Hitzeman et al., J. Biol. Chem., 255:2073 (1980)] or other glycolytic enzymes [Hess et al., J. Adv. Enzyme Reg., 7:149 (1968); Holland, Biochemistry, 17:4900 (1978)], such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in EP 73,657.

Anti-TAT antibody or TAT polypeptide transcription from vectors in mammalian host cells is controlled, for example, by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g., the actin promoter or an immunoglobulin promoter, and from heat-shock promoters, provided such promoters are compatible with the host cell systems.

Transcription of a DNA encoding the anti-TAT antibody or TAT polypeptide by higher eukaryotes may be increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the

late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. The enhancer may be spliced into the vector at a position 5' or 3' to the anti-TAT antibody or TAT polypeptide coding sequence, but is preferably located at a site 5' from the promoter.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3', untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding anti-TAT antibody or TAT polypeptide.

Still other methods, vectors, and host cells suitable for adaptation to the synthesis of anti-TAT antibody or TAT polypeptide in recombinant vertebrate cell culture are described in Gething et al., Nature, 293:620-625 (1981); Mantei et al., Nature, 281:40-46 (1979); EP 117,060; and EP 117,058.

4. Culturing the Host Cells

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The host cells used to produce the anti-TAT antibody or TAT polypeptide of this invention may be cultured in a variety of media. Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ((MEM), (Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ((DMEM), Sigma) are suitable for culturing the host cells. In addition, any of the media described in Ham et alMeth. Enz. 58:44 (1979), Barnes et al., Anal. Biochem.102:255 (1980), U.S. Pat. Nos. 4,767,704; 4,657,866; 4,927,762; 4,560,655; or 5,122,469; WO 90/03430; WO 87/00195; or U.S. Patent Re. 30,985 may be used as culture media for the host cells. Any of these media may be supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleotides (such as adenosine and thymidine), antibiotics (such as GENTAMYCINTM drug), trace elements (defined as inorganic compounds usually present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

5. Detecting Gene Amplification/Expression

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Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA [Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 (1980)], dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of cells or tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. Antibodies useful for immunohistochemical staining and/or assay of sample fluids may be either monoclonal or polyclonal, and may be prepared in any mammal. Conveniently, the antibodies may be prepared against a native sequence TAT polypeptide or against a synthetic peptide based on the DNA sequences provided herein or against exogenous sequence fused to TAT DNA and encoding a specific antibody epitope.

6. Purification of Anti-TAT Antibody and TAT Polypeptide

Forms of anti-TAT antibody and TAT polypeptide may be recovered from culture medium or from host cell lysates. If membrane-bound, it can be released from the membrane using a suitable detergent solution (e.g. Triton-X 100) or by enzymatic cleavage. Cells employed in expression of anti-TAT antibody and TAT polypeptide can be disrupted by various physical or chemical means, such as freeze-thaw cycling, sonication, mechanical disruption, or cell lysing agents.

It may be desired to purify anti-TAT antibody and TAT polypeptide from recombinant cell proteins or polypeptides. The following procedures are exemplary of suitable purification procedures: by fractionation on an ion-exchange column; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation-exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel filtration using, for example, Sephadex G-75; protein A Sepharose columns to remove contaminants such as IgG; and metal chelating columns to bind epitope-tagged forms of the anti-TAT antibody and TAT polypeptide. Various methods of protein purification may be employed and such methods are known in the art and described for example in Deutscher, Methods in Enzymology, 182 (1990); Scopes, Protein Purification: Principles and Practice, Springer-Verlag, New York (1982). The purification step(s) selected will depend, for example, on the nature of the production process used and the particular anti-TAT antibody or TAT polypeptide produced.

When using recombinant techniques, the antibody can be produced intracellularly, in the periplasmic space, or directly secreted into the medium. If the antibody is produced intracellularly, as a first step, the particulate debris, either host cells or lysed fragments, are removed, for example, by centrifugation or ultrafiltration. Carter et al., <u>Bio/Technology</u> 10:163-167 (1992) describe a procedure for isolating antibodies which are secreted to the periplasmic space of *E. coli*. Briefly, cell paste is thawed in the presence of sodium

acetate (pH 3.5), EDTA, and phenylmethylsulfonylfluoride (PMSF) over about 30 min. Cell debris can be removed by centrifugation. Where the antibody is secreted into the medium, supernatants from such expression systems are generally first concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit. A protease inhibitor such as PMSF may be included in any of the foregoing steps to inhibit proteolysis and antibiotics may be included to prevent the growth of adventitious contaminants.

The antibody composition prepared from the cells can be purified using, for example, hydroxylapatite chromatography, gel electrophoresis, dialysis, and affinity chromatography, with affinity chromatography being the preferred purification technique. The suitability of protein A as an affinity ligand depends on the species and isotype of any immunoglobulin Fc domain that is present in the antibody. Protein A can be used to purify antibodies that are based on human $\gamma 1$, $\gamma 2$ or $\gamma 4$ heavy chains (Lindmark et al., J. Immunol. Meth. 62:1-13 (1983)). Protein G is recommended for all mouse isotypes and for human y3 (Guss et al., EMBO J. 5:15671575 (1986)). The matrix to which the affinity ligand is attached is most often agarose, but other Mechanically stable matrices such as controlled pore glass or matrices are available. poly(styrenedivinyl)benzene allow for faster flow rates and shorter processing times than can be achieved with agarose. Where the antibody comprises a C₁3 domain, the Bakerbond ABX™resin (J. T. Baker, Phillipsburg, NJ) is useful for purification. Other techniques for protein purification such as fractionation on an ion-exchange column, ethanol precipitation, Reverse Phase HPLC, chromatography on silica, chromatography on heparin SEPHAROSE™ chromatography on an anion or cation exchange resin (such as a polyaspartic acid column), chromatofocusing, SDS-PAGE, and ammonium sulfate precipitation are also available depending on the antibody to be recovered.

Following any preliminary purification step(s), the mixture comprising the antibody of interest and contaminants may be subjected to low pH hydrophobic interaction chromatography using an elution buffer at a pH between about 2.5-4.5, preferably performed at low salt concentrations (e.g., from about 0-0.25M salt).

J. Pharmaceutical Formulations

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Therapeutic formulations of the anti-TAT antibodies, TAT binding oligopeptides, TAT binding organic molecules and/or TAT polypeptides used in accordance with the present invention are prepared for storage by mixing the antibody, polypeptide, oligopeptide or organic molecule having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers (Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. (1980)), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as acetate, Tris, phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine,

histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; tonicifiers such as trehalose and sodium chloride; sugars such as sucrose, mannitol, trehalose or sorbitol; surfactant such as polysorbate; salt-forming counter-ions such as sodium; metal complexes (e.g., Zn-protein complexes); and/or non-ionic surfactants such as TWEEN®, PLURONICS® or polyethylene glycol (PEG). The antibody preferably comprises the antibody at a concentration of between 5-200 mg/ml, preferably between 10-100 mg/ml.

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The formulations herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. For example, in addition to an anti-TAT antibody, TAT binding oligopeptide, or TAT binding organic molecule, it may be desirable to include in the one formulation, an additional antibody, e.g., a second anti-TAT antibody which binds a different epitope on the TAT polypeptide, or an antibody to some other target such as a growth factor that affects the growth of the particular cancer. Alternatively, or additionally, the composition may further comprise a chemotherapeutic agent, cytotoxic agent, cytokine, growth inhibitory agent, anti-hormonal agent, and/or cardioprotectant. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences, 16th edition, Osol, A. Ed. (1980).

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semi-permeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, e.g., films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919), copolymers of L-glutamic acid and γ ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT® (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid.

The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

K. <u>Diagnosis and Treatment with Anti-TAT Antibodies, TAT Binding Oligopeptides and TAT</u>
 Binding Organic Molecules

To determine TAT expression in the cancer, various diagnostic assays are available. In one embodiment, TAT polypeptide overexpression may be analyzed by immunohistochemistry (IHC). Parrafin embedded tissue sections from a tumor biopsy may be subjected to the IHC assay and accorded a TAT protein staining intensity criteria as follows:

Score 0 - no staining is observed or membrane staining is observed in less than 10% of tumor cells.

Score 1+ - a faint/barely perceptible membrane staining is detected in more than 10% of the tumor cells. The cells are only stained in part of their membrane.

Score 2+ - a weak to moderate complete membrane staining is observed in more than 10% of the tumor cells.

Score 3+ - a moderate to strong complete membrane staining is observed in more than 10% of the tumor cells.

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Those tumors with 0 or 1+ scores for TAT polypeptide expression may be characterized as not overexpressing TAT, whereas those tumors with 2+ or 3+ scores may be characterized as overexpressing TAT.

Alternatively, or additionally, FISH assays such as the INFORM® (sold by Ventana, Arizona) or PATHVISION® (Vysis, Illinois) may be carried out on formalin-fixed, paraffin-embedded tumor tissue to determine the extent (if any) of TAT overexpression in the tumor.

TAT overexpression or amplification may be evaluated using an *in vivo* diagnostic assay, e.g., by administering a molecule (such as an antibody, oligopeptide or organic molecule) which binds the molecule to be detected and is tagged with a detectable label (e.g., a radioactive isotope or a fluorescent label) and externally scanning the patient for localization of the label.

As described above, the anti-TAT antibodies, oligopeptides and organic molecules of the invention have various non-therapeutic applications. The anti-TAT antibodies, oligopeptides and organic molecules of the present invention can be useful for diagnosis and staging of TAT polypeptide-expressing cancers (e.g., in radioimaging). The antibodies, oligopeptides and organic molecules are also useful for purification or immunoprecipitation of TAT polypeptide from cells, for detection and quantitation of TAT polypeptide in vitro, e.g., in an ELISA or a Western blot, to kill and eliminate TAT-expressing cells from a population of mixed cells as a step in the purification of other cells.

Currently, depending on the stage of the cancer, cancer treatment involves one or a combination of the following therapies: surgery to remove the cancerous tissue, radiation therapy, and chemotherapy. Anti-TAT antibody, oligopeptide or organic molecule therapy may be especially desirable in elderly patients who do not tolerate the toxicity and side effects of chemotherapy well and in metastatic disease where radiation therapy has limited usefulness. The tumor targeting anti-TAT antibodies, oligopeptides and organic molecules of the invention are useful to alleviate TAT-expressing cancers upon initial diagnosis of the disease or during relapse. For therapeutic applications, the anti-TAT antibody, oligopeptide or organic molecule can be used alone, or in combination therapy with, e.g., hormones, antiangiogens, or radiolabelled compounds, or with surgery, cryotherapy, and/or radiotherapy. Anti-TAT antibody, oligopeptide or organic molecule treatment can be administered in conjunction with other forms of conventional therapy, either consecutively with, pre- or post-conventional therapy. Chemotherapeutic drugs such as TAXOTERE® (docetaxel), TAXOL® (palictaxel), estramustine and mitoxantrone are used in treating cancer, in particular, in good risk patients. In the present method of the invention for treating or alleviating cancer, the cancer patient can be administered anti-TAT

antibody, oligopeptide or organic molecule in conjuction with treatment with the one or more of the preceding chemotherapeutic agents. In particular, combination therapy with palictaxel and modified derivatives (see, e.g., EP0600517) is contemplated. The anti-TAT antibody, oligopeptide or organic molecule will be administered with a therapeutically effective dose of the chemotherapeutic agent. In another embodiment, the anti-TAT antibody, oligopeptide or organic molecule is administered in conjunction with chemotherapy to enhance the activity and efficacy of the chemotherapeutic agent, e.g., paclitaxel. The Physicians' Desk Reference (PDR) discloses dosages of these agents that have been used in treatment of various cancers. The dosing regimen and dosages of these aforementioned chemotherapeutic drugs that are therapeutically effective will depend on the particular cancer being treated, the extent of the disease and other factors familiar to the physician of skill in the art and can be determined by the physician.

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In one particular embodiment, a conjugate comprising an anti-TAT antibody, oligopeptide or organic molecule conjugated with a cytotoxic agent is administered to the patient. Preferably, the immunoconjugate bound to the TAT protein is internalized by the cell, resulting in increased therapeutic efficacy of the immunoconjugate in killing the cancer cell to which it binds. In a preferred embodiment, the cytotoxic agent targets or interferes with the nucleic acid in the cancer cell. Examples of such cytotoxic agents are described above and include maytansinoids, calicheamicins, ribonucleases and DNA endonucleases.

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The anti-TAT antibodies, oligopeptides, organic molecules or toxin conjugates thereof are administered to a human patient, in accord with known methods, such as intravenous administration, e.g.,, as a bolus or by continuous infusion over a period of time, by intramuscular, intraperitoneal, intracerobrospinal, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. Intravenous or subcutaneous administration of the antibody, oligopeptide or organic molecule is preferred.

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Other therapeutic regimens may be combined with the administration of the anti-TAT antibody, oligopeptide or organic molecule. The combined administration includes co-administration, using separate formulations or a single pharmaceutical formulation, and consecutive administration in either order, wherein preferably there is a time period while both (or all) active agents simultaneously exert their biological activities. Preferably such combined therapy results in a synergistic therapeutic effect.

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It may also be desirable to combine administration of the anti-TAT antibody or antibodies, oligopeptides or organic molecules, with administration of an antibody directed against another tumor antigen associated with the particular cancer.

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In another embodiment, the therapeutic treatment methods of the present invention involves the combined administration of an anti-TAT antibody (or antibodies), oligopeptides or organic molecules and one or more chemotherapeutic agents or growth inhibitory agents, including co-administration of cocktails of different chemotherapeutic agents. Chemotherapeutic agents include estramustine phosphate, prednimustine, cisplatin, 5-fluorouracil, melphalan, cyclophosphamide, hydroxyurea and hydroxyureataxanes (such as paclitaxel and doxetaxel) and/or anthracycline antibiotics. Preparation and dosing schedules for such chemotherapeutic agents may be used according to manufacturers' instructions or as determined empirically by the skilled practitioner. Preparation and dosing schedules for such chemotherapy are also described in Chemotherapy

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Service Ed., M.C. Perry, Williams & Wilkins, Baltimore, MD (1992).

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The antibody, oligopeptide or organic molecule may be combined with an anti-hormonal compound; e.g., an anti-estrogen compound such as tamoxifen; an anti-progesterone such as onapristone (see, EP 616 812); or an anti-androgen such as flutamide, in dosages known for such molecules. Where the cancer to be treated is androgen independent cancer, the patient may previously have been subjected to anti-androgen therapy and, after the cancer becomes androgen independent, the anti-TAT antibody, oligopeptide or organic molecule (and optionally other agents as described herein) may be administered to the patient.

Sometimes, it may be beneficial to also co-administer a cardioprotectant (to prevent or reduce myocardial dysfunction associated with the therapy) or one or more cytokines to the patient. In addition to the above therapeutic regimes, the patient may be subjected to surgical removal of cancer cells and/or radiation therapy, before, simultaneously with, or post antibody, oligopeptide or organic molecule therapy. Suitable dosages for any of the above co-administered agents are those presently used and may be lowered due to the combined action (synergy) of the agent and anti-TAT antibody, oligopeptide or organic molecule.

For the prevention or treatment of disease, the dosage and mode of administration will be chosen by the physician according to known criteria. The appropriate dosage of antibody, oligopeptide or organic molecule will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether the antibody, oligopeptide or organic molecule is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the antibody, oligopeptide or organic molecule, and the discretion of the attending physician. The antibody, oligopeptide or organic molecule is suitably administered to the patient at one time or over a series of treatments. Preferably, the antibody, oligopeptide or organic molecule is administered by intravenous infusion or by subcutaneous injections. Depending on the type and severity of the disease, about 1 μ g/kg to about 50 mg/kg body weight (e.g., about 0.1-15mg/kg/dose) of antibody can be an initial candidate dosage for administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. A dosing regimen can comprise administering an initial loading dose of about 4 mg/kg, followed by a weekly maintenance dose of about 2 mg/kg of the anti-TAT antibody. However, other dosage regimens may be useful. A typical daily dosage might range from about 1 µg/kg to 100 mg/kg or more, depending on the factors mentioned above. For repeated administrations over several days or longer, depending on the condition, the treatment is sustained until a desired suppression of disease symptoms occurs. The progress of this therapy can be readily monitored by conventional methods and assays and based on criteria known to the physician or other persons of skill in the art.

Aside from administration of the antibody protein to the patient, the present application contemplates administration of the antibody by gene therapy. Such administration of nucleic acid encoding the antibody is encompassed by the expression "administering a therapeutically effective amount of an antibody". See, for example, WO96/07321 published March 14, 1996 concerning the use of gene therapy to generate intracellular antibodies.

There are two major approaches to getting the nucleic acid (optionally contained in a vector) into the patient's cells; in vivo and ex vivo. For in vivo delivery the nucleic acid is injected directly into the patient,

usually at the site where the antibody is required. For ex vivo treatment, the patient's cells are removed, the nucleic acid is introduced into these isolated cells and the modified cells are administered to the patient either directly or, for example, encapsulated within porous membranes which are implanted into the patient (see, e.g., U.S. Patent Nos. 4,892,538 and 5,283,187). There are a variety of techniques available for introducing nucleic acids into viable cells. The techniques vary depending upon whether the nucleic acid is transferred into cultured cells in vitro, or in vivo in the cells of the intended host. Techniques suitable for the transfer of nucleic acid into mammalian cells in vitro include the use of liposomes, electroporation, microinjection, cell fusion, DEAE-dextran, the calcium phosphate precipitation method, etc. A commonly used vector for ex vivo delivery of the gene is a retroviral vector.

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The currently preferred *in vivo* nucleic acid transfer techniques include transfection with viral vectors (such as adenovirus, Herpes simplex I virus, or adeno-associated virus) and lipid-based systems (useful lipids for lipid-mediated transfer of the gene are DOTMA, DOPE and DC-Chol, for example). For review of the currently known gene marking and gene therapy protocols see Anderson et al., <u>Science</u> 256:808-813 (1992). See also WO 93/25673 and the references cited therein.

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The anti-TAT antibodies of the invention can be in the different forms encompassed by the definition of "antibody" herein. Thus, the antibodies include full length or intact antibody, antibody fragments, native sequence antibody or amino acid variants, humanized, chimeric or fusion antibodies, immunoconjugates, and functional fragments thereof. In fusion antibodies an antibody sequence is fused to a heterologous polypeptide sequence. The antibodies can be modified in the Fc region to provide desired effector functions. As discussed in more detail in the sections herein, with the appropriate Fc regions, the naked antibody bound on the cell surface can induce cytotoxicity, e.g., via antibody-dependent cellular cytotoxicity (ADCC) or by recruiting complement in complement dependent cytotoxicity, or some other mechanism. Alternatively, where it is desirable to eliminate or reduce effector function, so as to minimize side effects or therapeutic complications, certain other Fc regions may be used.

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In one embodiment, the antibody competes for binding or bind substantially to, the same epitope as the antibodies of the invention. Antibodies having the biological characteristics of the present anti-TAT antibodies of the invention are also contemplated, specifically including the *in vivo* tumor targeting and any cell proliferation inhibition or cytotoxic characteristics.

Methods of producing the above antibodies are described in detail herein.

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The present anti-TAT antibodies, oligopeptides and organic molecules are useful for treating a TAT-expressing cancer or alleviating one or more symptoms of the cancer in a mammal. Such a cancer includes prostate cancer, cancer of the urinary tract, lung cancer, breast cancer, colon cancer and ovarian cancer, more specifically, prostate adenocarcinoma, renal cell carcinomas, colorectal adenocarcinomas, lung adenocarcinomas, lung squamous cell carcinomas, and pleural mesothelioma. The cancers encompass metastatic cancers of any of the preceding. The antibody, oligopeptide or organic molecule is able to bind to at least a portion of the cancer cells that express TAT polypeptide in the mammal. In a preferred embodiment, the

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antibody, oligopeptide or organic molecule is effective to destroy or kill TAT-expressing tumor cells or inhibit

the growth of such tumor cells, in vitro or in vivo, upon binding to TAT polypeptide on the cell. Such an antibody includes a naked anti-TAT antibody (not conjugated to any agent). Naked antibodies that have cytotoxic or cell growth inhibition properties can be further harnessed with a cytotoxic agent to render them even more potent in tumor cell destruction. Cytotoxic properties can be conferred to an anti-TAT antibody by, e.g., conjugating the antibody with a cytotoxic agent, to form an immunoconjugate as described herein. The cytotoxic agent or a growth inhibitory agent is preferably a small molecule. Toxins such as calicheamicin or a maytansinoid and analogs or derivatives thereof, are preferable.

The invention provides a composition comprising an anti-TAT antibody, oligopeptide or organic molecule of the invention, and a carrier. For the purposes of treating cancer, compositions can be administered to the patient in need of such treatment, wherein the composition can comprise one or more anti-TAT antibodies present as an immunoconjugate or as the naked antibody. In a further embodiment, the compositions can comprise these antibodies, oligopeptides or organic molecules in combination with other therapeutic agents such as cytotoxic or growth inhibitory agents, including chemotherapeutic agents. The invention also provides formulations comprising an anti-TAT antibody, oligopeptide or organic molecule of the invention, and a carrier. In one embodiment, the formulation is a therapeutic formulation comprising a pharmaceutically acceptable carrier.

Another aspect of the invention is isolated nucleic acids encoding the anti-TAT antibodies. Nucleic acids encoding both the H and L chains and especially the hypervariable region residues, chains which encode the native sequence antibody as well as variants, modifications and humanized versions of the antibody, are encompassed.

The invention also provides methods useful for treating a TAT polypeptide-expressing cancer or alleviating one or more symptoms of the cancer in a mammal, comprising administering a therapeutically effective amount of an anti-TAT antibody, oligopeptide or organic molecule to the mammal. The antibody, oligopeptide or organic molecule therapeutic compositions can be administered short term (acute) or chronic, or intermittent as directed by physician. Also provided are methods of inhibiting the growth of, and killing a TAT polypeptide-expressing cell.

The invention also provides kits and articles of manufacture comprising at least one anti-TAT antibody, oligopeptide or organic molecule. Kits containing anti-TAT antibodies, oligopeptides or organic molecules find use, e.g., for TAT cell killing assays, for purification or immunoprecipitation of TAT polypeptide from cells. For example, for isolation and purification of TAT, the kit can contain an anti-TAT antibody, oligopeptide or organic molecule coupled to beads (e.g., sepharose beads). Kits can be provided which contain the antibodies, oligopeptides or organic molecules for detection and quantitation of TAT in vitro, e.g., in an ELISA or a Western blot. Such antibody, oligopeptide or organic molecule useful for detection may be provided with a label such as a fluorescent or radiolabel.

L. Articles of Manufacture and Kits

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Another embodiment of the invention is an article of manufacture containing materials useful for the treatment of anti-TAT expressing cancer. The article of manufacture comprises a container and a label or

package insert on or associated with the container. Suitable containers include, for example, bottles, vials, syringes, etc. The containers may be formed from a variety of materials such as glass or plastic. The container holds a composition which is effective for treating the cancer condition and may have a sterile access port (for example the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection needle). At least one active agent in the composition is an anti-TAT antibody, oligopeptide or organic molecule of the invention. The label or package insert indicates that the composition is used for treating cancer. The label or package insert will further comprise instructions for administering the antibody, oligopeptide or organic molecule composition to the cancer patient. Additionally, the article of manufacture may further comprise a second container comprising a pharmaceutically-acceptable buffer, such as bacteriostatic water for injection (BWFI), phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, and syringes.

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Kits are also provided that are useful for various purposes, e.g., for TAT-expressing cell killing assays, for purification or immunoprecipitation of TAT polypeptide from cells. For isolation and purification of TAT polypeptide, the kit can contain an anti-TAT antibody, oligopeptide or organic molecule coupled to beads (e.g., sepharose beads). Kits can be provided which contain the antibodies, oligopeptides or organic molecules for detection and quantitation of TAT polypeptide *in vitro*, e.g., in an ELISA or a Western blot. As with the article of manufacture, the kit comprises a container and a label or package insert on or associated with the container. The container holds a composition comprising at least one anti-TAT antibody, oligopeptide or organic molecule of the invention. Additional containers may be included that contain, e.g., diluents and buffers, control antibodies. The label or package insert may provide a description of the composition as well as instructions for the intended in vitro or diagnostic use.

M. <u>Uses for TAT Polypeptides and TAT-Polypeptide Encoding Nucleic Acids</u>

Nucleotide sequences (or their complement) encoding TAT polypeptides have various applications in the art of molecular biology, including uses as hybridization probes, in chromosome and gene mapping and in the generation of anti-sense RNA and DNA probes. TAT-encoding nucleic acid will also be useful for the preparation of TAT polypeptides by the recombinant techniques described herein, wherein those TAT polypeptides may find use, for example, in the preparation of anti-TAT antibodies as described herein.

The full-length native sequence TAT gene, or portions thereof, may be used as hybridization probes for a cDNA library to isolate the full-length TAT cDNA or to isolate still other cDNAs (for instance, those encoding naturally-occurring variants of TAT or TAT from other species) which have a desired sequence identity to the native TAT sequence disclosed herein. Optionally, the length of the probes will be about 20 to about 50 bases. The hybridization probes may be derived from at least partially novel regions of the full length native nucleotide sequence wherein those regions may be determined without undue experimentation or from genomic sequences including promoters, enhancer elements and introns of native sequence TAT. By way of example, a screening method will comprise isolating the coding region of the TAT gene using the known DNA sequence to synthesize a selected probe of about 40 bases. Hybridization probes may be labeled by a variety

of labels, including radionucleotides such as ³²P or ³⁵S, or enzymatic labels such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems. Labeled probes having a sequence complementary to that of the TAT gene of the present invention can be used to screen libraries of human cDNA, genomic DNA or mRNA to determine which members of such libraries the probe hybridizes to. Hybridization techniques are described in further detail in the Examples below. Any EST sequences disclosed in the present application may similarly be employed as probes, using the methods disclosed herein.

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Other useful fragments of the TAT-encoding nucleic acids include antisense or sense oligonucleotides comprising a singe-stranded nucleic acid sequence (either RNA or DNA) capable of binding to target TAT mRNA (sense) or TAT DNA (antisense) sequences. Antisense or sense oligonucleotides, according to the present invention, comprise a fragment of the coding region of TAT DNA. Such a fragment generally comprises at least about 14 nucleotides, preferably from about 14 to 30 nucleotides. The ability to derive an antisense or a sense oligonucleotide, based upon a cDNA sequence encoding a given protein is described in, for example, Stein and Cohen (Cancer Res. 48:2659, 1988) and van der Krol et al. (BioTechniques 6:958, 1988).

Binding of antisense or sense oligonucleotides to target nucleic acid sequences results in the formation of duplexes that block transcription or translation of the target sequence by one of several means, including enhanced degradation of the duplexes, premature termination of transcription or translation, or by other means. Such methods are encompassed by the present invention. The antisense oligonucleotides thus may be used to block expression of TAT proteins, wherein those TAT proteins may play a role in the induction of cancer in mammals. Antisense or sense oligonucleotides further comprise oligonucleotides having modified sugarphosphodiester backbones (or other sugar linkages, such as those described in WO 91/06629) and wherein such sugar linkages are resistant to endogenous nucleases. Such oligonucleotides with resistant sugar linkages are stable *in vivo* (i.e., capable of resisting enzymatic degradation) but retain sequence specificity to be able to bind to target nucleotide sequences.

Preferred intragenic sites for antisense binding include the region incorporating the translation initiation/start codon (5'-AUG/5'-ATG) or termination/stop codon (5'-UAA, 5'-UAG and 5-UGA/5'-TAA, 5'-TAG and 5'-TGA) of the open reading frame (ORF) of the gene. These regions refer to a portion of the mRNA or gene that encompasses from about 25 to about 50 contiguous nucleotides in either direction (i.e., 5' or 3') from a translation initiation or termination codon. Other preferred regions for antisense binding include: introns; exons; intron-exon junctions; the open reading frame (ORF) or "coding region," which is the region between the translation initiation codon and the translation termination codon; the 5' cap of an mRNA which comprises an N7-methylated guanosine residue joined to the 5'-most residue of the mRNA via a 5'-5' triphosphate linkage and includes 5' cap structure itself as well as the first 50 nucleotides adjacent to the cap; the 5' untranslated region (5'UTR), the portion of an mRNA in the 5' direction from the translation initiation codon, and thus including nucleotides between the 5' cap site and the translation initiation codon of an mRNA or corresponding nucleotides between the translation termination codon, and thus including nucleotides between the translation termination codon and 3' end of an mRNA or corresponding nucleotides on the gene.

Specific examples of preferred antisense compounds useful for inhibiting expression of TAT proteins include oligonucleotides containing modified backbones or non-natural internucleoside linkages. Oligonucleotides having modified backbones include those that retain a phosphorus atom in the backbone and those that do not have a phosphorus atom in the backbone. For the purposes of this specification, and as sometimes referenced in the art, modified oligonucleotides that do not have a phosphorus atom in their internucleoside backbone can also be considered to be oligonucleosides. Preferred modified oligonucleotide backbones include, for example, phosphorothioates, chiral phosphorothioates, phosphorodithioates, phosphotriesters, aminoalkylphosphotri-esters, methyl and other alkyl phosphonates including 3'-alkylene phosphonates, 5'-alkylene phosphonates and chiral phosphonates, phosphinates, phosphoramidates including 3'-amino phosphoramidate and aminoalkylphosphoramidates, thionophosphoramidates, thionoalkylphosphonates, thionoalkylphosphotriesters, selenophosphates and borano-phosphates having normal 3'-5' linkages, 2'-5' linked analogs of these, and those having inverted polarity wherein one or more internucleotide linkages is a 3' to 3', 5' to 5' or 2' to 2' linkage. Preferred oligonucleotides having inverted polarity comprise a single 3' to 3' linkage at the 3'-most internucleotide linkage i.e. a single inverted nucleoside residue which may be abasic (the nucleobase is missing or has a hydroxyl group in place thereof). Various salts, mixed salts and free acid forms are also included. Representative United States patents that teach the preparation of phosphorus-containing linkages include, but are not limited to, U.S. Pat. Nos.: 3,687,808; 4,469,863; 4,476,301; 5,023,243; 5,177,196; 5,188,897; 5,264,423; 5,276,019; 5,278,302; 5,286,717; 5,321,131; 5,399,676; 5,405,939; 5,453,496; 5,455,233; 5,466,677; 5,476,925; 5,519,126; 5,536,821; 5,541,306; 5,550,111; 5,563,253; 5,571,799; 5,587,361; 5,194,599; 5,565,555; 5,527,899; 5,721,218; 5,672,697 and 5,625,050, each of which is herein incorporated by reference.

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Preferred modified oligonucleotide backbones that do not include a phosphorus atom therein have backbones that are formed by short chain alkyl or cycloalkyl internucleoside linkages, mixed heteroatom and alkyl or cycloalkyl internucleoside linkages, or one or more short chain heteroatomic or heterocyclic internucleoside linkages. These include those having morpholino linkages (formed in part from the sugar portion of a nucleoside); siloxane backbones; sulfide, sulfoxide and sulfone backbones; formacetyl and thioformacetyl backbones; methylene formacetyl and thioformacetyl backbones; riboacetyl backbones; alkene containing backbones; sulfamate backbones; methyleneimino and methylenehydrazino backbones; sulfonate and sulfonamide backbones; amide backbones; and others having mixed N, O, S and CH.sub.2 component parts. Representative United States patents that teach the preparation of such oligonucleosides include, but are not limited to,. U.S. Pat. Nos.: 5,034,506; 5,166,315; 5,185,444; 5,214,134; 5,216,141; 5,235,033; 5,264,562; 5,264,564; 5,405,938; 5,434,257; 5,466,677; 5,470,967; 5,489,677; 5,541,307; 5,561,225; 5,596,086; 5,602,240; 5,610,289; 5,602,240; 5,608,046; 5,610,289; 5,618,704; 5,623,070; 5,663,312; 5,633,360; 5,677,437; 5,792,608; 5,646,269 and 5,677,439, each of which is herein incorporated by reference.

In other preferred antisense oligonucleotides, both the sugar and the internucleoside linkage, i.e., the backbone, of the nucleotide units are replaced with novel groups. The base units are maintained for hybridization with an appropriate nucleic acid target compound. One such oligomeric compound, an oligonucleotide mimetic

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that has been shown to have excellent hybridization properties, is referred to as a peptide nucleic acid (PNA). In PNA compounds, the sugar-backbone of an oligonucleotide is replaced with an amide containing backbone, in particular an aminoethylglycine backbone. The nucleobases are retained and are bound directly or indirectly to aza nitrogen atoms of the amide portion of the backbone. Representative United States patents that teach the preparation of PNA compounds include, but are not limited to, U.S. Pat. Nos.: 5,539,082; 5,714,331; and 5,719,262, each of which is herein incorporated by reference. Further teaching of PNA compounds can be found in Nielsen et al., Science, 1991, 254, 1497-1500.

Preferred antisense oligonucleotides incorporate phosphorothioate backbones and/or heteroatom backbones, and in particular -CH₂-NH-O-CH₂-, -CH₂-N(CH₃)-O-CH₂- [known as a methylene (methylimino) or MMI backbone], -CH2-O-N(CH3)-CH2-, -CH2-N(CH3)-N(CH3)-CH2- and -O-N(CH3)-CH2- [wherein the native phosphodiester backbone is represented as -O-P-O-CH₂-] described in the above referenced U.S. Pat. No. 5,489,677, and the amide backbones of the above referenced U.S. Pat. No. 5,602,240. Also preferred are antisense oligonucleotides having morpholino backbone structures of the above-referenced U.S. Pat. No. 5,034,506.

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Modified oligonucleotides may also contain one or more substituted sugar moieties. Preferred oligonucleotides comprise one of the following at the 2' position: OH; F; O-alkyl, S-alkyl, or N-alkyl; Oalkenyl, S-alkeynyl, or N-alkenyl; O-alkynyl, S-alkynyl or N-alkynyl; or O-alkyl-O-alkyl, wherein the alkyl, alkenyl and alkynyl may be substituted or unsubstituted C $_{1}$ to C_{10} alkyl or C_{2} to C_{10} alkenyl and alkynyl. Particularly preferred are O[(CH₂)_nO]_mCH₃, O(CH₂)_nOCH₃, O(CH₂)_nNH₂, O(CH₂)_nCH₃, O(CH₂)_nONH₂, and O(CH₂)_nON[(CH₂)_nCH₃)]₂, where n and m are from 1 to about 10. Other preferred antisense oligonucleotides comprise one of the following at the 2' position: C1 to C10 lower alkyl, substituted lower alkyl, alkenyl, alkynyl, alkaryl, aralkyl, O-alkaryl or O-aralkyl, SH, SCH3, OCN, Cl, Br, CN, CF3, OCF3, SOCH3, SO2 CH3, ONO2, NO2, N3, NH2, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalkylamino, substituted silyl, an RNA cleaving group, a reporter group, an intercalator, a group for improving the pharmacokinetic properties of an oligonucleotide, or a group for improving the pharmacodynamic properties of an oligonucleotide, and other substituents having similar properties. A preferred modification includes 2'-methoxyethoxy (2'-O-CH2CH2OCH3, also known as 2'-O-(2-methoxyethyl) or 2'-MOE) (Martin et al., Helv. Chim. Acta, 1995, 78, 486-504) i.e., an alkoxyalkoxy group. A further preferred modification includes 2'-dimethylaminooxyethoxy, i.e., a O(CH 2)2ON(CH3)2 group, also known as 2'-DMAOE, as described in examples hereinbelow, and 2'-dimethylaminoethoxyethoxy (also known in the art as

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2'-O-dimethylaminoethoxyethyl or 2'-DMAEOE), i.e., 2'-O-CH₂-O-CH₂-N(CH₂).

A further prefered modification includes Locked Nucleic Acids (LNAs) in which the 2'-hydroxyl group is linked to the 3' or 4' carbon atom of the sugar ring thereby forming a bicyclic sugar moiety. The linkage is preferably a methelyne (-CH2-)n group bridging the 2' oxygen atom and the 4' carbon atom wherein n is 1 or 2. LNAs and preparation thereof are described in WO 98/39352 and WO 99/14226.

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Other preferred modifications include 2'-methoxy (2'-O-CH₃), 2'-aminopropoxy (2'-OCH₂CH₂CH₂

NH₂), 2'-allyl (2'-CH₂-CH=CH₂), 2'-O-allyl (2'-O-CH₂-CH=CH₂) and 2'-fluoro (2'-F). The 2'-modification may be in the arabino (up) position or ribo (down) position. A preferred 2'-arabino modification is 2'-F. Similar modifications may also be made at other positions on the oligonucleotide, particularly the 3' position of the sugar on the 3' terminal nucleotide or in 2'-5' linked oligonucleotides and the 5' position of 5' terminal nucleotide. Oligonucleotides may also have sugar mimetics such as cyclobutyl moieties in place of the pentofuranosyl sugar. Representative United States patents that teach the preparation of such modified sugar structures include, but are not limited to, U.S. Pat. Nos.: 4,981,957; 5,118,800; 5,319,080; 5,359,044; 5,393,878; 5,446,137; 5,466,786; 5,514,785; 5,519,134; 5,567,811; 5,576,427; 5,591,722; 5,597,909; 5,610,300; 5,627,053; 5,639,873; 5,646,265; 5,658,873; 5,670,633; 5,792,747; and 5,700,920, each of which is herein incorporated by reference in its entirety.

Oligonucleotides may also include nucleobase (often referred to in the art simply as "base")

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modifications or substitutions. As used herein, "unmodified" or "natural" nucleobases include the purine bases adenine (A) and guanine (G), and the pyrimidine bases thymine (T), cytosine (C) and uracil (U). Modified nucleobases include other synthetic and natural nucleobases such as 5-methylcytosine (5-me-C), 5-hydroxymethyl cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl (-C=C-CH₃ or -CH₂-C=CH) uracil and cytosine and other alkynyl derivatives of pyrimidine bases, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 2-F-adenine, 2-amino-adenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine and 3-deazaguanine and 3-deazaadenine. Further modified nucleobases include tricyclic pyrimidines such as phenoxazine cytidine(1H-pyrimido[5,4-b][1,4]benzoxazin-2(3H)-one), phenothiazine cytidine (1H-pyrimido[5,4-b][1,4]benzothiazin-2(3H)-one), G-clamps such as a substituted phenoxazine cytidine 9-(2-aminoethoxy)-H-pyrimido[5,4-b][1,4]benzoxazin-2(3H)-one), carbazole cytidine (2H-pyrimido[4,5-b]indol-2-one), pyridoindole cytidine (H-pyrido[3',2':4,5]pyrrolo[2,3-d]pyrimidin-2-one). Modified nucleobases may also include those in which the purine or pyrimidine base is replaced with other heterocycles, for example 7-deaza-adenine, 7-deazaguanosine, 2-aminopyridine and 2-pyridone. Further nucleobases include those disclosed in U.S. Pat. No. 3,687,808, those disclosed in The Concise Encyclopedia Of Polymer Science And Engineering, pages 858-859, Kroschwitz, J. I., ed. John Wiley & Sons, 1990, and those disclosed by Englisch et al., Angewandte Chemie, International Edition, 1991, 30, 613. Certain of these nucleobases are particularly useful for increasing the binding affinity of the oligomeric compounds of the invention. These include 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted pyrimes, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine. 5-methylcytosine substitutions have been shown to increase nucleic acid duplex stability by 0.6-1.2.degree. C. (Sanghvi et al, Antisense Research and Applications, CRC Press, Boca Raton, 1993, pp. 276-278) and are preferred base substitutions, even more particularly when combined with 2'-O-methoxyethyl sugar modifications. Representative United States patents

that teach the preparation of modified nucleobases include, but are not limited to: U.S. Pat. No. 3,687,808, as well as U.S. Pat. Nos.: 4,845,205; 5,130,302; 5,134,066; 5,175,273; 5,367,066; 5,432,272; 5,457,187; 5,459,255; 5,484,908; 5,502,177; 5,525,711; 5,552,540; 5,587,469; 5,594,121, 5,596,091; 5,614,617; 5,645,985; 5,830,653; 5,763,588; 6,005,096; 5,681,941 and 5,750,692, each of which is herein incorporated by reference.

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Another modification of antisense oligonucleotides chemically linking to the oligonucleotide one or more mojeties or conjugates which enhance the activity, cellular distribution or cellular uptake of the oligonucleotide. The compounds of the invention can include conjugate groups covalently bound to functional groups such as primary or secondary hydroxyl groups. Conjugate groups of the invention include intercalators, reporter molecules, polyamines, polyamides, polyethylene glycols, polyethers, groups that enhance the pharmacodynamic properties of oligomers, and groups that enhance the pharmacokinetic properties of oligomers. Typical conjugates groups include cholesterols, lipids, cation lipids, phospholipids, cationic phospholipids, biotin, phenazine, folate, phenanthridine, anthraquinone, acridine, fluoresceins, rhodamines, coumarins, and dyes. Groups that enhance the pharmacodynamic properties, in the context of this invention, include groups that improve oligomer uptake, enhance oligomer resistance to degradation, and/or strengthen sequence-specific hybridization with RNA. Groups that enhance the pharmacokinetic properties, in the context of this invention, include groups that improve oligomer uptake, distribution, metabolism or excretion. Conjugate moieties include but are not limited to lipid moieties such as a cholesterol moiety (Letsinger et al., Proc. Natl. Acad. Sci. USA, 1989, 86, 6553-6556), cholic acid (Manoharan et al., Bioorg. Med. Chem. Let., 1994, 4, 1053-1060), a thioether, e.g., hexyl-S-tritylthiol (Manoharan et al., Ann. N.Y. Acad. Sci., 1992, 660, 306-309; Manoharan et al., Bioorg. Med. Chem. Let., 1993, 3, 2765-2770), a thiocholesterol (Oberhauser et al., Nucl. Acids Res., 1992, 20, 533-538), an aliphatic chain, e.g., dodecandiol or undecyl residues (Saison-Behmoaras et al., EMBO J., 1991, 10, 1111-1118; Kabanov et al., FEBS Lett., 1990, 259, 327-330; Svinarchuk et al., Biochimie, 1993, 75, 49-54), a phospholipid, e.g., di-hexadecyl-rac-glycerol or triethyl-ammonium 1,2-di-O-hexadecyl-rac-glycero-3-H-phosphonate (Manoharan et al., Tetrahedron Lett., 1995, 36, 3651-3654; Shea et al., Nucl. Acids Res., 1990, 18, 3777-3783), a polyamine or a polyethylene glycol chain (Manoharan et al., Nucleosides & Nucleotides, 1995, 14, 969-973), or adamantane acetic acid (Manoharan et al., Tetrahedron Lett., 1995, 36, 3651-3654), a palmityl moiety (Mishra et al., Biochim. Biophys. Acta, 1995, 1264, 229-237), or an octadecylamine or hexylamino-carbonyl-oxycholesterol moiety. Oligonucleotides of the invention may also be conjugated to active drug substances, for example, aspirin, warfarin, phenylbutazone, ibuprofen, suprofen, fenbufen, ketoprofen, (S)-(+)-pranoprofen, carprofen, dansylsarcosine, 2.3.5-triiodobenzoic acid, flufenamic acid, folinic acid, a benzothiadiazide, chlorothiazide, a diazepine, indomethicin, a barbiturate, a cephalosporin, a sulfa drug, an antidiabetic, an antibacterial or an antibiotic. Oligonucleotide-drug conjugates and their preparation are described in U.S. patent application Ser. No. 09/334,130 (filed Jun. 15, 1999) and United States patents Nos.: 4,828,979; 4,948,882; 5,218,105; 5,525,465; 5,541,313; 5,545,730; 5,552,538; 5,578,717, 5,580,731; 5,580,731; 5,591,584; 5,109,124; 5,118,802;

5,138,045; 5,414,077; 5,486,603; 5,512,439; 5,578,718; 5,608,046; 4,587,044; 4,605,735; 4,667,025;

4,762,779; 4,789,737; 4,824,941; 4,835,263; 4,876,335; 4,904,582; 4,958,013; 5,082,830; 5,112,963; 5,214,136; 5,082,830; 5,112,963; 5,214,136; 5,082,830; 5,112,963; 5,245,022; 5,254,469; 5,258,506; 5,262,536; 5,272,250; 5,292,873; 5,317,098; 5,371,241, 5,391,723; 5,416,203, 5,451,463; 5,510,475; 5,512,667; 5,514,785; 5,565,552; 5,567,810; 5,574,142; 5,585,481; 5,587,371; 5,595,726; 5,597,696; 5,599,923; 5,599,928 and 5,688,941, each of which is herein incorporated by reference.

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It is not necessary for all positions in a given compound to be uniformly modified, and in fact more than one of the aforementioned modifications may be incorporated in a single compound or even at a single nucleoside within an oligonucleotide. The present invention also includes antisense compounds which are chimeric compounds. "Chimeric" antisense compounds or "chimeras," in the context of this invention, are antisense compounds, particularly oligonucleotides, which contain two or more chemically distinct regions, each made up of at least one monomer unit, i.e., a nucleotide in the case of an oligonucleotide compound. These oligonucleotides typically contain at least one region wherein the oligonucleotide is modified so as to confer upon the oligonucleotide increased resistance to nuclease degradation, increased cellular uptake, and/or increased binding affinity for the target nucleic acid. An additional region of the oligonucleotide may serve as a substrate for enzymes capable of cleaving RNA:DNA or RNA:RNA hybrids. By way of example, RNase H is a cellular endonuclease which cleaves the RNA strand of an RNA:DNA duplex. Activation of RNase H, therefore, results in cleavage of the RNA target, thereby greatly enhancing the efficiency of oligonucleotide inhibition of gene expression. Consequently, comparable results can often be obtained with shorter oligonucleotides when chimeric oligonucleotides are used, compared to phosphorothioate deoxyoligonucleotides hybridizing to the same target region. Chimeric antisense compounds of the invention may be formed as composite structures of two or more oligonucleotides, modified oligonucleotides, oligonucleosides and/or oligonucleotide mimetics as described above. Preferred chimeric antisense oligonucleotides incorporate at least one 2' modified sugar (preferably 2'-O-(CH₂)₂-O-CH₃) at the 3' terminal to confer nuclease resistance and a region with at least 4 contiguous 2'-H sugars to confer RNase H activity. Such compounds have also been referred to in the art as hybrids or gapmers. Preferred gapmers have a region of 2' modified sugars (preferably 2'-O-(CH₂)₂-O-CH₃) at the 3'-terminal and at the 5' terminal separated by at least one region having at least 4 contiguous 2'-H sugars and preferably incorporate phosphorothioate backbone linkages. Representative United States patents that teach the preparation of such hybrid structures include, but are not limited to, U.S. Pat. Nos. 5,013,830; 5,149,797; 5,220,007; 5,256,775; 5,366,878; 5,403,711; 5,491,133; 5,565,350; 5,623,065; 5,652,355; 5,652,356; and 5,700,922,

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The antisense compounds used in accordance with this invention may be conveniently and routinely made through the well-known technique of solid phase synthesis. Equipment for such synthesis is sold by several vendors including, for example, Applied Biosystems (Foster City, Calif.). Any other means for such synthesis known in the art may additionally or alternatively be employed. It is well known to use similar techniques to prepare oligonucleotides such as the phosphorothioates and alkylated derivatives. The compounds of the invention may also be admixed, encapsulated, conjugated or otherwise associated with other molecules, molecule structures or mixtures of compounds, as for example, liposomes, receptor targeted molecules, oral,

each of which is herein incorporated by reference in its entirety.

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rectal, topical or other formulations, for assisting in uptake, distribution and/or absorption. Representative United States patents that teach the preparation of such uptake, distribution and/or absorption assisting formulations include, but are not limited to, U.S. Pat. Nos. 5,108,921; 5,354,844; 5,416,016; 5,459,127; 5,521,291; 5,543,158; 5,547,932; 5,583,020; 5,591,721; 4,426,330; 4,534,899; 5,013,556; 5,108,921; 5,213,804; 5,227,170; 5,264,221; 5,356,633; 5,395,619; 5,416,016; 5,417,978; 5,462,854; 5,512,295; 5,527,528; 5,534,259; 5,543,152; 5,556,948; 5,580,575; and 5,595,756, each of which is herein incorporated by reference.

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Other examples of sense or antisense oligonucleotides include those oligonucleotides which are covalently linked to organic moieties, such as those described in WO 90/10048, and other moieties that increases affinity of the oligonucleotide for a target nucleic acid sequence, such as poly-(L-lysine). Further still, intercalating agents, such as ellipticine, and alkylating agents or metal complexes may be attached to sense or antisense oligonucleotides to modify binding specificities of the antisense or sense oligonucleotide for the target nucleotide sequence.

Antisense or sense oligonucleotides may be introduced into a cell containing the target nucleic acid sequence by any gene transfer method, including, for example, CaPO 4-mediated DNA transfection, electroporation, or by using gene transfer vectors such as Epstein-Barr virus. In a preferred procedure, an antisense or sense oligonucleotide is inserted into a suitable retroviral vector. A cell containing the target nucleic acid sequence is contacted with the recombinant retroviral vector, either *in vivo* or *ex vivo*. Suitable retroviral vectors include, but are not limited to, those derived from the murine retrovirus M-MuLV, N2 (a retrovirus derived from M-MuLV), or the double copy vectors designated DCT5A, DCT5B and DCT5C (see WO 90/13641).

Sense or antisense oligonucleotides also may be introduced into a cell containing the target nucleotide sequence by formation of a conjugate with a ligand binding molecule, as described in WO 91/04753. Suitable ligand binding molecules include, but are not limited to, cell surface receptors, growth factors, other cytokines, or other ligands that bind to cell surface receptors. Preferably, conjugation of the ligand binding molecule does not substantially interfere with the ability of the ligand binding molecule to bind to its corresponding molecule or receptor, or block entry of the sense or antisense oligonucleotide or its conjugated version into the cell.

Alternatively, a sense or an antisense oligonucleotide may be introduced into a cell containing the target nucleic acid sequence by formation of an oligonucleotide-lipid complex, as described in WO 90/10448. The sense or antisense oligonucleotide-lipid complex is preferably dissociated within the cell by an endogenous lipase.

Antisense or sense RNA or DNA molecules are generally at least about 5 nucleotides in length, alternatively at least about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600, 610, 620, 630, 640, 650, 660, 670, 680, 690, 700, 710,

720, 730, 740, 750, 760, 770, 780, 790, 800, 810, 820, 830, 840, 850, 860, 870, 880, 890, 900, 910, 920, 930, 940, 950, 960, 970, 980, 990, or 1000 nucleotides in length, wherein in this context the term "about" means the referenced nucleotide sequence length plus or minus 10% of that referenced length.

The probes may also be employed in PCR techniques to generate a pool of sequences for identification of closely related TAT coding sequences.

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Nucleotide sequences encoding a TAT can also be used to construct hybridization probes for mapping the gene which encodes that TAT and for the genetic analysis of individuals with genetic disorders. The nucleotide sequences provided herein may be mapped to a chromosome and specific regions of a chromosome using known techniques, such as *in situ* hybridization, linkage analysis against known chromosomal markers, and hybridization screening with libraries.

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When the coding sequences for TAT encode a protein which binds to another protein (example, where the TAT is a receptor), the TAT can be used in assays to identify the other proteins or molecules involved in the binding interaction. By such methods, inhibitors of the receptor/ligand binding interaction can be identified. Proteins involved in such binding interactions can also be used to screen for peptide or small molecule inhibitors or agonists of the binding interaction. Also, the receptor TAT can be used to isolate correlative ligand(s). Screening assays can be designed to find lead compounds that mimic the biological activity of a native TAT or a receptor for TAT. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates. Small molecules contemplated include synthetic organic or inorganic compounds. The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays and cell based assays, which are well characterized in the art.

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Nucleic acids which encode TAT or its modified forms can also be used to generate either transgenic animals or "knock out" animals which, in turn, are useful in the development and screening of therapeutically useful reagents. A transgenic animal (e.g., a mouse or rat) is an animal having cells that contain a transgene, which transgene was introduced into the animal or an ancestor of the animal at a prenatal, e.g., an embryonic stage. A transgene is a DNA which is integrated into the genome of a cell from which a transgenic animal develops. In one embodiment, cDNA encoding TAT can be used to clone genomic DNA encoding TAT in accordance with established techniques and the genomic sequences used to generate transgenic animals that contain cells which express DNA encoding TAT. Methods for generating transgenic animals, particularly animals such as mice or rats, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009. Typically, particular cells would be targeted for TAT transgene incorporation with tissue-specific enhancers. Transgenic animals that include a copy of a transgene encoding TAT introduced into the germ line of the animal at an embryonic stage can be used to examine the effect of increased expression of DNA encoding TAT. Such animals can be used as tester animals for reagents thought to confer protection from, for example, pathological conditions associated with its overexpression. In accordance with this facet of the invention, an animal is treated with the reagent and a reduced incidence of the pathological condition, compared to untreated animals bearing the transgene, would indicate a potential

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therapeutic intervention for the pathological condition.

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Alternatively, non-human homologues of TAT can be used to construct a TAT "knock out" animal which has a defective or altered gene encoding TAT as a result of homologous recombination between the endogenous gene encoding TAT and altered genomic DNA encoding TAT introduced into an embryonic stem cell of the animal. For example, cDNA encoding TAT can be used to clone genomic DNA encoding TAT in accordance with established techniques. A portion of the genomic DNA encoding TAT can be deleted or replaced with another gene, such as a gene encoding a selectable marker which can be used to monitor integration. Typically, several kilobases of unaltered flanking DNA (both at the 5' and 3' ends) are included in the vector [see e.g., Thomas and Capecchi, Cell, 51:503 (1987) for a description of homologous recombination vectors]. The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced DNA has homologously recombined with the endogenous DNA are selected [see e.g., Li et al., Cell, 69:915 (1992)]. The selected cells are then injected into a blastocyst of an animal (e.g., a mouse or rat) to form aggregation chimeras [see e.g., Bradley, in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E. J. Robertson, ed. (IRL, Oxford, 1987), pp. 113-152]. A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term to create a "knock out" animal. Progeny harboring the homologously recombined DNA in their germ cells can be identified by standard techniques and used to breed animals in which all cells of the animal contain the homologously recombined DNA. Knockout animals can be characterized for instance, for their ability to defend against certain pathological conditions and for their development of pathological conditions due to absence of the TAT polypeptide.

Nucleic acid encoding the TAT polypeptides may also be used in gene therapy. In gene therapy applications, genes are introduced into cells in order to achieve *in vivo* synthesis of a therapeutically effective genetic product, for example for replacement of a defective gene. "Gene therapy" includes both conventional gene therapy where a lasting effect is achieved by a single treatment, and the administration of gene therapeutic agents, which involves the one time or repeated administration of a therapeutically effective DNA or mRNA. Antisense RNAs and DNAs can be used as therapeutic agents for blocking the expression of certain genes *in vivo*. It has already been shown that short antisense oligonucleotides can be imported into cells where they act as inhibitors, despite their low intracellular concentrations caused by their restricted uptake by the cell membrane. (Zamecnik *et al.*, Proc. Natl. Acad. Sci. USA 83:4143-4146 [1986]). The oligonucleotides can be modified to enhance their uptake, e.g. by substituting their negatively charged phosphodiester groups by uncharged groups.

There are a variety of techniques available for introducing nucleic acids into viable cells. The techniques vary depending upon whether the nucleic acid is transferred into cultured cells in vitro, or in vivo in the cells of the intended host. Techniques suitable for the transfer of nucleic acid into mammalian cells in vitro include the use of liposomes, electroporation, microinjection, cell fusion, DEAE-dextran, the calcium phosphate precipitation method, etc. The currently preferred in vivo gene transfer techniques include transfection with viral (typically retroviral) vectors and viral coat protein-liposome mediated transfection (Dzau

et al., <u>Trends in Biotechnology</u> 11, 205-210 [1993]). In some situations it is desirable to provide the nucleic acid source with an agent that targets the target cells, such as an antibody specific for a cell surface membrane protein or the target cell, a ligand for a receptor on the target cell, etc. Where liposomes are employed, proteins which bind to a cell surface membrane protein associated with endocytosis may be used for targeting and/or to facilitate uptake, e.g. capsid proteins or fragments thereof tropic for a particular cell type, antibodies for proteins which undergo internalization in cycling, proteins that target intracellular localization and enhance intracellular half-life. The technique of receptor-mediated endocytosis is described, for example, by Wu et al., <u>J. Biol. Chem.</u> 262, 4429-4432 (1987); and Wagner et al., <u>Proc. Natl. Acad. Sci. USA</u> 87, 3410-3414 (1990). For review of gene marking and gene therapy protocols see Anderson et al., <u>Science</u> 256, 808-813 (1992).

The nucleic acid molecules encoding the TAT polypeptides or fragments thereof described herein are useful for chromosome identification. In this regard, there exists an ongoing need to identify new chromosome markers, since relatively few chromosome marking reagents, based upon actual sequence data are presently available. Each TAT nucleic acid molecule of the present invention can be used as a chromosome marker.

The TAT polypeptides and nucleic acid molecules of the present invention may also be used diagnostically for tissue typing, wherein the TAT polypeptides of the present invention may be differentially expressed in one tissue as compared to another, preferably in a diseased tissue as compared to a normal tissue of the same tissue type. TAT nucleic acid molecules will find use for generating probes for PCR, Northern analysis, Southern analysis and Western analysis.

This invention encompasses methods of screening compounds to identify those that mimic the TAT polypeptide (agonists) or prevent the effect of the TAT polypeptide (antagonists). Screening assays for antagonist drug candidates are designed to identify compounds that bind or complex with the TAT polypeptides encoded by the genes identified herein, or otherwise interfere with the interaction of the encoded polypeptides with other cellular proteins, including e.g., inhibiting the expression of TAT polypeptide from cells. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates.

The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays, and cell-based assays, which are well characterized in the art.

All assays for antagonists are common in that they call for contacting the drug candidate with a TAT polypeptide encoded by a nucleic acid identified herein under conditions and for a time sufficient to allow these two components to interact.

In binding assays, the interaction is binding and the complex formed can be isolated or detected in the reaction mixture. In a particular embodiment, the TAT polypeptide encoded by the gene identified herein or the drug candidate is immobilized on a solid phase, e.g., on a microtiter plate, by covalent or non-covalent attachments. Non-covalent attachment generally is accomplished by coating the solid surface with a solution of the TAT polypeptide and drying. Alternatively, an immobilized antibody, e.g., a monoclonal antibody, specific for the TAT polypeptide to be immobilized can be used to anchor it to a solid surface. The assay is performed by adding the non-immobilized component, which may be labeled by a detectable label, to the

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immobilized component, e.g., the coated surface containing the anchored component. When the reaction is complete, the non-reacted components are removed, e.g., by washing, and complexes anchored on the solid surface are detected. When the originally non-immobilized component carries a detectable label, the detection of label immobilized on the surface indicates that complexing occurred. Where the originally non-immobilized component does not carry a label, complexing can be detected, for example, by using a labeled antibody specifically binding the immobilized complex.

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If the candidate compound interacts with but does not bind to a particular TAT polypeptide encoded by a gene identified herein, its interaction with that polypeptide can be assayed by methods well known for detecting protein-protein interactions. Such assays include traditional approaches, such as, e.g., cross-linking, co-immunoprecipitation, and co-purification through gradients or chromatographic columns. In addition, protein-protein interactions can be monitored by using a yeast-based genetic system described by Fields and coworkers (Fields and Song, Nature (London), 340:245-246 (1989); Chien et al., Proc. Natl. Acad. Sci. USA, 88:9578-9582 (1991)) as disclosed by Chevray and Nathans, Proc. Natl. Acad. Sci. USA, 89: 5789-5793 (1991). Many transcriptional activators, such as yeast GALA, consist of two physically discrete modular domains, one acting as the DNA-binding domain, the other one functioning as the transcription-activation domain. The yeast expression system described in the foregoing publications (generally referred to as the "twohybrid system") takes advantage of this property, and employs two hybrid proteins, one in which the target protein is fused to the DNA-binding domain of GAL4, and another, in which candidate activating proteins are fused to the activation domain. The expression of a GAL1- lacZ reporter gene under control of a GAL4activated promoter depends on reconstitution of GALA activity via protein-protein interaction. Colonies containing interacting polypeptides are detected with a chromogenic substrate for \beta-galactosidase. A complete kit (MATCHMAKER™) for identifying protein-protein interactions between two specific proteins using the twohybrid technique is commercially available from Clontech. This system can also be extended to map protein domains involved in specific protein interactions as well as to pinpoint amino acid residues that are crucial for these interactions.

Compounds that interfere with the interaction of a gene encoding a TAT polypeptide identified herein and other intra- or extracellular components can be tested as follows: usually a reaction mixture is prepared containing the product of the gene and the intra- or extracellular component under conditions and for a time allowing for the interaction and binding of the two products. To test the ability of a candidate compound to inhibit binding, the reaction is run in the absence and in the presence of the test compound. In addition, a placebo may be added to a third reaction mixture, to serve as positive control. The binding (complex formation) between the test compound and the intra- or extracellular component present in the mixture is monitored as described hereinabove. The formation of a complex in the control reaction(s) but not in the reaction mixture containing the test compound indicates that the test compound interferes with the interaction of the test compound and its reaction partner.

To assay for antagonists, the TAT polypeptide may be added to a cell along with the compound to be screened for a particular activity and the ability of the compound to inhibit the activity of interest in the presence

of the TAT polypeptide indicates that the compound is an antagonist to the TAT polypeptide. Alternatively, antagonists may be detected by combining the TAT polypeptide and a potential antagonist with membrane-bound TAT polypeptide receptors or recombinant receptors under appropriate conditions for a competitive inhibition assay. The TAT polypeptide can be labeled, such as by radioactivity, such that the number of TAT polypeptide molecules bound to the receptor can be used to determine the effectiveness of the potential antagonist. The gene encoding the receptor can be identified by numerous methods known to those of skill in the art, for example, ligand panning and FACS sorting. Coligan et al., Current Protocols in Immun., 1(2): Chapter 5 (1991). Preferably, expression cloning is employed wherein polyadenylated RNA is prepared from a cell responsive to the TAT polypeptide and a cDNA library created from this RNA is divided into pools and used to transfect COS cells or other cells that are not responsive to the TAT polypeptide. Transfected cells that are grown on glass slides are exposed to labeled TAT polypeptide. The TAT polypeptide can be labeled by a variety of means including iodination or inclusion of a recognition site for a site-specific protein kinase. Following fixation and incubation, the slides are subjected to autoradiographic analysis. Positive pools are identified and sub-pools are prepared and re-transfected using an interactive sub-pooling and re-screening process, eventually yielding a single clone that encodes the putative receptor.

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As an alternative approach for receptor identification, labeled TAT polypeptide can be photoaffinity-linked with cell membrane or extract preparations that express the receptor molecule. Cross-linked material is resolved by PAGE and exposed to X-ray film. The labeled complex containing the receptor can be excised, resolved into peptide fragments, and subjected to protein micro-sequencing. The amino acid sequence obtained from micro- sequencing would be used to design a set of degenerate oligonucleotide probes to screen a cDNA library to identify the gene encoding the putative receptor.

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In another assay for antagonists, mammalian cells or a membrane preparation expressing the receptor would be incubated with labeled TAT polypeptide in the presence of the candidate compound. The ability of the compound to enhance or block this interaction could then be measured.

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More specific examples of potential antagonists include an oligonucleotide that binds to the fusions of immunoglobulin with TAT polypeptide, and, in particular, antibodies including, without limitation, poly- and monoclonal antibodies and antibody fragments, single-chain antibodies, anti-idiotypic antibodies, and chimeric or humanized versions of such antibodies or fragments, as well as human antibodies and antibody fragments. Alternatively, a potential antagonist may be a closely related protein, for example, a mutated form of the TAT polypeptide that recognizes the receptor but imparts no effect, thereby competitively inhibiting the action of the TAT polypeptide.

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Another potential TAT polypeptide antagonist is an antisense RNA or DNA construct prepared using antisense technology, where, e.g., an antisense RNA or DNA molecule acts to block directly the translation of mRNA by hybridizing to targeted mRNA and preventing protein translation. Antisense technology can be used to control gene expression through triple-helix formation or antisense DNA or RNA, both of which methods are based on binding of a polynucleotide to DNA or RNA. For example, the 5' coding portion of the polynucleotide sequence, which encodes the mature TAT polypeptides herein, is used to design an antisense

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RNA oligonucleotide of from about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene involved in transcription (triple helix - see Lee et al., Nucl. Acids Res., 6:3073 (1979); Cooney et al., Science, 241: 456 (1988); Dervan et al., Science, 251:1360 (1991)), thereby preventing transcription and the production of the TAT polypeptide. The antisense RNA oligonucleotide hybridizes to the mRNA in vivo and blocks translation of the mRNA molecule into the TAT polypeptide (antisense - Okano, Neurochem., 56:560 (1991); Oligodeoxynucleotides as Antisense Inhibitors of Gene Expression (CRC Press: Boca Raton, FL, 1988). The oligonucleotides described above can also be delivered to cells such that the antisense RNA or DNA may be expressed in vivo to inhibit production of the TAT polypeptide. When antisense DNA is used, oligodeoxyribonucleotides derived from the translation-initiation site, e.g., between about -10 and +10 positions of the target gene nucleotide sequence, are preferred.

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Potential antagonists include small molecules that bind to the active site, the receptor binding site, or growth factor or other relevant binding site of the TAT polypeptide, thereby blocking the normal biological activity of the TAT polypeptide. Examples of small molecules include, but are not limited to, small peptides or peptide-like molecules, preferably soluble peptides, and synthetic non-peptidyl organic or inorganic compounds.

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Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. Ribozymes act by sequence-specific hybridization to the complementary target RNA, followed by endonucleolytic cleavage. Specific ribozyme cleavage sites within a potential RNA target can be identified by known techniques. For further details see, e.g., Ross<u>Current Biology</u>, 4:469-471 (1994), and PCT publication No. WO 97/33551 (published September 18, 1997).

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Nucleic acid molecules in triple-helix formation used to inhibit transcription should be single-stranded and composed of deoxynucleotides. The base composition of these oligonucleotides is designed such that it promotes triple-helix formation via Hoogsteen base-pairing rules, which generally require sizeable stretches of purines or pyrimidines on one strand of a duplex. For further details see, e.g., PCT publication No. WO 97/33551, supra.

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These small molecules can be identified by any one or more of the screening assays discussed hereinabove and/or by any other screening techniques well known for those skilled in the art.

Isolated TAT polypeptide-encoding nucleic acid can be used herein for recombinantly producing TAT polypeptide using techniques well known in the art and as described herein. In turn, the produced TAT polypeptides can be employed for generating anti-TAT antibodies using techniques well known in the art and as described herein.

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Antibodies specifically binding a TAT polypeptide identified herein, as well as other molecules identified by the screening assays disclosed hereinbefore, can be administered for the treatment of various disorders, including cancer, in the form of pharmaceutical compositions.

If the TAT polypeptide is intracellular and whole antibodies are used as inhibitors, internalizing antibodies are preferred. However, lipofections or liposomes can also be used to deliver the antibody, or an antibody fragment, into cells. Where antibody fragments are used, the smallest inhibitory fragment that specifically binds to the binding domain of the target protein is preferred. For example, based upon the variable-region sequences of an antibody, peptide molecules can be designed that retain the ability to bind the target protein sequence. Such peptides can be synthesized chemically and/or produced by recombinant DNA technology. See, e.g., Marasco et al., Proc. Natl. Acad. Sci. USA, 90: 7889-7893 (1993).

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The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. Alternatively, or in addition, the composition may comprise an agent that enhances its function, such as, for example, a cytotoxic agent, cytokine, chemotherapeutic agent, or growth-inhibitory agent. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The following examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way.

All patent and literature references cited in the present specification are hereby incorporated by reference in their entirety.

EXAMPLES

Commercially available reagents referred to in the examples were used according to manufacturer's instructions unless otherwise indicated. The source of those cells identified in the following examples, and throughout the specification, by ATCC accession numbers is the American Type Culture Collection, Manassas, VA.

EXAMPLE 1: Analysis of Differential TAT Polypeptide Expression by GEPIS

An expressed sequence tag (EST) DNA database (LIFESEQ®, Incyte Pharmaceuticals, Palo Alto, CA) was searched and interesting EST sequences were identified by GEPIS. Gene expression profiling in silico (GEPIS) is a bioinformatics tool developed at Genentech, Inc. that characterizes genes of interest for new cancer therapeutic targets. GEPIS takes advantage of large amounts of EST sequence and library information to determine gene expression profiles. GEPIS is capable of determining the expression profile of a gene based upon its proportional correlation with the number of its occurrences in EST databases, and it works by integrating the LIFESEQ® EST relational database and Genentech proprietary information in a stringent and statistically meaningful way. In this example, GEPIS is used to identify and cross-validate novel tumor antigens, although GEPIS can be configured to perform either very specific analyses or broad screening tasks. For the initial screen, GEPIS is used to identify EST sequences from the LIFESEQ® database that correlate

to expression in a particular tissue or tissues of interest (often a tumor tissue of interest). Then, GEPIS was employed to generate a complete tissue expression profile for the various sequences of interest. Using this type of screening bioinformatics, various TAT polypeptides (and their encoding nucleic acid molecules) were identified as being significantly overexpressed in a particular type of cancer or certain cancers as compared to other cancers and/or normal non-cancerous tissues. The rating of GEPIS hits is based upon several criteria including, for example, tissue specificity, tumor specificity and expression level in normal essential and/or normal proliferating tissues. The following is a list of molecules whose tissue expression profile as determined by GEPIS evidences significant upregulation of expression in a specific tumor or tumors as compared to other tumor(s) and/or normal tissues and optionally relatively low expression in normal essential and/or normal proliferating tissues.

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Under each tissue heading shown below is a list of the cDNA sequences that are detectably overexpressed in tumor tissue of the indicated tissue type as compared to normal non-tumor tissue of the same tissue type. As such, the molecules listed below (and the polypeptides they encode) are excellent nucleic acid (and polypeptide) targets for the diagnosis and therapy of cancer in mammals.

15	PERIPHERAL NERVOUS SYSTEM							
	DNA324303	DNA324573	DNA324681	DNA325296	DNA325405	DNA325407		
	DNA325408	DNA325409	DNA325410	DNA325449	DNA325503	DNA326083		
	DNA326231	DNA188229	DNA327080	DNA327081	DNA327082			
20	BRAIN							
	DNA323721	DNA323722	DNA323723	DNA323724	DNA323726	DNA323727		
	DNA323728	DNA323729	DNA323731	DNA323732	DNA287173	DNA151148		
	DNA323740	DNA323742	DNA323743	DNA323744	DNA323751	DNA323753		
	DNA323755	DNA323757	DNA323759	DNA323764	DNA323765	DNA323778		
25	DNA323781	DNA323783	DNA323785	DNA323795	DNA323796	DNA323797		
	DNA323805	DNA323810	DNA323811	DNA323812	DNA323814	DNA83085		
	DNA323817	DNA323821	DNA273060	DNA323823	DNA323824	DNA256503		
	DNA323825	DNA323826	DNA323828	DNA323829	DNA323830	DNA323833		
	DNA103214	DNA323834	DNA323837	DNA323838	DNA323839	DNA323846		
30	DNA323856	DNA323859	DNA323863	DNA323869	DNA323871	DNA323874		
	DNA323882	DNA323887	DNA323888	DNA323892	DNA323893	DNA323897		
	DNA323898	DNA323900	DNA323901	DNA323902	DNA323908	DNA210134		
	DNA323912	DNA323918	DNA323921	DNA323922	DNA323923	DNA323924		
	DNA323925	DNA323926	DNA257916	DNA323927	DNA323931	DNA323936		
35	DNA323937	DNA323938	DNA323939	DNA323940	DNA323942	DNA226793		
	DNA294794	DNA323943	DNA323944	DNA323946	DNA323947	DNA323950		

	DNA323951	DNA103436	DNA323953	DNA323958	DNA323959	DNA323961
	DNA226619	DNA323962	DNA323964	DNA323969	DNA323970	DNA323973
	DNA323974	DNA323975	DNA323976	DNA323977	DNA323979	DNA323980
	DNA323991	DNA323992	DNA323994	DNA323995	DNA324000	DNA324001
	DNA324002	DNA324003	DNA227246	DNA324004	DNA324008	DNA324009
5	DNA324010	DNA324011	DNA324012	DNA196344	DNA193882	DNA324024
	DNA324034	DNA324037	DNA324042	DNA324046	DNA324047	DNA324048
	DNA324050	DNA324051	DNA324055	DNA275195	DNA324059	DNA324060
	DNA275049	DNA324063	DNA324065	DNA324066	DNA324067	DNA324071
	DNA324072	DNA324073	DNA227165	DNA324074	DNA324076	DNA324077
10	DNA324078	DNA324079	DNA324080	DNA271243	DNA324081	DNA324082
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	DNA324101	DNA324106	DNA324109	DNA324111	DNA324112	DNA324121
	DNA324122	DNA324123	DNA324128	DNA324129	DNA227795	DNA324130
	DNA324131	DNA324132	DNA324133	DNA227528	DNA324134	DNA150725
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	DNA194740	DNA324166	DNA324175	DNA324176	DNA272127	DNA324177
	DNA324182	DNA324184	DNA324186	DNA324188	DNA324194	DNA324197
	DNA324198	DNA324203	DNA324204	DNA324207	DNA324209	DNA324210
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	DNA324224	DNA324227	DNA324228	DNA194827	DNA324230	DNA324231
	DNA324233	DNA324234	DNA324235	DNA324237	DNA324239	DNA254204
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	DNA150884	DNA324256	DNA324258	DNA324260	DNA324262	DNA324264
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	DNA83020	DNA324626	DNA103380	DNA226872	DNA324632	DNA324640
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	DNA327025	DNA327060	DNA327062	DNA327067	DNA327114	

EXAMPLE 2: Use of TAT as a hybridization probe

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The following method describes use of a nucleotide sequence encoding TAT as a hybridization probe for, i.e., diagnosis of the presence of a tumor in a mammal.

DNA comprising the coding sequence of full-length or mature TAT as disclosed herein can also be employed as a probe to screen for homologous DNAs (such as those encoding naturally-occurring variants of TAT) in human tissue cDNA libraries or human tissue genomic libraries.

Hybridization and washing of filters containing either library DNAs is performed under the following high stringency conditions. Hybridization of radiolabeled TAT-derived probe to the filters is performed in a solution of 50% formamide, 5x SSC, 0.1% SDS, 0.1% sodium pyrophosphate, 50 mM sodium phosphate, pH 6.8, 2x Denhardt's solution, and 10% dextran sulfate at 42°C for 20 hours. Washing of the filters is performed in an aqueous solution of 0.1x SSC and 0.1% SDS at 42°C.

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DNAs having a desired sequence identity with the DNA encoding full-length native sequence TAT can then be identified using standard techniques known in the art.

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EXAMPLE 3: Expression of TAT in E. coli

This example illustrates preparation of an unglycosylated form of TAT by recombinant expression in E. coli.

The DNA sequence encoding TAT is initially amplified using selected PCR primers. The primers should contain restriction enzyme sites which correspond to the restriction enzyme sites on the selected expression vector. A variety of expression vectors may be employed. An example of a suitable vector is pBR322 (derived from *E. coli*; see Bolivar et al., Gene, 2:95 (1977)) which contains genes for amplicillin and tetracycline resistance. The vector is digested with restriction enzyme and dephosphorylated. The PCR amplified sequences are then ligated into the vector. The vector will preferably include sequences which encode for an antibiotic resistance gene, a trp promoter, a polyhis leader (including the first six STII codons, polyhis sequence, and enterokinase cleavage site), the TAT coding region, lambda transcriptional terminator, and an argU gene.

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The ligation mixture is then used to transform a selected *E. coli* strain using the methods described in Sambrook et al., <u>supra</u>. Transformants are identified by their ability to grow on LB plates and antibiotic resistant colonies are then selected. Plasmid DNA can be isolated and confirmed by restriction analysis and DNA sequencing.

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Selected clones can be grown overnight in liquid culture medium such as LB broth supplemented with antibiotics. The overnight culture may subsequently be used to inoculate a larger scale culture. The cells are then grown to a desired optical density, during which the expression promoter is turned on.

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After culturing the cells for several more hours, the cells can be harvested by centrifugation. The cell pellet obtained by the centrifugation can be solubilized using various agents known in the art, and the solubilized TAT protein can then be purified using a metal chelating column under conditions that allow tight binding of the protein.

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TAT may be expressed in *E. coli* in a poly-His tagged form, using the following procedure. The DNA encoding TAT is initially amplified using selected PCR primers. The primers will contain restriction enzyme sites which correspond to the restriction enzyme sites on the selected expression vector, and other useful sequences providing for efficient and reliable translation initiation, rapid purification on a metal chelation column, and proteolytic removal with enterokinase. The PCR-amplified, poly-His tagged sequences are then ligated into an expression vector, which is used to transform an *E. coli* host based on strain 52 (W3110)

fuhA(tonA) lon galE rpoHts(htpRts) clpP(lacIq). Transformants are first grown in LB containing 50 mg/ml carbenicillin at 30°C with shaking until an O.D.600 of 3-5 is reached. Cultures are then diluted 50-100 fold into CRAP media (prepared by mixing 3.57 g (NH₄)₂SO₄, 0.71 g sodium citrate•2H2O, 1.07 g KCl, 5.36 g Difco yeast extract, 5.36 g Sheffield hycase SF in 500 mL water, as well as 110 mM MPOS, pH 7.3, 0.55% (w/v) glucose and 7 mM MgSO₄) and grown for approximately 20-30 hours at 30°C with shaking. Samples are removed to verify expression by SDS-PAGE analysis, and the bulk culture is centrifuged to pellet the cells. Cell pellets are frozen until purification and refolding.

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E. coli paste from 0.5 to 1 L fermentations (6-10 g pellets) is resuspended in 10 volumes (w/v) in 7 M guanidine, 20 mM Tris, pH 8 buffer. Solid sodium sulfite and sodium tetrathionate is added to make final concentrations of 0.1M and 0.02 M, respectively, and the solution is stirred overnight at 4°C. This step results in a denatured protein with all cysteine residues blocked by sulfitolization. The solution is centrifuged at 40,000 rpm in a Beckman Ultracentifuge for 30 min. The supernatant is diluted with 3-5 volumes of metal chelate column buffer (6 M guanidine, 20 mM Tris, pH 7.4) and filtered through 0.22 micron filters to clarify. The clarified extract is loaded onto a 5 ml Qiagen Ni-NTA metal chelate column equilibrated in the metal chelate column buffer. The column is washed with additional buffer containing 50 mM imidazole (Calbiochem, Utrol grade), pH 7.4. The protein is eluted with buffer containing 250 mM imidazole. Fractions containing the desired protein are pooled and stored at 4°C. Protein concentration is estimated by its absorbance at 280 nm using the calculated extinction coefficient based on its amino acid sequence.

The proteins are refolded by diluting the sample slowly into freshly prepared refolding buffer consisting of: 20 mM Tris, pH 8.6, 0.3 M NaCl, 2.5 M urea, 5 mM cysteine, 20 mM glycine and 1 mM EDTA. Refolding volumes are chosen so that the final protein concentration is between 50 to 100 micrograms/ml. The refolding solution is stirred gently at 4°C for 12-36 hours. The refolding reaction is quenched by the addition of TFA to a final concentration of 0.4% (pH of approximately 3). Before further purification of the protein, the solution is filtered through a 0.22 micron filter and acetonitrile is added to 2-10% final concentration. The refolded protein is chromatographed on a Poros R1/H reversed phase column using a mobile buffer of 0.1% TFA with elution with a gradient of acetonitrile from 10 to 80%. Aliquots of fractions with A280 absorbance are analyzed on SDS polyacrylamide gels and fractions containing homogeneous refolded protein are pooled. Generally, the properly refolded species of most proteins are eluted at the lowest concentrations of acetonitrile since those species are the most compact with their hydrophobic interiors shielded from interaction with the reversed phase resin. Aggregated species are usually eluted at higher acetonitrile concentrations. In addition to resolving misfolded forms of proteins from the desired form, the reversed phase step also removes endotoxin from the samples.

Fractions containing the desired folded TAT polypeptide are pooled and the acetonitrile removed using a gentle stream of nitrogen directed at the solution. Proteins are formulated into 20 mM Hepes, pH 6.8 with 0.14 M sodium chloride and 4% mannitol by dialysis or by gel filtration using G25 Superfine (Pharmacia) resins equilibrated in the formulation buffer and sterile filtered.

Certain of the TAT polypeptides disclosed herein have been successfully expressed and purified using

this technique(s).

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EXAMPLE 4: Expression of TAT in mammalian cells

This example illustrates preparation of a potentially glycosylated form of TAT by recombinant expression in mammalian cells.

The vector, pRK5 (see EP 307,247, published March 15, 1989), is employed as the expression vector. Optionally, the TAT DNA is ligated into pRK5 with selected restriction enzymes to allow insertion of the TAT DNA using ligation methods such as described in Sambrook et al., <u>supra</u>. The resulting vector is called pRK5-TAT.

In one embodiment, the selected host cells may be 293 cells. Human 293 cells (ATCC CCL 1573) are grown to confluence in tissue culture plates in medium such as DMEM supplemented with fetal calf serum and optionally, nutrient components and/or antibiotics. About 10 µg pRK5-TAT DNA is mixed with about 1 µg DNA encoding the VA RNA gene [Thimmappaya et al., Cell, 31:543 (1982)] and dissolved in 500 µl of 1 mM Tris-HCl, 0.1 mM EDTA, 0.227 M CaCl₂. To this mixture is added, dropwise, 500µl of 50 mM HEPES (pH 7.35), 280 mM NaCl, 1.5 mM NaPO ₄, and a precipitate is allowed to form for 10 minutes at 25°C. The precipitate is suspended and added to the 293 cells and allowed to settle for about four hours at 37°C. The culture medium is aspirated off and 2 ml of 20% glycerol in PBS is added for 30 seconds. The 293 cells are then washed with serum free medium, fresh medium is added and the cells are incubated for about 5 days.

Approximately 24 hours after the transfections, the culture medium is removed and replaced with culture medium (alone) or culture medium containing 200 µCi/ml ³⁵S-cysteine and 200 µCi/ml ³⁵S-methionine. After a 12 hour incubation, the conditioned medium is collected, concentrated on a spin filter, and loaded onto a 15% SDS gel. The processed gel may be dried and exposed to film for a selected period of time to reveal the presence of TAT polypeptide. The cultures containing transfected cells may undergo further incubation (in serum free medium) and the medium is tested in selected bioassays.

In an alternative technique, TAT may be introduced into 293 cells transiently using the dextran sulfate method described by Somparyrac et al., <u>Proc. Natl. Acad. Sci.</u>, <u>12</u>:7575 (1981). 293 cells are grown to maximal density in a spinner flask and 700 μ g pRK5-TAT DNA is added. The cells are first concentrated from the spinner flask by centrifugation and washed with PBS. The DNA-dextran precipitate is incubated on the cell pellet for four hours. The cells are treated with 20% glycerol for 90 seconds, washed with tissue culture medium, and re-introduced into the spinner flask containing tissue culture medium, 5 μ g/ml bovine insulin and 0.1 μ g/ml bovine transferrin. After about four days, the conditioned media is centrifuged and filtered to remove cells and debris. The sample containing expressed TAT can then be concentrated and purified by any selected method, such as dialysis and/or column chromatography.

In another embodiment, TAT can be expressed in CHO cells. The pRK5-TAT can be transfected into CHO cells using known reagents such as CaPO₄ or DEAE-dextran. As described above, the cell cultures can be incubated, and the medium replaced with culture medium (alone) or medium containing a radiolabel such as ³⁵S-methionine. After determining the presence of TAT polypeptide, the culture medium may be replaced

with serum free medium. Preferably, the cultures are incubated for about 6 days, and then the conditioned medium is harvested. The medium containing the expressed TAT can then be concentrated and purified by any selected method.

Epitope-tagged TAT may also be expressed in host CHO cells. The TAT may be subcloned out of the pRK5 vector. The subclone insert can undergo PCR to fuse in frame with a selected epitope tag such as a polyhis tag into a Baculovirus expression vector. The poly-his tagged TAT insert can then be subcloned into a SV40 driven vector containing a selection marker such as DHFR for selection of stable clones. Finally, the CHO cells can be transfected (as described above) with the SV40 driven vector. Labeling may be performed, as described above, to verify expression. The culture medium containing the expressed poly-His tagged TAT can then be concentrated and purified by any selected method, such as by Ni²⁺-chelate affinity chromatography.

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TAT may also be expressed in CHO and/or COS cells by a transient expression procedure or in CHO cells by another stable expression procedure.

Stable expression in CHO cells is performed using the following procedure. The proteins are expressed as an IgG construct (immunoadhesin), in which the coding sequences for the soluble forms (e.g. extracellular domains) of the respective proteins are fused to an IgG1 constant region sequence containing the hinge, CH2 and CH2 domains and/or is a poly-His tagged form.

Following PCR amplification, the respective DNAs are subcloned in a CHO expression vector using standard techniques as described in Ausubel et al., <u>Current Protocols of Molecular Biology</u>, Unit 3.16, John Wiley and Sons (1997). CHO expression vectors are constructed to have compatible restriction sites 5' and 3' of the DNA of interest to allow the convenient shuttling of cDNA's. The vector used expression in CHO cells is as described in Lucas et al., <u>Nucl. Acids Res.</u> 24:9 (1774-1779 (1996), and uses the SV40 early promoter/enhancer to drive expression of the cDNA of interest and dihydrofolate reductase (DHFR). DHFR expression permits selection for stable maintenance of the plasmid following transfection.

Twelve micrograms of the desired plasmid DNA is introduced into approximately 10 million CHO cells using commercially available transfection reagents Superfect[®] (Quiagen), Dosper[®] or Fugene[®] (Boehringer Mannheim). The cells are grown as described in Lucas et al., <u>supra</u>. Approximately 3 x 10⁷ cells are frozen in an ampule for further growth and production as described below.

The ampules containing the plasmid DNA are thawed by placement into water bath and mixed by vortexing. The contents are pipetted into a centrifuge tube containing 10 mLs of media and centrifuged at 1000 rpm for 5 minutes. The supernatant is aspirated and the cells are resuspended in 10 mL of selective media (0.2 μ m filtered PS20 with 5% 0.2 μ m diafiltered fetal bovine serum). The cells are then aliquoted into a 100 mL spinner containing 90 mL of selective media. After 1-2 days, the cells are transferred into a 250 mL spinner filled with 150 mL selective growth medium and incubated at 37°C. After another 2-3 days, 250 mL, 500 mL and 2000 mL spinners are seeded with 3 x 10⁵ cells/mL. The cell media is exchanged with fresh media by centrifugation and resuspension in production medium. Although any suitable CHO media may be employed, a production medium described in U.S. Patent No. 5,122,469, issued June 16, 1992 may actually be used. A 3L production spinner is seeded at 1.2 x 10⁶ cells/mL. On day 0, the cell number pH ie determined. On day

1, the spinner is sampled and sparging with filtered air is commenced. On day 2, the spinner is sampled, the temperature shifted to 33°C, and 30 mL of 500 g/L glucose and 0.6 mL of 10% antifoam (e.g., 35% polydimethylsiloxane emulsion, Dow Corning 365 Medical Grade Emulsion) taken. Throughout the production, the pH is adjusted as necessary to keep it at around 7.2. After 10 days, or until the viability dropped below 70%, the cell culture is harvested by centrifugation and filtering through a 0.22 μ m filter. The filtrate was either stored at 4°C or immediately loaded onto columns for purification.

For the poly-His tagged constructs, the proteins are purified using a Ni-NTA column (Qiagen). Before purification, imidazole is added to the conditioned media to a concentration of 5 mM. The conditioned media is pumped onto a 6 ml Ni-NTA column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 4°C. After loading, the column is washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein is subsequently desalted into a storage buffer containing 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc-containing) constructs are purified from the conditioned media as follows. The conditioned medium is pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column is washed extensively with equilibration buffer before elution with 100 mM citric acid, pH 3.5. The eluted protein is immediately neutralized by collecting 1 ml fractions into tubes containing 275 μ L of 1 M Tris buffer, pH 9. The highly purified protein is subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity is assessed by SDS polyacrylamide gels and by N-terminal amino acid sequencing by Edman degradation.

Certain of the TAT polypeptides disclosed herein have been successfully expressed and purified using this technique(s).

EXAMPLE 5: Expression of TAT in Yeast

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The following method describes recombinant expression of TAT in yeast.

First, yeast expression vectors are constructed for intracellular production or secretion of TAT from the ADH2/GAPDH promoter. DNA encoding TAT and the promoter is inserted into suitable restriction enzyme sites in the selected plasmid to direct intracellular expression of TAT. For secretion, DNA encoding TAT can be cloned into the selected plasmid, together with DNA encoding the ADH2/GAPDH promoter, a native TAT signal peptide or other mammalian signal peptide, or, for example, a yeast alpha-factor or invertase secretory signal/leader sequence, and linker sequences (if needed) for expression of TAT.

Yeast cells, such as yeast strain AB110, can then be transformed with the expression plasmids described above and cultured in selected fermentation media. The transformed yeast supernatants can be analyzed by precipitation with 10% trichloroacetic acid and separation by SDS-PAGE, followed by staining of the gels with Coomassie Blue stain.

Recombinant TAT can subsequently be isolated and purified by removing the yeast cells from the fermentation medium by centrifugation and then concentrating the medium using selected cartridge filters. The

concentrate containing TAT may further be purified using selected column chromatography resins.

Certain of the TAT polypeptides disclosed herein have been successfully expressed and purified using this technique(s).

EXAMPLE 6: Expression of TAT in Baculovirus-Infected Insect Cells

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The following method describes recombinant expression of TAT in Baculovirus-infected insect cells. The sequence coding for TAT is fused upstream of an epitope tag contained within a baculovirus expression vector. Such epitope tags include poly-his tags and immunoglobulin tags (like Fc regions of IgG). A variety of plasmids may be employed, including plasmids derived from commercially available plasmids such as pVL1393 (Novagen). Briefly, the sequence encoding TAT or the desired portion of the coding sequence of TAT such as the sequence encoding an extracellular domain of a transmembrane protein or the sequence encoding the mature protein if the protein is extracellular is amplified by PCR with primers complementary to the 5' and 3' regions. The 5' primer may incorporate flanking (selected) restriction enzyme sites. The product is then digested with those selected restriction enzymes and subcloned into the expression vector.

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Recombinant baculovirus is generated by co-transfecting the above plasmid and BaculoGold TM virus DNA (Pharmingen) into Spodoptera frugiperda ("Sf9") cells (ATCC CRL 1711) using lipofectin (commercially available from GIBCO-BRL). After 4 - 5 days of incubation at 28°C, the released viruses are harvested and used for further amplifications. Viral infection and protein expression are performed as described by O'Reilley et al., Baculovirus expression vectors: A Laboratory Manual, Oxford: Oxford University Press (1994).

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Expressed poly-his tagged TAT can then be purified, for example, by Ni ²⁺-chelate affinity chromatography as follows. Extracts are prepared from recombinant virus-infected Sf9 cells as described by Rupert et al., Nature, 362:175-179 (1993). Briefly, Sf9 cells are washed, resuspended in sonication buffer (25 mL Hepes, pH 7.9; 12.5 mM MgCl₂; 0.1 mM EDTA; 10% glycerol; 0.1% NP-40; 0.4 M KCl), and sonicated twice for 20 seconds on ice. The sonicates are cleared by centrifugation, and the supernatant is diluted 50-fold in loading buffer (50 mM phosphate, 300 mM NaCl, 10% glycerol, pH 7.8) and filtered through a 0.45 µm filter. A Ni²⁺-NTA agarose column (commercially available from Qiagen) is prepared with a bed volume of 5 mL, washed with 25 mL of water and equilibrated with 25 mL of loading buffer. The filtered cell extract is loaded onto the column at 0.5 mL per minute. The column is washed to baseline A₂₈₀ with loading buffer, at which point fraction collection is started. Next, the column is washed with a secondary wash buffer (50 mM phosphate; 300 mM NaCl, 10% glycerol, pH 6.0), which elutes nonspecifically bound protein. After reaching A₂₈₀ baseline again, the column is developed with a 0 to 500 mM Imidazole gradient in the secondary wash buffer. One mL fractions are collected and analyzed by SDS-PAGE and silver staining or Western blot with Ni²⁺-NTA-conjugated to alkaline phosphatase (Qiagen). Fractions containing the eluted His₀-tagged TAT are pooled and dialyzed against loading buffer.

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Alternatively, purification of the IgG tagged (or Fc tagged) TAT can be performed using known chromatography techniques, including for instance, Protein A or protein G column chromatography.

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Certain of the TAT polypeptides disclosed herein have been successfully expressed and purified using

this technique(s).

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EXAMPLE 7: Preparation of Antibodies that Bind TAT

This example illustrates preparation of monoclonal antibodies which can specifically bind TAT.

Techniques for producing the monoclonal antibodies are known in the art and are described, for instance, in Goding, <u>supra</u>. Immunogens that may be employed include purified TAT, fusion proteins containing TAT, and cells expressing recombinant TAT on the cell surface. Selection of the immunogen can be made by the skilled artisan without undue experimentation.

Mice, such as Balb/c, are immunized with the TAT immunogen emulsified in complete Freund's adjuvant and injected subcutaneously or intraperitoneally in an amount from 1-100 micrograms. Alternatively, the immunogen is emulsified in MPL-TDM adjuvant (Ribi Immunochemical Research, Hamilton, MT) and injected into the animal's hind foot pads. The immunized mice are then boosted 10 to 12 days later with additional immunogen emulsified in the selected adjuvant. Thereafter, for several weeks, the mice may also be boosted with additional immunization injections. Serum samples may be periodically obtained from the mice by retro-orbital bleeding for testing in ELISA assays to detect anti-TAT antibodies.

After a suitable antibody titer has been detected, the animals "positive" for antibodies can be injected with a final intravenous injection of TAT. Three to four days later, the mice are sacrificed and the spleen cells are harvested. The spleen cells are then fused (using 35% polyethylene glycol) to a selected murine myeloma cell line such as P3X63AgU.1, available from ATCC, No. CRL 1597. The fusions generate hybridoma cells which can then be plated in 96 well tissue culture plates containing HAT (hypoxanthine, aminopterin, and thymidine) medium to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.

The hybridoma cells will be screened in an ELISA for reactivity against TAT. Determination of "positive" hybridoma cells secreting the desired monoclonal antibodies against TAT is within the skill in the art.

The positive hybridoma cells can be injected intraperitoneally into syngeneic Balb/c mice to produce ascites containing the anti-TAT monoclonal antibodies. Alternatively, the hybridoma cells can be grown in tissue culture flasks or roller bottles. Purification of the monoclonal antibodies produced in the ascites can be accomplished using ammonium sulfate precipitation, followed by gel exclusion chromatography. Alternatively, affinity chromatography based upon binding of antibody to protein A or protein G can be employed.

EXAMPLE 8: Purification of TAT Polypeptides Using Specific Antibodies

Native or recombinant TAT polypeptides may be purified by a variety of standard techniques in the art of protein purification. For example, pro-TAT polypeptide, mature TAT polypeptide, or pre-TAT polypeptide is purified by immunoaffinity chromatography using antibodies specific for the TAT polypeptide of interest. In general, an immunoaffinity column is constructed by covalently coupling the anti-TAT polypeptide antibody to an activated chromatographic resin.

Polyclonal immunoglobulins are prepared from immune sera either by precipitation with ammonium

sulfate or by purification on immobilized Protein A (Pharmacia LKB Biotechnology, Piscataway, N.J.). Likewise, monoclonal antibodies are prepared from mouse ascites fluid by ammonium sulfate precipitation or chromatography on immobilized Protein A. Partially purified immunoglobulin is covalently attached to a chromatographic resin such as CnBr-activated SEPHAROSETM (Pharmacia LKB Biotechnology). The antibody is coupled to the resin, the resin is blocked, and the derivative resin is washed according to the manufacturer's instructions.

Such an immunoaffinity column is utilized in the purification of TAT polypeptide by preparing a fraction from cells containing TAT polypeptide in a soluble form. This preparation is derived by solubilization of the whole cell or of a subcellular fraction obtained via differential centrifugation by the addition of detergent or by other methods well known in the art. Alternatively, soluble TAT polypeptide containing a signal sequence may be secreted in useful quantity into the medium in which the cells are grown.

A soluble TAT polypeptide-containing preparation is passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of TAT polypeptide (e.g., high ionic strength buffers in the presence of detergent). Then, the column is eluted under conditions that disrupt antibody/TAT polypeptide binding (e.g., a low pH buffer such as approximately pH 2-3, or a high concentration of a chaotrope such as urea or thiocyanate ion), and TAT polypeptide is collected.

EXAMPLE 9: In Vitro Tumor Cell Killing Assay

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Mammalian cells expressing the TAT polypeptide of interest may be obtained using standard expression vector and cloning techniques. Alternatively, many tumor cell lines expressing TAT polypeptides of interest are publicly available, for example, through the ATCC and can be routinely identified using standard ELISA or FACS analysis. Anti-TAT polypeptide monoclonal antibodies (and toxin conjugated derivatives thereof) may then be employed in assays to determine the ability of the antibody to kill TAT polypeptide expressing cells in vitro.

For example, cells expressing the TAT polypeptide of interest are obtained as described above and plated into 96 well dishes. In one analysis, the antibody/toxin conjugate (or naked antibody) is included throughout the cell incubation for a period of 4 days. In a second independent analysis, the cells are incubated for 1 hour with the antibody/toxin conjugate (or naked antibody) and then washed and incubated in the absence of antibody/toxin conjugate for a period of 4 days. Cell viability is then measured using the CellTiter-Glo Luminescent Cell Viability Assay from Promega (Cat# G7571). Untreated cells serve as a negative control.

EXAMPLE 10: In Vivo Tumor Cell Killing Assay

To test the efficacy of conjugated or unconjugated anti-TAT polypeptide monoclonal antibodies, anti-TAT antibody is injected intraperitoneally into nude mice 24 hours prior to receiving tumor promoting cells subcutaneously in the flank. Antibody injections continue twice per week for the remainder of the study. Tumor volume is then measured twice per week.

The foregoing written specification is considered to be sufficient to enable one skilled in the art to

practice the invention. The present invention is not to be limited in scope by the construct deposited, since the deposited embodiment is intended as a single illustration of certain aspects of the invention and any constructs that are functionally equivalent are within the scope of this invention. The deposit of material herein does not constitute an admission that the written description herein contained is inadequate to enable the practice of any aspect of the invention, including the best mode thereof, nor is it to be construed as limiting the scope of the claims to the specific illustrations that it represents. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and fall within the scope of the appended claims.

WHAT IS CLAIMED IS:

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1. Isolated nucleic acid having a nucleotide sequence that has at least 80% nucleic acid sequence identity to:

- (a) a DNA molecule encoding the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) a DNA molecule encoding the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) a DNA molecule encoding an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEO ID NOS:1-6355), with its associated signal peptide;
- (d) a DNA molecule encoding an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
 - (e) the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (f) the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
 - (g) the complement of (a), (b), (c), (d), (e) or (f).
- Isolated nucleic acid having:
- (a) a nucleotide sequence that encodes the amino acid sequence shown in any one of Figures 1-6355 (SEO ID NOS:1-6355);
- (b) a nucleotide sequence that encodes the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) a nucleotide sequence that encodes an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) a nucleotide sequence that encodes an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
 - (e) the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (f) the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
 - (g) the complement of (a), (b), (c), (d), (e) or (f).
 - 3. Isolated nucleic acid that hybridizes to:
- (a) a nucleic acid that encodes the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
 - (b) a nucleic acid that encodes the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
 - (c) a nucleic acid that encodes an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) a nucleic acid that encodes an extracellular domain of the polypeptide shown in any one of Figures
 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;

- (e) the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (f) the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
 - (g) the complement of (a), (b), (c), (d), (e) or (f).

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- 4. The nucleic acid of Claim 3, wherein the hybridization occurs under stringent conditions.
- 5. The nucleic acid of Claim 3 which is at least about 5 nucleotides in length.
- 6. An expression vector comprising the nucleic acid of Claim 1, 2 or 3.
- 7. The expression vector of Claim 6, wherein said nucleic acid is operably linked to control sequences recognized by a host cell transformed with the vector.
 - 8. A host cell comprising the expression vector of Claim 7.
 - 9. The host cell of Claim 8 which is a CHO cell, an E. coli cell or a yeast cell.
- 10. A process for producing, a polypeptide comprising culturing the host cell of Claim 8 under conditions suitable for expression of said polypeptide and recovering said polypeptide from the cell culture.
 - 11. An isolated polypeptide having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
 - 12. An isolated polypeptide having:
 - (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;
 - (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
 - (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- 35 (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).

13. A chimeric polypeptide comprising the polypeptide of Claim 11 or 12 fused to a heterologous polypeptide.

- 14. The chimeric polypeptide of Claim 13, wherein said heterologous polypeptide is an epitope tag sequence or an Fc region of an immunoglobulin.
- 15. An isolated antibody that binds to a polypeptide having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);

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- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
 - 16. An isolated antibody that binds to a polypeptide having:
 - (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;
- (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
 - 17. The antibody of Claim 15 or 16 which is a monoclonal antibody.
- 18. The antibody of Claim 15 or 16 which is an antibody fragment.
 - 19. The antibody of Claim 15 or 16 which is a chimeric or a humanized antibody.
 - 20. The antibody of Claim 15 or 16 which is conjugated to a growth inhibitory agent.
 - 21. The antibody of Claim 15 or 16 which is conjugated to a cytotoxic agent.
- The antibody of Claim 21, wherein the cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.
 - 23. The antibody of Claim 21, wherein the cytotoxic agent is a toxin.

24. The antibody of Claim 23, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.

- 25. The antibody of Claim 23, wherein the toxin is a maytansinoid.
- 26. The antibody of Claim 15 or 16 which is produced in bacteria.
- 27. The antibody of Claim 15 or 16 which is produced in CHO cells.
- 28. The antibody of Claim 15 or 16 which induces death of a cell to which it binds.
- 29. The antibody of Claim 15 or 16 which is detectably labeled.

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- 30. An isolated nucleic acid having a nucleotide sequence that encodes the antibody of Claim 15 or 16.
- 31. An expression vector comprising the nucleic acid of Claim 30 operably linked to control sequences recognized by a host cell transformed with the vector.
 - 32. A host cell comprising the expression vector of Claim 31.
 - 33. The host cell of Claim 32 which is a CHO cell, an E. coli cell or a yeast cell.
 - 34. A process for producing an antibody comprising culturing the host cell of Claim 32 under conditions suitable for expression of said antibody and recovering said antibody from the cell culture.
 - 35. An isolated oligopeptide that binds to a polypeptide having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
 - (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
 - (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
 - (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
 - (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
 - (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
 - 36. An isolated oligopeptide that binds to a polypeptide having:
 - (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
 - (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
 - (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;
 - (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
 - (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355

(SEQ ID NOS:1-6355); or

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(f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).

- 37. The oligopeptide of Claim 35 or 36 which is conjugated to a growth inhibitory agent.
- 38. The oligopeptide of Claim 35 or 36 which is conjugated to a cytotoxic agent.
- 39. The oligopeptide of Claim 38, wherein the cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.
 - 40. The oligopeptide of Claim 38, wherein the cytotoxic agent is a toxin.
- 41. The oligopeptide of Claim 40, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.
 - 42. The oligopeptide of Claim 40, wherein the toxin is a maytansinoid.
 - 43. The oligopeptide of Claim 35 or 36 which induces death of a cell to which it binds.
 - 44. The oligopeptide of Claim 35 or 36 which is detectably labeled.
- 45. A TAT binding organic molecule that binds to a polypeptide having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
 - 46. The organic molecule of Claim 45 that binds to a polypeptide having:
 - (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
 - (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
 - (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;
 - (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
 - (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown

in any one of Figures 1-6355 (SEQ ID NOS:1-6355).

- 47. The organic molecule of Claim 45 or 46 which is conjugated to a growth inhibitory agent.
- 48. The organic molecule of Claim 45 or 46 which is conjugated to a cytotoxic agent.
- 49. The organic molecule of Claim 48, wherein the cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.
 - 50. The organic molecule of Claim 48, wherein the cytotoxic agent is a toxin.
- 51. The organic molecule of Claim 50, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.
 - 52. The organic molecule of Claim 50, wherein the toxin is a maytansinoid.
 - 53. The organic molecule of Claim 45 or 46 which induces death of a cell to which it binds.
- The organic molecule of Claim 45 or 46 which is detectably labeled.
 - 55. A composition of matter comprising:
 - (a) the polypeptide of Claim 11;
 - (b) the polypeptide of Claim 12;
 - (c) the chimeric polypeptide of Claim 13;
- 15 (d) the antibody of Claim 15;

- (e) the antibody of Claim 16;
- (f) the oligopeptide of Claim 35;
- (g) the oligopeptide of Claim 36;
- (h) the TAT binding organic molecule of Claim 45; or
- 20 (i) the TAT binding organic molecule of Claim 46; in combination with a carrier.
 - 56. The composition of matter of Claim 55, wherein said carrier is a pharmaceutically acceptable carrier.
 - 57. An article of manufacture comprising:
 - (a) a container; and
- 25 (b) the composition of matter of Claim 55 contained within said container.
 - 58. The article of manufacture of Claim 57 further comprising a label affixed to said container, or a package insert included with said container, referring to the use of said composition of matter for the therapeutic treatment of or the diagnostic detection of a cancer.
- 59. A method of inhibiting the growth of a cell that expresses a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
 - (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
 - (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-

6355), lacking its associated signal peptide;

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(e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or

(f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), said method comprising contacting said cell with an antibody, oligopeptide or organic molecule that binds to said protein, the binding of said antibody, oligopeptide or organic molecule to said protein thereby causing an inhibition of growth of said cell.

- 60. The method of Claim 59, wherein said antibody is a monoclonal antibody.
- 61. The method of Claim 59, wherein said antibody is an antibody fragment.
- 62. The method of Claim 59, wherein said antibody is a chimeric or a humanized antibody.
- 63. The method of Claim 59, wherein said antibody, oligopeptide or organic molecule is conjugated to a growth inhibitory agent.
 - 64. The method of Claim 59, wherein said antibody, oligopeptide or organic molecule is conjugated to a cytotoxic agent.
- 65. The method of Claim 64, wherein said cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.
 - 66. The method of Claim 64, wherein the cytotoxic agent is a toxin.
- 67. The method of Claim 66, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.
 - 68. The method of Claim 66, wherein the toxin is a maytansinoid.
 - 69. The method of Claim 59, wherein said antibody is produced in bacteria.
 - 70. The method of Claim 59, wherein said antibody is produced in CHO cells.
 - 71. The method of Claim 59, wherein said cell is a cancer cell.
- 72. The method of Claim 71, wherein said cancer cell is further exposed to radiation treatment or a chemotherapeutic agent.
- 73. The method of Claim 71, wherein said cancer cell is selected from the group consisting of a breast cancer cell, a colorectal cancer cell, a lung cancer cell, an ovarian cancer cell, a central nervous system cancer cell, a liver cancer cell, a bladder cancer cell, a pancreatic cancer cell, a cervical cancer cell, a melanoma cell and a leukemia cell.
- 74. The method of Claim 71, wherein said protein is more abundantly expressed by said cancer cell as compared to a normal cell of the same tissue origin.
 - 75. The method of Claim 59 which causes the death of said cell.
 - 76. The method of Claim 59, wherein said protein has:
 - (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
 - (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
 - (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures

1-6355 (SEO ID NOS:1-6355), with its associated signal peptide sequence;

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- (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
- 77. A method of therapeutically treating a mammal having a cancerous tumor comprising cells that express a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), said method comprising administering to said mammal a therapeutically effective amount of an antibody, oligopeptide or organic molecule that binds to said protein, thereby effectively treating said mammal.
 - 78. The method of Claim 77, wherein said antibody is a monoclonal antibody.
 - 79. The method of Claim 77, wherein said antibody is an antibody fragment.
 - 80. The method of Claim 77, wherein said antibody is a chimeric or a humanized antibody.
- 81. The method of Claim 77, wherein said antibody, oligopeptide or organic molecule is conjugated to a growth inhibitory agent.
- 82. The method of Claim 77, wherein said antibody, oligopeptide or organic molecule is conjugated to a cytotoxic agent.
- 83. The method of Claim 82, wherein said cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.
 - 84. The method of Claim 82, wherein the cytotoxic agent is a toxin.
 - 85. The method of Claim 84, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.
 - 86. The method of Claim 84, wherein the toxin is a maytansinoid.
- 35 87. The method of Claim 77, wherein said antibody is produced in bacteria.
 - 88. The method of Claim 77, wherein said antibody is produced in CHO cells.

89. The method of Claim 77, wherein said tumor is further exposed to radiation treatment or a chemotherapeutic agent.

- 90. The method of Claim 77, wherein said tumor is a breast tumor, a colorectal tumor, a lung tumor, an ovarian tumor, a central nervous system tumor, a liver tumor, a bladder tumor, a pancreatic tumor, or a cervical tumor.
- 91. The method of Claim 77, wherein said protein is more abundantly expressed by the cancerous cells of said tumor as compared to a normal cell of the same tissue origin.
 - 92. The method of Claim 77, wherein said protein has:

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- (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;
- (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
- 93. A method of determining the presence of a protein in a sample suspected of containing said protein, wherein said protein has at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), said method comprising exposing said sample to an antibody, oligopeptide or organic molecule that binds to said protein and determining binding of said antibody, oligopeptide or organic molecule to said protein in said sample, wherein binding of the antibody, oligopeptide or organic molecule to said protein is indicative of the presence of said protein in said sample.
- 94. The method of Claim 93, wherein said sample comprises a cell suspected of expressing said protein.

- 95. The method of Claim 94, wherein said cell is a cancer cell.
- 96. The method of Claim 93, wherein said antibody, oligopeptide or organic molecule is detectably labeled.
 - 97. The method of Claim 93, wherein said protein has:

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- (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEO ID NOS:1-6355), with its associated signal peptide sequence;
- (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
- 98. A method of diagnosing the presence of a tumor in a mammal, said method comprising determining the level of expression of a gene encoding a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), in a test sample of tissue cells obtained from said mammal and in a control sample of known normal cells of the same tissue origin, wherein a higher level of expression of said protein in the test sample, as compared to the control sample, is indicative of the presence of tumor in the mammal from which the test sample was obtained.
- 99. The method of Claim 98, wherein the step of determining the level of expression of a gene encoding said protein comprises employing an oligonucleotide in an *in situ* hybridization or RT-PCR analysis.
- 100. The method of Claim 98, wherein the step determining the level of expression of a gene encoding said protein comprises employing an antibody in an immunohistochemistry or Western blot analysis.
 - 101. The method of Claim 98, wherein said protein has:

- (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEO ID NOS:1-6355), with its associated signal peptide sequence;
- (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;

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- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
- 102. A method of diagnosing the presence of a tumor in a mammal, said method comprising contacting a test sample of tissue cells obtained from said mammal with an antibody, oligopeptide or organic molecule that binds to a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), and detecting the formation of a complex between said antibody, oligopeptide or organic molecule and said protein in the test sample, wherein the formation of a complex is indicative of the presence of a tumor in said mammal.
- 103. The method of Claim 102, wherein said antibody, oligopeptide or organic molecule is detectably labeled.
- 104. The method of Claim 102, wherein said test sample of tissue cells is obtained from an individual suspected of having a cancerous tumor.
 - 105. The method of Claim 102, wherein said protein has:
 - (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;

(d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;

- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
 - 106. A method for treating or preventing a cell proliferative disorder associated with increased expression or activity of a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);

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- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), said method comprising administering to a subject in need of such treatment an effective amount of an antagonist of said protein, thereby effectively treating or preventing said cell proliferative disorder.
 - 107. The method of Claim 106, wherein said cell proliferative disorder is cancer.
- 108. The method of Claim 106, wherein said antagonist is an anti-TAT polypeptide antibody, TAT binding oligopeptide, TAT binding organic molecule or antisense oligonucleotide.
- 109. A method of binding an antibody, oligopeptide or organic molecule to a cell that expresses a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- 35 (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), said method comprising contacting said cell with an antibody,

oligopeptide or organic molecule that binds to said protein and allowing the binding of the antibody, oligopeptide or organic molecule to said protein to occur, thereby binding said antibody, oligopeptide or organic molecule to said cell.

- 110. The method of Claim 109, wherein said antibody is a monoclonal antibody.
- 111. The method of Claim 109, wherein said antibody is an antibody fragment.

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- 112. The method of Claim 109, wherein said antibody is a chimeric or a humanized antibody.
- 113. The method of Claim 109, wherein said antibody, oligopeptide or organic molecule is conjugated to a growth inhibitory agent.
- 114. The method of Claim 109, wherein said antibody, oligopeptide or organic molecule is conjugated to a cytotoxic agent.
- 115. The method of Claim 114, wherein said cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.
 - 116. The method of Claim 114, wherein the cytotoxic agent is a toxin.
- 117. The method of Claim 116, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.
 - 118. The method of Claim 116, wherein the toxin is a maytansinoid.
 - 119. The method of Claim 109, wherein said antibody is produced in bacteria.
 - 120. The method of Claim 109, wherein said antibody is produced in CHO cells.
 - 121. The method of Claim 109, wherein said cell is a cancer cell.
- 122. The method of Claim 121, wherein said cancer cell is further exposed to radiation treatment or a chemotherapeutic agent.
- 123. The method of Claim 121, wherein said cancer cell is selected from the group consisting of a breast cancer cell, a colorectal cancer cell, a lung cancer cell, an ovarian cancer cell, a central nervous system cancer cell, a liver cancer cell, a bladder cancer cell, a pancreatic cancer cell, a cervical cancer cell, a melanoma cell and a leukemia cell.
- 124. The method of Claim 123, wherein said protein is more abundantly expressed by said cancer cell as compared to a normal cell of the same tissue origin.
 - 125. The method of Claim 109 which causes the death of said cell.
- 126. Use of a nucleic acid as claimed in any of Claims 1 to 5 or 30 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
- 127. Use of a nucleic acid as claimed in any of Claims 1 to 5 or 30 in the preparation of a medicament for treating a tumor.
- 128. Use of a nucleic acid as claimed in any of Claims 1 to 5 or 30 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.
- 129. Use of an expression vector as claimed in any of Claims 6, 7 or 31 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
 - 130. Use of an expression vector as claimed in any of Claims 6, 7 or 31 in the preparation of

medicament for treating a tumor.

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131. Use of an expression vector as claimed in any of Claims 6, 7 or 31 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.

- 132. Use of a host cell as claimed in any of Claims 8, 9, 32, or 33 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
- 133. Use of a host cell as claimed in any of Claims 8, 9, 32 or 33 in the preparation of a medicament for treating a tumor.
- 134. Use of a host cell as claimed in any of Claims 8, 9, 32 or 33 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.

135. Use of a polypeptide as claimed in any of Claims 11 to 14 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.

- 136. Use of a polypeptide as claimed in any of Claims 11 to 14 in the preparation of a medicament for treating a tumor.
- 137. Use of a polypeptide as claimed in any of Claims 11 to 14 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.

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- 138. Use of an antibody as claimed in any of Claims 15 to 29 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
- 139. Use of an antibody as claimed in any of Claims 15 to 29 in the preparation of a medicament for treating a tumor.
- 140. Use of an antibody as claimed in any of Claims 15 to 29 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.
- 141. Use of an oligopeptide as claimed in any of Claims 35 to 44 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
- 142. Use of an oligopeptide as claimed in any of Claims 35 to 44 in the preparation of a medicament for treating a tumor.
- 143. Use of an oligopeptide as claimed in any of Claims 35 to 44 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.
- 144. Use of a TAT binding organic molecule as claimed in any of Claims 45 to 54 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
- 145. Use of a TAT binding organic molecule as claimed in any of Claims 45 to 54 in the preparation of a medicament for treating a tumor.
- 146. Use of a TAT binding organic molecule as claimed in any of Claims 45 to 54 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.
- 147. Use of a composition of matter as claimed in any of Claims 55 or 56 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
- 148. Use of a composition of matter as claimed in any of Claims 55 or 56 in the preparation of a medicament for treating a tumor.
- 149. Use of a composition of matter as claimed in any of Claims 55 or 56 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.
- 150. Use of an article of manufacture as claimed in any of Claims 57 or 58 in the preparation of a medicament for the therapeutic treatment or diagnostic detection of a cancer.
- 151. Use of an article of manufacture as claimed in any of Claims 57 or 58 in the preparation of a medicament for treating a tumor.

152. Use of an article of manufacture as claimed in any of Claims 57 or 58 in the preparation of a medicament for treatment or prevention of a cell proliferative disorder.

- 153. A method for inhibiting the growth of a cell, wherein the growth of said cell is at least in part dependent upon a growth potentiating effect of a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);

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- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), said method comprising contacting said protein with an antibody, oligopeptide or organic molecule that binds to said protein, there by inhibiting the growth of said cell.
 - 154. The method of Claim 153, wherein said cell is a cancer cell.
 - 155. The method of Claim 153, wherein said protein is expressed by said cell.
- 156. The method of Claim 153, wherein the binding of said antibody, oligopeptide or organic molecule to said protein antagonizes a cell growth-potentiating activity of said protein.
- 157. The method of Claim 153, wherein the binding of said antibody, oligopeptide or organic molecule to said protein induces the death of said cell.
 - 158. The method of Claim 153, wherein said antibody is a monoclonal antibody.
 - 159. The method of Claim 153, wherein said antibody is an antibody fragment.
 - 160. The method of Claim 153, wherein said antibody is a chimeric or a humanized antibody.
- 161. The method of Claim 153, wherein said antibody, oligopeptide or organic molecule is conjugated to a growth inhibitory agent.
 - 162. The method of Claim 153, wherein said antibody, oligopeptide or organic molecule is conjugated to a cytotoxic agent.
- 163. The method of Claim 162, wherein said cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.
 - 164. The method of Claim 162, wherein the cytotoxic agent is a toxin.
 - 165. The method of Claim 164, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.
 - 166. The method of Claim 164, wherein the toxin is a maytansinoid.
- 35 167. The method of Claim 153, wherein said antibody is produced in bacteria.
 - 168. The method of Claim 153, wherein said antibody is produced in CHO cells.

169. The method of Claim 153, wherein said protein has:

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- (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;
- (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEO ID NOS:1-6355), lacking its associated signal peptide sequence;
- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).
- 170. A method of therapeutically treating a tumor in a mammal, wherein the growth of said tumor is at least in part dependent upon a growth potentiating effect of a protein having at least 80% amino acid sequence identity to:
 - (a) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);
- (b) the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (c) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide;
- (d) an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide;
- (e) a polypeptide encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) a polypeptide encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), said method comprising contacting said protein with an antibody, oligopeptide or organic molecule that binds to said protein, thereby effectively treating said tumor.
 - 171. The method of Claim 170, wherein said protein is expressed by cells of said tumor.
- 172. The method of Claim 170, wherein the binding of said antibody, oligopeptide or organic molecule to said protein antagonizes a cell growth-potentiating activity of said protein.
 - 173. The method of Claim 170, wherein said antibody is a monoclonal antibody.
 - 174. The method of Claim 170, wherein said antibody is an antibody fragment.
 - 175. The method of Claim 170, wherein said antibody is a chimeric or a humanized antibody.
- 176. The method of Claim 170, wherein said antibody, oligopeptide or organic molecule is conjugated to a growth inhibitory agent.
- The method of Claim 170, wherein said antibody, oligopeptide or organic molecule is conjugated to a cytotoxic agent.

178. The method of Claim 177, wherein said cytotoxic agent is selected from the group consisting of toxins, antibiotics, radioactive isotopes and nucleolytic enzymes.

- 179. The method of Claim 177, wherein the cytotoxic agent is a toxin.
- 180. The method of Claim 179, wherein the toxin is selected from the group consisting of maytansinoid and calicheamicin.

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- 181. The method of Claim 179, wherein the toxin is a maytansinoid.
- 182. The method of Claim 170, wherein said antibody is produced in bacteria.
- 183. The method of Claim 170, wherein said antibody is produced in CHO cells.
- 184. The method of Claim 170, wherein said protein has:
- (a) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355);

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- (b) the amino acid sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (c) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), with its associated signal peptide sequence;
- (d) an amino acid sequence of an extracellular domain of the polypeptide shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355), lacking its associated signal peptide sequence;
- (e) an amino acid sequence encoded by the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355); or
- (f) an amino acid sequence encoded by the full-length coding region of the nucleotide sequence shown in any one of Figures 1-6355 (SEQ ID NOS:1-6355).

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FIGURE 3

CAAGCTCATGACTCACAATGGCCTATTTAGGCCCATACCCTACGTCACGGCAGCCTCCGCAGATGAGCCTACTGC
CTCACAACAGCCTCCACAGGCACAGCTCCATCGTTACAATGGCCTCTTTAGACCCAGCTCCTGCCTCCCAGCCTT
CTCTCCAGGCTCTGAACTTTCTCAGGTCTCCCTCTGTTGTCCAAGGCTGGAGTGTAGTAGTGCTATCGCAGCTGA
CTGCAGCCTCAACCTTCCAGGCTGAAGCGATCCTCCCACCTCCACCTCCCACGTGGCTGAGACTACAGGTGCTTG
CCACTATGCCCAACTAACATTTGGAATTTTCGTATACGTGGATTCCAGAGGGGTGACAGCGAAACGTGGGACCAT
TCAGTTGCAGGAAAACAAGCTTAACACGCCCACTAATTCTACATTATGCTCCTACCTCCCGGCAGCCTCTCCAGG
CCCAGAACTTTCTCCAGTCAGCCCTCTACAGACCAAGCTCATGACTCACAATG

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CGTCGGCCCCCGGCCCCCAGCAGCCTCCAAAGCCCTGTGACTCACAGCCCTGCTTCCACGGGGGGACCTGCCAG GGCGCCCTGTGCCGGCCTTCGAGGGCCGCTCCTTCCTGGCCTTCCCCACTCTCCGCGCCTACCACACGCTGCGC CTGGCACTGGAATTCCGGGCGCTGGAGCCTCAGGGGCTGCTGCTGCAATGGCAACGCCCGGGGCAAGGACTTC CTGGCATTGGCGCTGCTAGATGGCCGCGTGCAGCTCAGGTTTGACACAGGTTCGGGGCCGGCGGTGCTGACCAGT GCCGTGCCGGTAGAGCCGGGCCAGTGGCACCGCCTGGAGCTGTCCCGGCACTGGCGCCCGGGGCACCCTCTCGGTG GATGGTGAGACCCCTGTTCTGGGCGAGAGTCCCAGTGGCACCGACGGCCTCAACCTGGACACAGACCTCTTTGTG GGCGGCGTACCCGAGGACCAGGCTGCCGTGGCGCTGGAGCGGACCTTCGTGGGCGCCCGGCCTGAGGGGGTGCATC CGTTTGCTGGACGTCAACAACCAGCGCCTGGAGCTTGGCATTGGGCCGGGGGCTGCCACCCGAGGCTCTGGCGTG GGCAAGTGCGGGGACCACCCCTGCCCAACCCCTGCCATGGCGGGGCCCCATGCCAGAACCTGGAGGCTGGA $\verb|CCCTGCCATGGGGCGCCCCTGCCGTGTGCCCGAGGGTGGTGCTCAGTGCGAGTGCCCCTGGGGCGTGAG| \\$ $\tt CTGGAGCTGAGAGGCCTGCACACCTTTGCACGGGACCTGGGGGGAGAAGATGGCGCTGGAGGTCGTGTTCCTGGCA$ $\tt CTGGGAGCCTGGACCAGGGTCTCACTGGAGCGAAACGGCCGCAAGGGTGCCCTGCGTGTGGGCGACGGCCCCCGT$ GTGTTGGGGGGGGTCCCGGCACACCGTCCTCAACCTGAAGGAGCCGCTCTACGTAGGGGGGCGCTCCCGAC TTCAGCAAGCTGGCCCGTGCTGCCGTGTCCTCTGGCTTCGACGGTGCCATCCAGCTGGTCTCCCTCGGAGGC CGCCAGCTGCTGACCCCGGAGCACGTGCTGCGGCAGGTGGACGTCACGTCCTTTGCAGGTCACCCCTGCACCCGG GGATTCTCAGGACCGCACTGCGAGAAGGGGCTGGTGGAGAAGTCAGCGGGGGACGTGGATACCTTGGCCTTTGAC GGGCGGACCTTTGTCGAGTACCTCAACGCTGTGACCGAGAGCGAGAGGCACTGCAGAGCAACCACTTTGAACTG AGCCTGCGCACTGAGGCCACGCAGGGGCTGGTGCTCTGGAGTGGCAAGGCCACGGAGCGGGCAGACTATGTGGCA CTGGCCATTGTGGACGGCACCTGCAACTGAGCTACAACCTGGGCTCCCAGCCCGTGGTGCTGCGTTCCACCGTG GAGGCCCCTGTGACCGGCTCCCCCGCTGGGCGCCACGCAGCTGGACACTGATGGAGCCCTGTGGCTTGGGGGC CTGCCGGAGCTGCCCGTGGGCCCAGCACTGCCCAAGGCCTACGGCACAGGCTTTGTGGGCTGCTTGCGGGACGTG GTGGTGGGCCGGCACCCGCTGCACCTGGAGGAGGACGCCGTCACCAAGCCAGAGCTGCGGCCCTGCCCCACCCCA TGAGCTGGCACCAGAGCCCCGCCGCCCGCTGTAATTATTTTCTATTTTTGTAAACTTGTTGCTTTTTGATATGATT CCTAGTGCCGAGGGATGGACAGGCGAGGTGGCAGCGTGGAGGGCTCGGCGTGGATGGCAGCCTCAGGACACAC CCCTGCCTCAAGGTGCTGAGCCCCCGCCTTGCACTGCGCCCACGGTGTCCCCGCCGGGAAGCAGCCCGG CTCCTGAATCACCCTCGCTCCGTCAGGCGGGACTCGTGTCCCAGAGAGGGAAGGGGCTGCTGAGGTCTGATGGGGC $\tt CTGCCTCGGCCTCCTGCGCCAATACTGTGACTTCCAAACAATGTTACTGCTGGGCACAGCTCTGCGTTGCTCCCG$ TGCTGCCTGCGCCAGCCCCAGGCTGCTGAGGAGCCAGAGCCAGGCCCGATCTGGGTGTCCTGACCCTCAG $\tt CTGGCCCTGCCAGCCACCCTGGACATGACCGTATCCCTCTGCCACACCCCAGGCCCTGCGAGGGGCTATCGAGA$ TGTGTTGATTTTATTTGACCCCTGGAGTGGTGGGTCTCATCTTTCCCATCTCGCCTGAGAGCGGCTGAGGGCTGC GACCAAGGTCAAGGGGCAGGTGCAGAGGTGGCAGGGATGGCTCCGAAGCCAGAAATGCCTTAAACTGCAACGTCC CGTCCCTTCCCCACCCCATCCCATCCCCACCCCAGCCCAGCCCAGTCCTCCTAGGAGCAGGACCCGATGAAG CGGGCGGCGGGGGGGGGGGGGGGGGTGTTACTAACTCTAGTATGTTTCTGTGTCAATCGCTGTGAAATAAAGTCT GAAAACTTT

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MLNSSLMRITLRNLEEVEFCVEDKPGTHFTPVPPTPPDACRGMLCGFGAVCEPNAEGPGRASCVCKKSPCPSVVA PVCGSDASTYSNECELQRAQCSQQRRIRLLSRGPCGSRDPCSNVTCSFGSTCARSADGLTASCLCPATCRGAPEG ${\tt TVCGSDGADYPGECQLLRRACARQENVFKKFDGPCDPCQGALPDPSRSCRVNPRTRRPEMLLRPESCPARQAPVC}$ GDDGVTYENDCVMGRSGAARGLLLQKVRSGQCQGRDQCPEPCRFNAVCLSRRGRPRCSCDRVTCDGAYRPVCAQD GRTYDSDCWRQQAECRQQRAIPSKHQGPCDQAPSPCLGVQCAFGATCAVKNGQAACECLQACSSLYDPVCGSDGV TYGSACELEATACTLGREIQVARKGPCDRCGQCRFGALCEAETGRCVCPSECVALAQPVCGSDGHTYPSECMLHV HACTHQISLHVASAGPCETCGDAVCAFGAVCSAGQCVCPRCEHPPPGPVCGSDGVTYGSACELREAACLQQTQIE EARAGPCEQAECGSGGSGSGEDGDCEQELCRQRGGIWDEDSEDGPCVCDFSCQSVPGSPVCGSDGVTYSTECELK ${\tt KARCESQRGLYVAAQGACRGPTFAPLPPVAPLHCAQTPYGCCQDNITAARGVGLAGCPSACQCNPHGSYGGTCDP}$ $\verb|ATGQCSCRPGVGGLRCDRCEPGFWNFRGIVTDGRSGCTPCSCDPQGAVRDDCEQMTGLCSCKPGVAGPKCGQCPD| \\$ GRALGPAGCEADASAPATCAEMRCEFGARCVEESGSAHCVCPMLTCPEANATKVCGSDGVTYGNECQLKTIACRQ GLQISIQSLGPCQEAVAPSTHPTSASVTVTTPGLLLSQALPAPPGALPLAPSSTAHSQTTPPPSSRPRTTASVPR TTVWPVLTVPPTAPSPAPSLVASAFGESGSTDGSSDEELSGDQEASGGGSGGLEPLEGSSVATPGPPVERASCYN SALGCCSDGKTPSLDAEGSNCPATKVFQGVLELEGVEGQELFYTPEMADPKSELFGETARSIESTLDDLFRNSDV KKDFRSVRLRDLGPGKSVRAIVDVHFDPTTAFRAPDVARALLRQIQVSRRRSLGVRRPLQEHVRFMDFDWFPAFI TGATSGAIAAGATARATTASRLPSSAVTPRAPHPSHTSQPVAKTTAAPTTRRPPTTAPSRVPGRRPPAPQQPPKP CDSQPCFHGGTCQDWALGGGFTCSCPAGRGGAVCEKVLGAPVPAFEGRSFLAFPTLRAYHTLRLALEFRALEPQG LLLYNGNARGKDFLALALLDGRVQLRFDTGSGPAVLTSAVPVEPGQWHRLELSRHWRRGTLSVDGETPVLGESPS ${\tt GTDGLNLDTDLFVGGVPEDQAAVALERTFVGAGLRGCIRLLDVNNQRLELGIGPGAATRGSGVGKCGDHPCLPNP}$ $\verb|CHGGAPCQNLEAGRFHCQCPPGRVGPTCADEKSPCQPNPCHGAAPCRVLPEGGAQCECPLGREGTFCQTASGQDG|\\$ ${\tt SGPFLADFNGFSHLELRGLHTFARDLGEKMALEVVFLARGPSGLLLYNGQKTDGKGDFVSLALRDRRLEFRYDLG}$ KGAAVIRSREPVTLGAWTRVSLERNGRKGALRVGDGPRVLGESPVPHTVLNLKEPLYVGGAPDFSKLARAAAVSS GFDGAIQLVSLGGRQLLTPEHVLRQVDVTSFAGHPCTRASGHPCLNGASCVPREAAYVCLCPGGFSGPHCEKGLV EKSAGDVDTLAFDGRTFVEYLNAVTESEKALQSNHFELSLRTEATQGLVLWSGKATERADYVALAIVDGHLQLSY NLGSQPVVLRSTVPVNTNRWLRVVAHREQREGSLQVGNEAPVTGSSPLGATQLDTDGALWLGGLPELPVGPALPK AYGTGFVGCLRDVVVGRHPLHLLEDAVTKPELRPCPTP

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FIGURE 6

ACAGAGACCCCGAGTTCTACAAGTTCCTGCAGGAGAATGACCAGAGCCTGCTAAACTTCAGCGACTCGGACAGCT ${\tt AGGAAGGAGAAGATGGGGACAGAGTCCCCAGAGGGCTGAAGGGGAAGAAGAATTCTGTTCCTGTGACCGTCGCCA}$ TGGTTGAGAGTGGAAGCAGCCAAAGCAACGCCTCACTCCAAAGCTGTTCCATGAAGTGGTACAGGCGTTCC CATTCAATGCTCTGGGTTACCTTCTGCATCAGAGACCTCATTGGCTGTCTCCAGAAGCTGCTGTTTGGAAAGGTGG ${\tt CAAAGGATAGCAGCAGCAGCCGTCCAGCAGCCGCTCTGGGGGAAGCTTCGTGTGGACATCAAGGCTT}$ ACCTGGGCTCGGCCATACAGCTGGTGTCCTGTCTGTCGGAGACGACGGTGTTGGCGGCCGTGCTGCGGCACATCA GCGTGCTGGTGCCCTGCTTCCTGACCTTCCCCAAGCAGTGCCGCATGCTGCTCAAGAGAATGGTGATCGTATGGA GCACTGGGGAAGAGTCTCTGCGGGTGCTGGCTTTCCTGGTCCTCAGCAGAGTCTGCCGGCACAAGAAGGACACTT TCCTTGGCCCCGTCCTCAAGCAAATGTACATCACGTATGTGAGGAACTGCAAGTTCACCTCGCCTGGTGCCCTCC $\verb|CCTTCATCAGTTCATGCAGTGGACCTTGACGGAGCTGGTGGCCTGGAGCCGGGTGTGGCCTACCAGCACGCCT| \\$ TCCTCTACATCCGCCAGCTCGCCATACACCTGCGCAACGCCATGACCACTCGCAAGAAGGAAACATACCAGTCTG TGTACAACTGGCAGTATGTGCACTGCCTCTTCCTGTGGTGCCGGGTCCTGAGCACTGCGGGCCCCAGCGAAGCCC CGCTGCGAATGCACTCCGTGCCCTGACGCTGCTCTCGGGGGGCCTCGGGGGCCTTCATCCCGGTGCTGCCTT TCATCCTGGAGATGTTCCAGCAGGTCGACTTCAACAGGAAGCCAGGGCGCATGAGCTCCAAGCCCATCAACTTCT CCGTGATCCTGAAGCTGTCCAATGTCAACCTGCAGGAGAAGGCGTACCGGGACGGCCTGGTGGAGCAGCTGTACG ACCTCACCCTGGAGTACCTGCACAGCCAGGCACACTGCATCGGCTTCCCGGAGCTGGTGCTGCCTGTGGTCCTGC AGCTGAAGTCGTTCCTCCGGGAGTGCAAGGTGGCCAACTACTGCCGGCAGGTGCAGCAGCTGCTTGGGAAGGTTC AGGAGAACTCGGCATACATCTGCAGCCGCCAGAGGGTTTCCTTCGGCGTCTCTGAGCAGCAGCAGTGGAAG CCTGGGAGAAGCTGACCCGGGAAGAGGGGACACCCCTGACCTTGTACTACAGCCACTGGCGCAAGCTGCGTGACC TGGCTGACAGGAAGGATGAGGACAGGAAGCAATTTAAAGACCTCTTTGACCTGAACAGCTCTGAAGAGGACGACA CCGAGGGATTCTCGGAGAGAGGGGATACTGAGGCCCCTGAGCACTCGGCATGGGGTGGAAGACGATGAAGAGGACG AGGAGGAGGGCGAGGACAGCAGCAACTCGGAGGATGGAGACCCAGACGCAGAGGCGGGGCTGGCCCCTGGGG AGCTGCAGCAGCTGGCCCAGGGGCCGGAGGACGAGCTGGAGGATCTGCAGCTCTCAGAGGACGACTGAGGCAGCC CATCTGGGGGGCCTGTAGGGCTGCCGGGCTGGTGGCCAGTGTTTCCACCTCCCTGGCAGTCAGGCCTAGAGGCT CGTATCGAGAGCTGGGCTGGGCTGGTGTGGCTGCTGAAGCCCCACAGCTGTGGGCTGCTGAAGTCAGCTC CGCGGGGGAGCTGACCTGACGTCAGCAGACCGAGACCAGTCCCAGTTCCAGGGGGAGGCCTGCAGGCCCCTGGC CCCTTCCACCACCTCTGCCCTCCGTCTGCAGACCTCGTCCATCTGCACCAGGCTCTGCCTTCACTCCCCAAGTC TTTGAAAATTTTGTTCCTTTCCAAAGTCACATTTTCTTTTAAAATTTTTTGTTTTGCATCCGAAACCGAAAGA AATAAAGCGGTGGGAGGCAGGGCCATTGTGTTG

7/6881 **FIGURE 7**

MAAAGSRKRRLAELTVDEFLASGFDSESESESENSPQAETREAREAARSPDKPGGSPSASRRKGRASEHKDQLSR LKDRDPEFYKFLQENDQSLLNFSDSDSSEEEEGPFHSLPDVLEEASEEEDGAEEGEDGDRVPRGLKGKKNSVPVT VAMVERWKQAAKQRLTPKLFHEVVQAFRAAVATTRGDQESAEANKFQVTDSAAFNALVTFCIRDLIGCLQKLLFG KVAKDSSRMLQPSSSPLWGKLRVDIKAYLGSAIQLVSCLSETTVLAAVLRHISVLVPCFLTFPKQCRMLLKRMVI VWSTGEESLRVLAFLVLSRVCRHKKDTFLGPVLKQMYITYVRNCKFTSPGALPFISFMQWTLTELLALEPGVAYQ HAFLYIRQLAIHLRNAMTTRKKETYQSVYNWQYVHCLFLWCRVLSTAGPSEALQPLVYPLAQVIIGCIKLIPTAR FYPLRMHCIRALTLLSGSSGAFIPVLPFILEMFQQVDFNRKPGRMSSKPINFSVILKLSNVNLQEKAYRDGLVEQ LYDLTLEYLHSQAHCIGFPELVLPVVLQLKSFLRECKVANYCRQVQQLLGKVQENSAYICSRRQRVSFGVSEQQA VEAWEKLTREEGTPLTLYYSHWRKLRDREIQLEISGKERLEDLNFPEIKRRKMADRKDEDRKQFKDLFDLNSSEE DDTEGFSERGILRPLSTRHGVEDDEEDEEGEEDSNSEDGDPDAEAGLAPGELQQLAQGPEDELEDLQLSEDD